The range-wide, eDNA-based inventory of bull trout: early results and an ongoing invitation

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Project Coordinators & Presentation Co-Authors

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Collaborators

Sponsors

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Idaho Power Company
Kalispel Tribes
Lewis River Bull Trout Recovery

Team Montana Department of Natural Resources Conservation Montana Fish, Wildlife & Parks

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Nez Perce Tribes
North Cascades National Park
Oregon Department of Fish and
Wildlife
Trout Unlimited

University of Washington
U.S. Fish and Wildlife Service

National Forests:

Beaverhead-Deer Lodge, Boise, Colville, Deschutes, Flathead, Gifford Pinchot, Helena, Idaho

Panhandle, Lolo, Mount Baker-Snoqualmie, Nez Perce-Clearwater, Payette, Salmon-Challis, Sawtooth,

Umatilla, Wallowa-Whitman,

Wenatchee

Regions 1, 4, and 6

Washington Department of Fish and

Wildlife

Whitefish Institute

Wild Fish Conservancy

Yakama Nation





Institutional Support



Outline

- Focal species/life stage
- Juvenile bull trout
- Habitat uncertainty
 - SDM
 - Explicit predictions of occupancy
- eDNA sampling
 - What is it
 - Why use it
- Bull trout + eDNA
- Deciding where to sample
- Early results
- Closing the circle
- Better decisions
- Other species

You

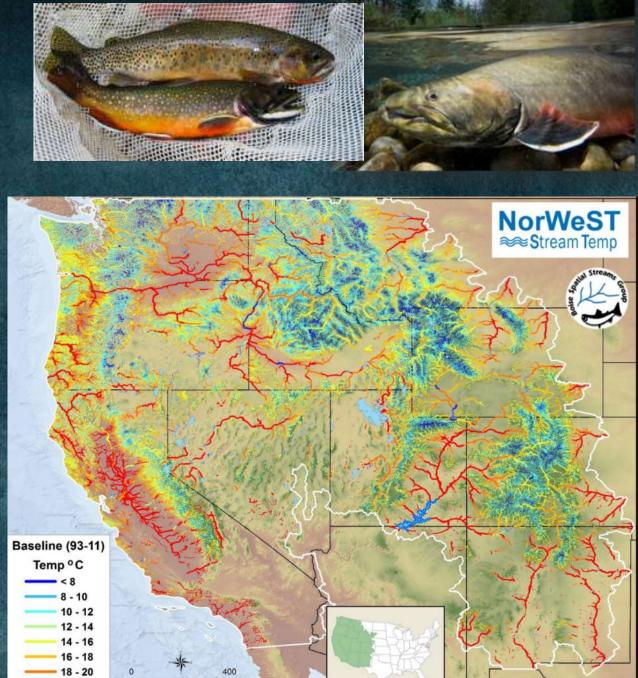


Why choose juvenile bull trout?

- ESA-listed as threatened
- Presence dictates land & water management & planning
- Widespread in PNW
- Often rare
- Difficult to detect
- Juveniles constrained by water temperature, vulnerable to nonnative spp.
- = candidate for occupancy modeling to identify suitable habitat



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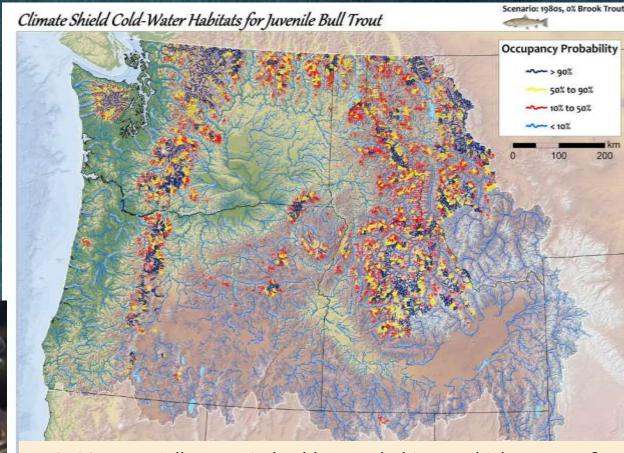
Identifying climate refugia for native trout – the Climate Shield

