

**A DIATOM CALIBRATION SET
FOR THE
CASCADE MOUNTAIN ECOREGION**

Prepared for

**PACIFICORP
Centralia, WA**

by

**J.M. Eilers¹
P.R. Sweets²
D. F. Charles³
K.B. Vaché¹**

**E&S Environmental Chemistry, Inc.
Corvallis, OR**

April 23, 1998

¹ E&S Environmental Chemistry, Inc., P.O. Box 609, Corvallis, OR 97330

² Department of Biology, University of Louisville, Louisville, KY 40292

³ Academy of Natural Sciences, 1900 Benjamin Franklin Pkwy., Philadelphia, PA 19103

TABLE OF CONTENTS

LIST OF TABLES	iii
LIST OF FIGURES	iv
TERMS, ABBREVIATIONS, AND UNITS	v
EXECUTIVE SUMMARY	vii
ACKNOWLEDGMENTS	viii
A. INTRODUCTION	1
B. BACKGROUND ON DIATOM INFERENCE TECHNIQUES	3
C. METHODS	8
1. Lake Selection	8
2. Field and Analytical Methods (1996)	18
3. Diatom Taxonomic Identification	19
4. Inference of Water Quality Characteristics Using Weighted Averaging (WA)	21
5. Diatom Inferences	25
D. RESULTS	27
1. Chemistry and Morphometry of Study Lakes	27
2. Diatom Taxonomy	34
3. Statistical Diatom Ordination and Regression	41
4. Summit Lake Reconstruction	61
E. DISCUSSION AND CONCLUSIONS	70
F. LITERATURE CITED	73
G. APPENDICES	78

EXECUTIVE SUMMARY

A diatom calibration set was constructed for the Washington and Oregon Cascade Range for the purpose of assessing water quality changes in Cascade lakes. The 48 lakes used for the analysis were based on 27 lakes sampled in 1996 and 21 lakes sampled previously. Among the 21 previously sampled lakes 19 were in Mount Rainier National Park. The relationship between diatom assemblages and water chemistry and other lake attributes was determined using correspondence analysis (CA) and canonical correspondence analysis (CCA). The measured environmental variables captured 90% of the variance along the first ordination axis. The amount of variance explained by subsequent axes was comparatively minor. The variables that best explained the diatom assemblages were pH, conductivity, acid neutralizing capacity, magnesium, lake depth, and total phosphorus. Among these variables, pH ($s=0.31$, $r^2=0.89$) provided the strongest predictive power for diatom-inferred (DI) reconstructions in the Cascades. Several subsets of lakes were explored to improve the pH calibration including a subset of lower pH ($\text{pH} < 7.5$) lakes ($n=38$, $s=0.27$, $r^2=0.76$) and a subset excluding the lakes from Mt. Rainier ($n=29$, $s=0.25$, $r^2=0.94$). However, other variables such as conductivity ($s=24.8 \mu\text{S}$, $r^2=0.93$) and total phosphorus ($s=5.1 \mu\text{g/L}$, $r^2=0.62$) also show potential for future applications in the Cascades. The calibration equations were used to reconstruct water quality changes in Summit Lake, WA, a low pH (5.3-5.9), low conductivity (3-5 μS) lake northwest of Mount Rainier. The diatom-inferred pH and conductivity values showed no significant changes over the previous 3150 years within the standard errors of the predictions (0.31 for pH and 4.9 μS for conductivity). The Cascade diatom calibration set is a valuable tool for reconstructing water quality changes for a variety of purposes including changes associated with atmospheric deposition, timber harvest, fisheries management, and recreational activities. The value of the calibration set can be enhanced by collecting additional water chemistry data for lakes presently in the data base, standardizing the diatom taxonomy from samples collected prior to 1996, resolving the taxonomic ambiguities associated with taxa such as *Aulacoseira distans*, and continuing to add additional lakes to the data base.

ACKNOWLEDGMENTS

This research was funded under Contract # PO24968 from PacifiCorp to E&S Environmental Chemistry, Inc. The technical representative for PacifiCorp was David Shilton and the senior procurement specialist was R. Craig Smith.

Assistance in collection of field samples was provided by Michael Wing and Joseph Bernert of E&S. Sediment samples from Mt. Rainier National Park were collected by Barbara Samora. Analysis of major ion chemistry was provided by the USDA Forest and Range Experiment Station in Fort Collins, CO under the direction of Louise O'Deen. Analysis of phosphorus and chlorophyll was conducted by Jim Sweet, Aquatic Analysts, Portland, OR. Kalina Manoylova, University of Louisville, assisted with the diatom counting and taxonomy. Dr. Mark Whiting performed the diatom taxonomy on slides from Mount Rainier National Park collected in 1990.

Supplemental funding for analysis of additional diatom samples was provided by Mount Rainier National Park through Barbara Samora and Mt. Baker-Snoqualmie National Forest through Janice Peterson.