- Climate to cold-water habitat
- Predictions
 - Accurate & sufficient
 - Address invasive species
 - Empirical
 - Precise
 - Range-wide
- Projections
 - Address climate change
- Validation

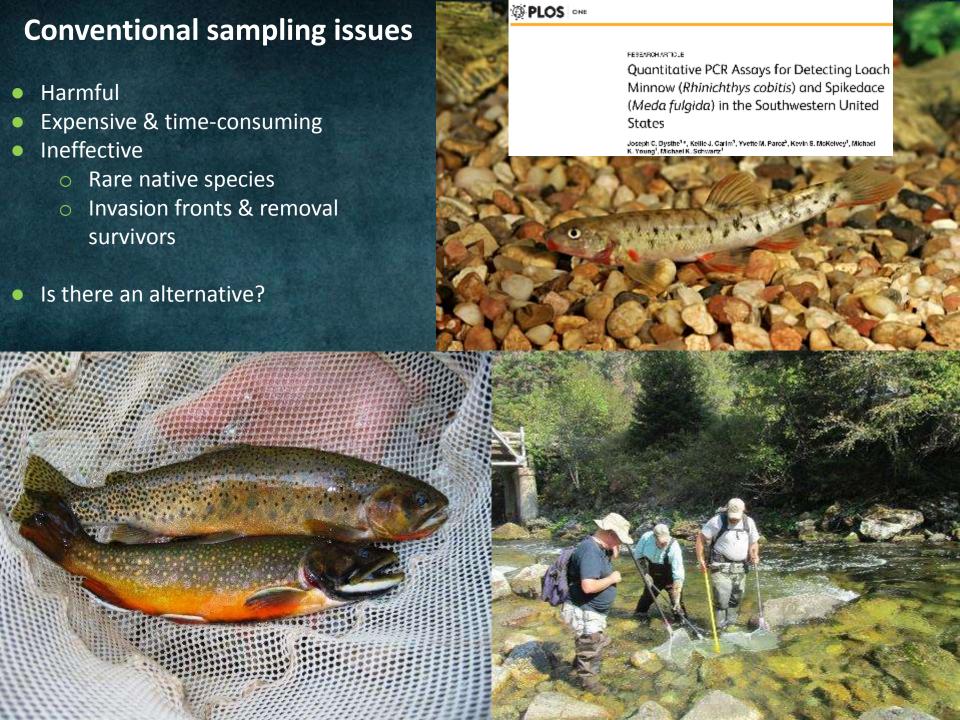




https://www.fs.fed.us/rm/boise/AWAE/projects/ClimateShield.html or Google "cold-water climate shield"



~3700 potentially occupied cold-water habitats: which ones are?



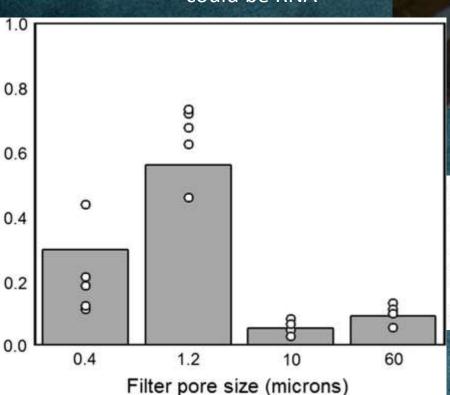


- Environmental = "free"
- Mitochondrial (usually)
 - Durable
 - Abundant
- Nuclear options

Proportion of eDNA captured

Dolly Varden & coastal bull trout

*could be RNA





Taylor M. Wilcox^{1,2} · Kevin S. McKelvey² · Michael K. Young² · Winsor H. Lowe¹ · Michael K. Schwartz³

Why use eDNA sampling: efficiency

- Fast
- Portable
- Stable
- Cost: pennies on the dollar, minutes on the hour
- Rapid, broad-scale surveys are feasible

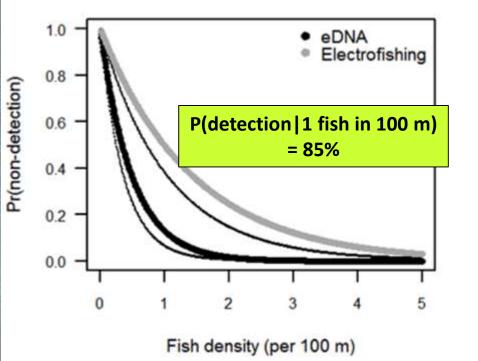




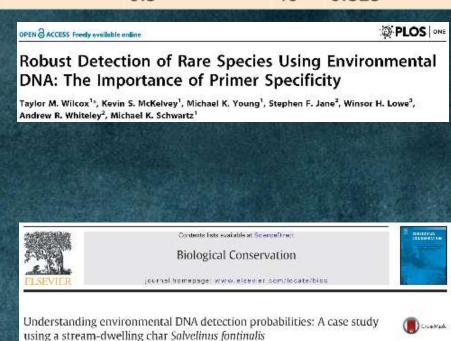


Why use eDNA sampling: accuracy

- Reliably* species-specific
- Sensitivity: high & quantified
 - Release rate: ~500 copies/sec
 - Detection threshold: 1 copy
- Very good at detecting rare species
- Occupancy estimates are robust



DNA Source	DNA Concentration Copies / ul	N	Proportion Successful						
Brook Trout	315.5	40	1						
	62.5	40	1						
	12.5	40	1						
	2.5	40	1						
	0.5	40	0.825						



Taylor M. Wilcox 3,8,4, Kevin S. McKelvey 3, Michael K. Young 5, Adam J. Sepulveda 6, Bradley B. Shepard 5.1,

Stephen F. Jane ", Andrew R. Whiteley", Winsor H. Lowe 1, Michael K. Schwartz

Why use eDNA sampling: revolutionary

- Apply a consistent approach
- Craft a sampling design
- Engage the stakeholder community
- Defensible, precise, broad-scale occupancy estimates for priority species in real time for reasonable cost



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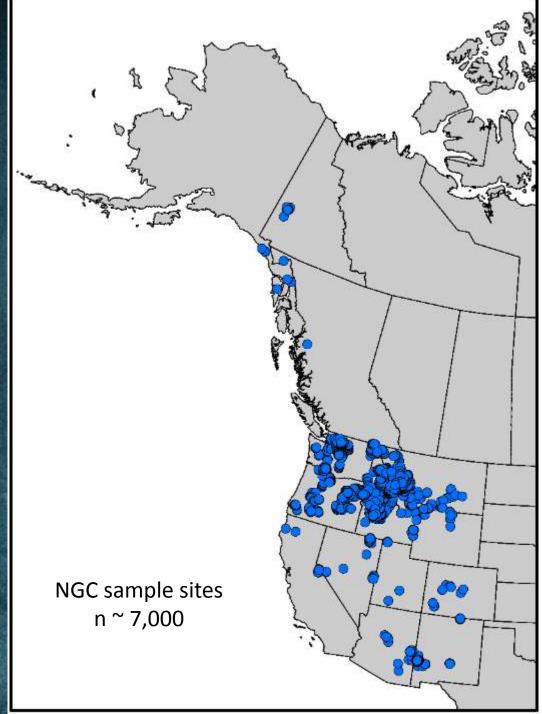
A Protocol for Collecting Environmental DNA Samples From Streams

Kellie J. Corim, Kevin S. McKelvey, Michael K. Young, Tayfor M. Wilcox, and Michael K. Schwartz.



Why eDNA: many applications

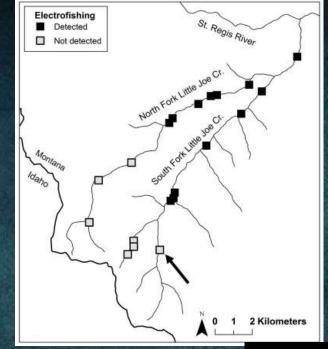
- Trout: rainbow, westslope cutthroat,
 Yellowstone cutthroat, brown
- Charr: bull, brook, Dolly Varden, lake,
 Arctic
- Salmon: Chinook, chum, coho, pink, sockeye
- Arctic grayling
- Any salmonid
- Pacific & brook lamprey
- Game fish/invaders: northern pike, sauger, walleye, smallmouth bass
- Non-game fish: sculpin (several), northern leatherside chub, loach minnow, spikedace
- Amphibians: Rocky Mountain tailed frog, western toad
- Mussels: western pearlshell, California floater
- Invertebrates: opossum shrimp
- North American river otter
- Harlequin duck
- Yuma clapper rail
- Your species here...

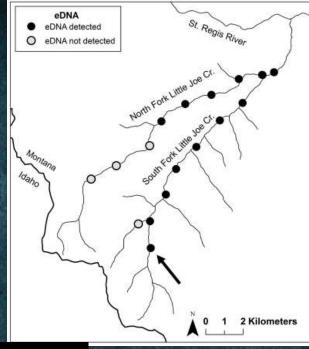


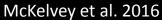
Using eDNA sampling to detect bull trout

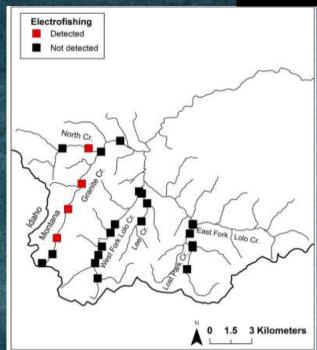
- ESA listed as threatened
- Dictates land & water management & planning
- Widespread rare
- Difficult to detect
- Juveniles constrained by environment/community
- = ideal candidate for eDNA sampling
- Test: Montana 2014
- Confirmed known habitats
- Discovered new ones

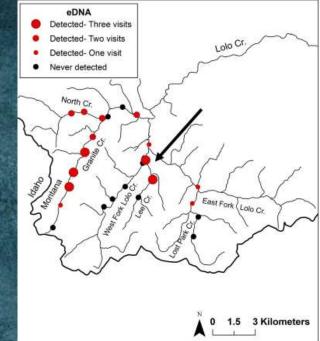






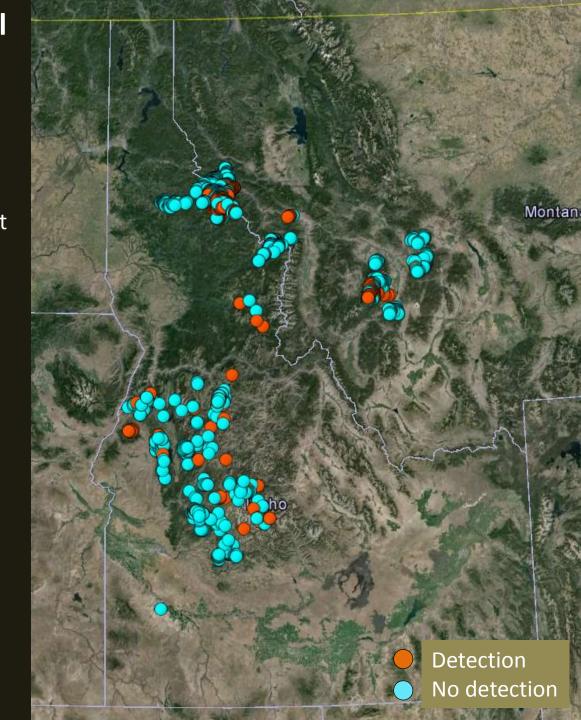


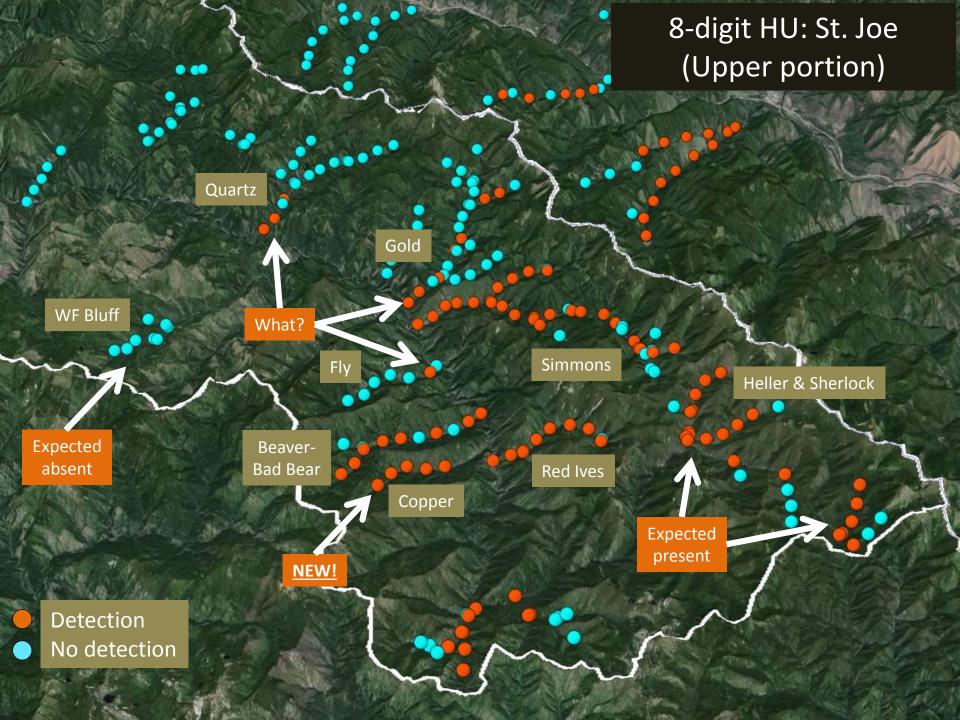


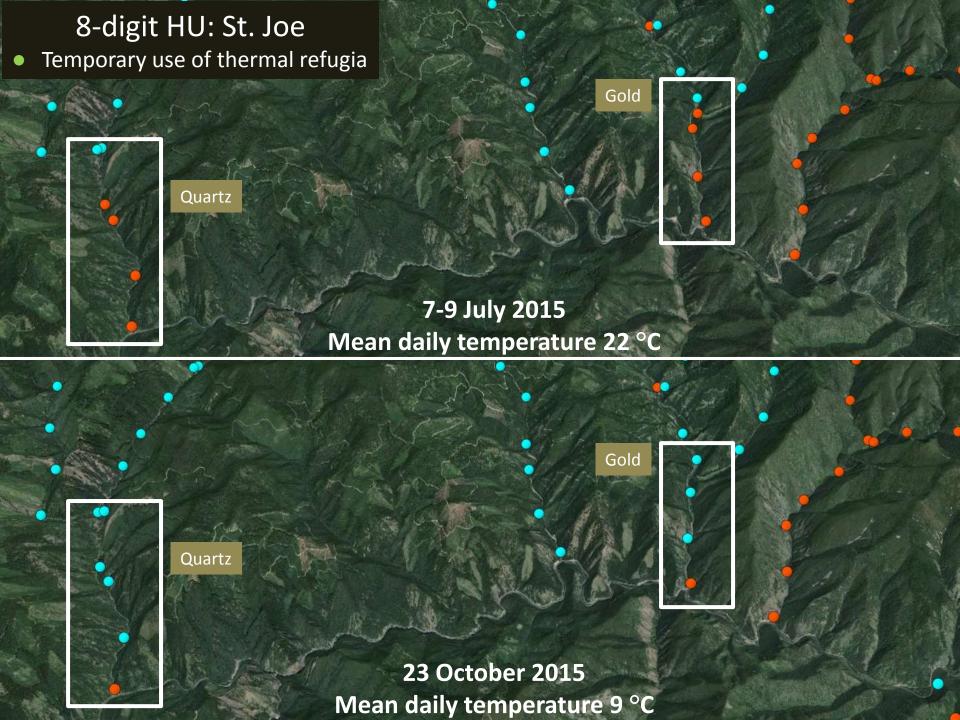


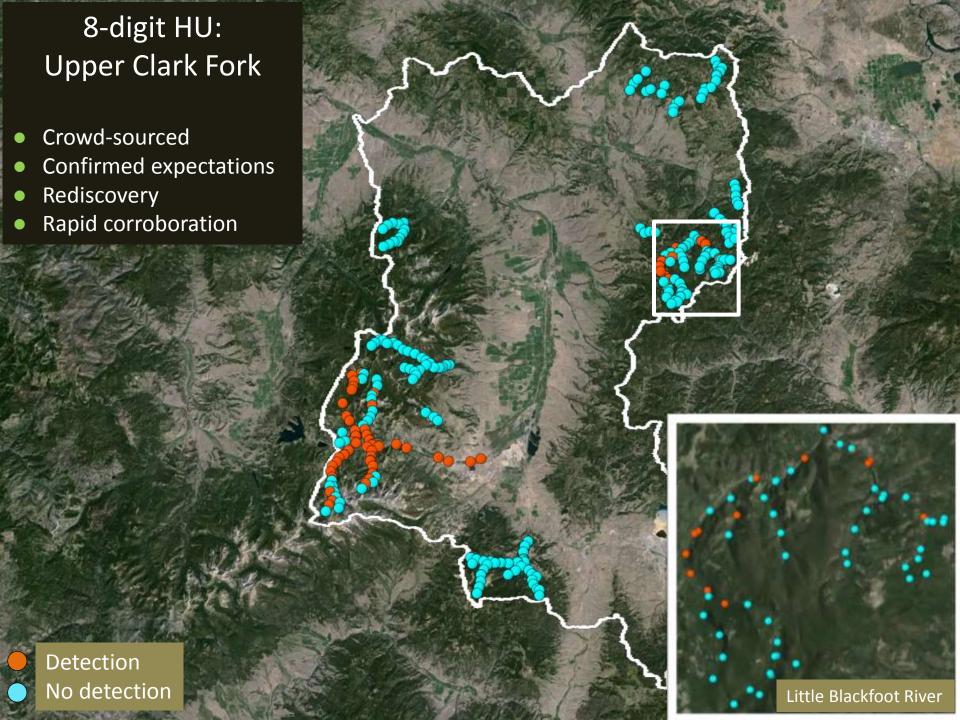
Update: the rangewide bull trout eDNA project

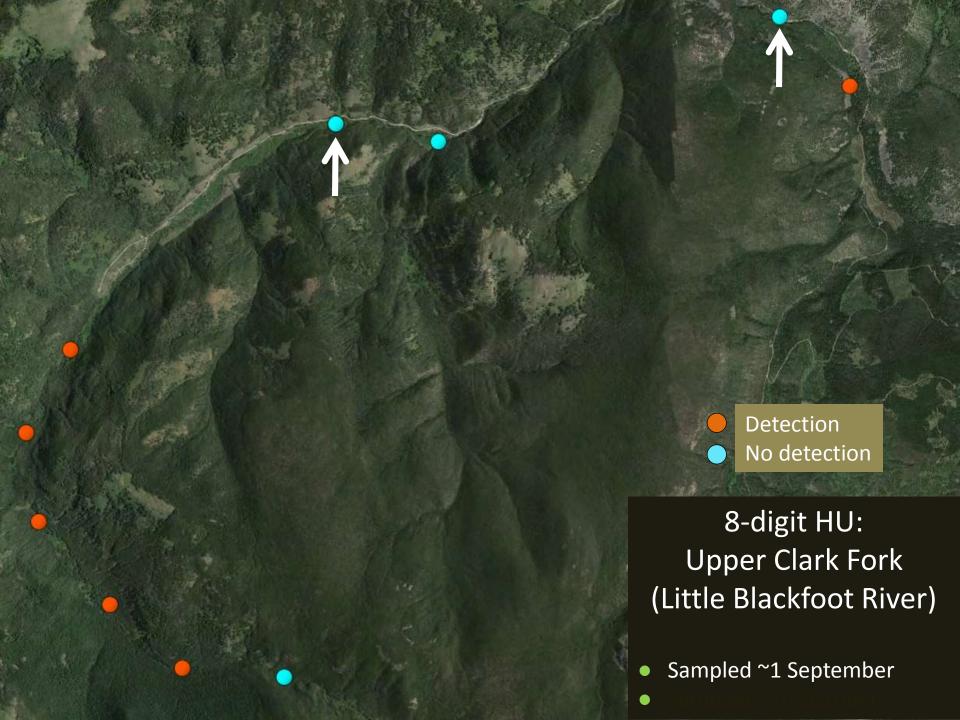
- Target: natal bull trout habitats
 - Cold-water habitats that are part of the Climate Shield
 - USFWS-designated critical habitat for bull trout spawning & rearing
 - Habitats about which recent occupancy is unknown
- Grain: sites at 1-km intervals
- Scope: all 8-digit U.S. HUs
- Timing
 - 2015: 500+ samples
 - 2016: 3,000+ samples
 - 2018: the rest of the range
- Cost
 - N. ID/W. MT: FREE!
 - S. ID/E. OR/WA: ½ price

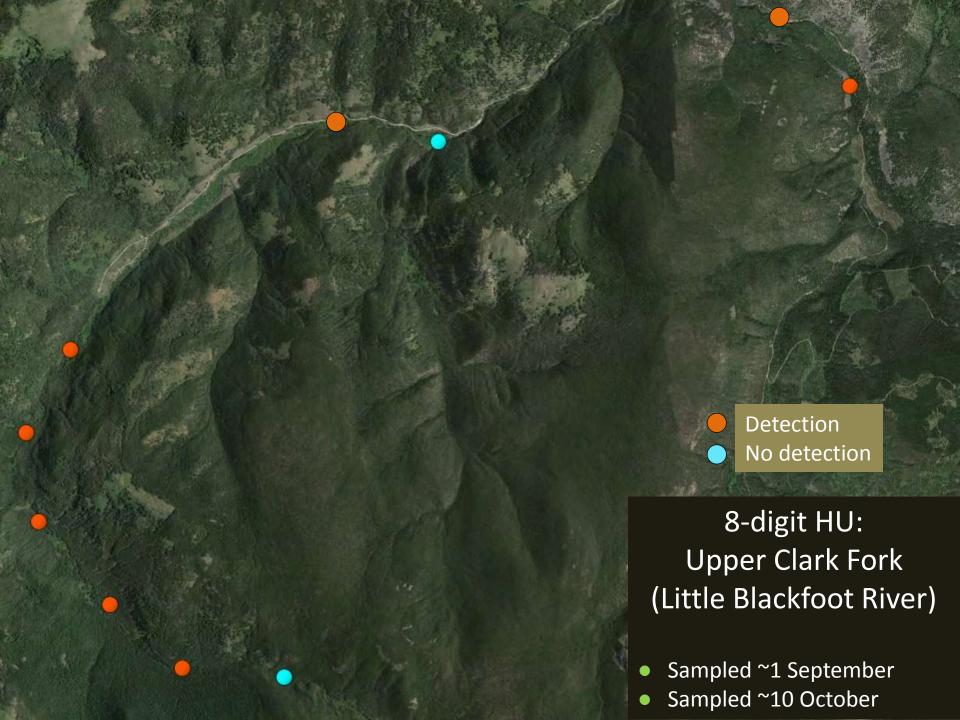












The Rangewide Bull Trout eDNA Project: want to help?

Visit our website: www.fs.fed.us/rm/boise/AWAE/projects/BullTrout eDNA.html

or Google "rangewide bull trout eDNA project"

- Contact us to get your library card
- Follow the simple instructions

A Protocol for Collecting Environmental DNA Samples From Streams

Kellie J. Carim, Kevin S. McKelvey, Michael K. Young, Taylor M. Wilcox, and Michael K. Schwartz





The built took is an ESA-listed species with a historical range that encompasses many waters across the Northwest. Though more abundant, built trout have declined an many locations and as at take from a therefore clinical encounters, and take the depositions histories interest and placeting select on sound and process information is a dearning select on sound and process information is a dearning and experience take. To overcome this problem, we employ 1 predictions from the range-rade, spatially precise Climate Shield model on the location of natal histories of built troot with 2) a sampling template from the range-rade, spatially precise Climate Shield model on the location of natal histories of built troot with 2) a sampling templater of the Natal Shield model on the probability of detecting built troot presence using environmental DNA (eRNA), sampling (McKelvey et al. 2016. The template constitute of a master set of geospatially referenced sampling locations at 1-km entervals within each cold-rester habitat. We also identified sampling locations at this same interval based on the USFMS's designation of critical spawrang and reason globals. Based on field tests of eNNA detection publishing conducted by the National Geomotoc Center for Wildlitts and Fish Consequentian, this scanding propriate most of built most, in well as provide insights on non-quanting labitats used by achief and sububbit feb. The zenit well be a rapid, obsert, and repeatable range-order assessment of natal habitates of this species.





Website: Get a bull trout hunting map

rather, if you can sample one or more cold-water habitats in their entirety, then we welcome your participation.

To make that possible, we will provide you with all you need to conduct eDNA field sampling for juvenile bull took. That includes:

1) A protocol that explains how to collect eDNA samples.

2) Additional guidelines specific to the bull trout eDNA survey project

3) A map and spreadsheet of eDNA points to guide your sampling.

4) The loan of a pemp set with a battery & charger. We operate a "tool library" i.e., you can sesserve a pump set for use during a particular time. The number of pump sets is limited and demand is high, so it's important to seserve one. It's also critical to return it when you are done to permit others to start their sampling. If you want to buy your own pump set—which gives you mose flexibility with respect to when you sample—we can give you the specifications.

 Field kits for the collection and storage of eDNA samples. To ensure consistency in sampling and guarantee stendiny of the supplies, we prefer to provide the field kits to you.

Once sampling is complete, return the pump set, field kits, and collected samples. In a few weeks, we'll share with you whether and where built trout were present. And at the end of each year, we'll post an interactive map of the results of sampling across the range of built tout on our results page.







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field sampling instructions ("Participating in the Bull Trout eDNA Survey, important caveats"). To get started, prant or plot the map of showing the 5-digit HLX to be sampled and identify which potential bull trout thesans are of inferent (and can be sampled in their entirety). Files may be periodically updated, so confirm that you have the most current versions before starting field work. Next, downdoad the Exod file with the eDNA sample are coordinates more that a GPS unit will be required for accurate field savingation.

Bull Trout eDNA Sample Sites

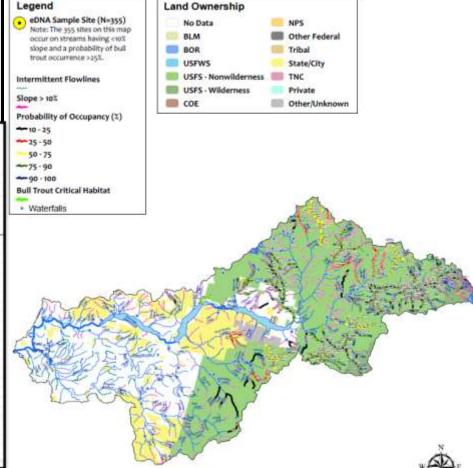
Scenario: 1980s, 0% Brook Trout

NHD Unit: 17080002 (Lewis)



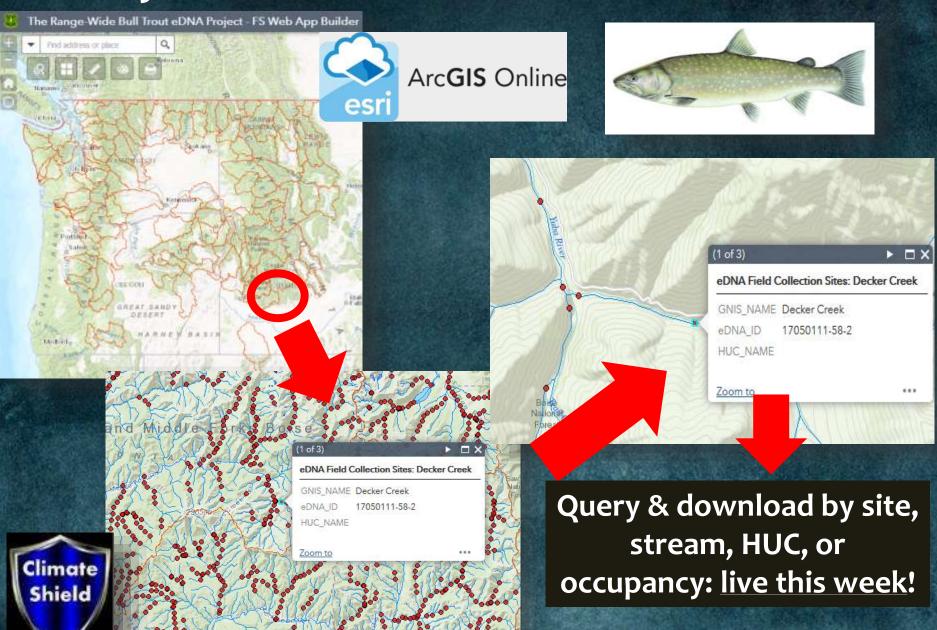
guide your eDNA surveys for

fats anywhere throughout their ims (>0.000) were identified using the severe in 512 streams (Isaak et al.





Dynamic Web Data Portal: eDNAtlas



eDNArchive

- 1 eDNA sample = many species
- Permanent archives of biodiversity
- ~10% of samples run for other spp.