LIST OF TABLES

1. Study lakes for the diatom calibration set	8
2. Analytical methods for analysis of lake samples collected in 1996 for the diatom calibration set	18
3. Water chemistry for calibration lakes	28
4. Summary statistics for calibration lakes and major subsets of study lakes	30
5. Location and physical characteristics of study lakes	31
6. Summary statistics for physical attributes of study lakes	33
7. Occurrence and optima (abundance weighted mean) for taxa and taxa groups used in ordinations and inference models	35
8. Correlation matrix for 12 environmental variables, 35 lakes	43
9. Eigenvectors calculated in CCA ordination analyses for Cascade Lake subset	44
10. Summary of inference statistics from weighted averaging regression of selected environmental variables against the diatom assemblages	50
11. pH calibration regression statistics(weighted-averaging, with tolerance) for the Cascade calibration set	57
12. Weighted-averaging regression statistics for diatom-inferred conductivity (μS) in the Cascade calibration set	59
13. Morphometry of Summit Lake, WA	61
14. Surface water chemistry of Summit Lake, WA and adjacent pond	
15. Diatom-inferred reconstruction of lake pH and conductivity (μS) for Summit Lake, WA using the Cascade calibration set from this study and the Sierra Nevada (Whiting et al. 1989) calibration set	64
	68

LIST OF FIGURES

1. Location of lakes in the Cascade Diatom Calibration Set	10
2. The distribution of Cascade diatom calibration set lakes superimposed over the cumulative frequency distribution of pH (a) and acid neutralizing capacity (ANC; $\mu\text{eq/L}$) (b) for the lake population in the Oregon and Washington Cascades as defined in the Western Lake Survey (Eilers et al. 1987)	11
3. The distribution of Cascade diatom calibration set lakes for a) conductivity (μS); b) total phosphorus (TP; $\mu\text{g/L}$)	12
4. The distribution of Cascade diatom calibration set lakes for a) calcium ($\mu\text{eq/L}$); b) magnesium ($\mu\text{eq/L}$)	13
5. The distribution of Cascade diatom calibration set lakes for a) sodium ($\mu\text{eq/L}$); b) potassium ($\mu\text{eq/L}$)	14
6. The distribution of Cascade diatom calibration set lakes for a) sulfate ($\mu\text{eq/L}$); b) chloride ($\mu\text{eq/L}$)	15
7. The distribution of Cascade diatom calibration set lakes for a) ammonium ($\mu\text{eq/L}$); b) elevation (m)	16
8. The distribution of Cascade diatom calibration set lakes for a) lake area (ha); b) depth (m) . .	17
9. Schematic representation of the process of developing and applying the diatom calibration set for the Cascade Mountain Ecoregion	24
10. <i>Aulacoseira distans</i> from Big Lake, OR.	34
11. Electron micrograph of <i>Cyclotella stelligera</i>	40
12. <i>Cymbella</i> sp. 6 from Lucky Lake, OR	41
13. CCA ordination diagrams of sediment diatoms	46
14. CCA ordination of species scores with environmental variables for Cascade lakes	47
15. Measured pH versus diatom-inferred pH for all lakes in the Cascade diatom calibration set	49
16. Measured pH versus the residual (from Figure 15) of measured and diatom-inferred pH.	51
17. Measured pH versus diatom-inferred pH with two outliers removed	52
18. Measured pH versus diatom-inferred pH for lakes with measured pH less than 7.5	54
19. Measured pH versus diatom-inferred pH for lakes with measured pH less than 6.5.	55
20. Measured pH versus diatom-inferred pH for lakes in the Cascade-only subset (i.e., all Rainier lakes omitted)	56
21. Measured conductivity versus diatom-inferred (DI) conductivity for lakes in the Cascade diatom calibration set	58
22. Measured acid neutralizing capacity (ANC) versus diatom-inferred (DI) ANC for lakes in the Cascade diatom calibration set	60
23. Measured depth versus diatom-inferred depth for lakes in the Cascade diatom calibration set	62
24. Measured total phosphorus (TP) versus diatom-inferred total phosphorus for lakes in the Cascade diatom calibration set	63
25. Relative abundance (%) of the five most abundant diatom taxa present in the sediments of Summit Lake	64
26. Diatom-inferred (DI) pH (top) and diatom-inferred conductivity (bottom) for sediments from Summit Lake, WA generated by applying the calibration equations developed in this study to diatom data reported in Eilers et al. (1996a)	66

TERMS, ABBREVIATIONS, AND UNITS

Terms

Base cations (C_B)	the sum of calcium, magnesium, sodium, and potassium
Benthic	an organism that is attached to the substrate
^{14}C	carbon-14, a naturally-occurring isotope with a half-life of 5730 years
Canonical Correspondence Analysis (CCA)	a method to find linear combinations of variables such that linear combinations have maximal correlations
Chrysophytes	a group of algae with siliceous shells, but with different anatomical and ecological properties than diatoms
Correspondence Analysis (CA)	a statistical ordination procedure used to generate hypotheses about the relationship between species composition and underlying environmental factors
Diatom	a class of algae characterized by silicon shells that remain preserved after the death of the cell
Epiphyte	a plant that grows on the surface of another plant
Euplankton	organisms that are truly planktonic, i.e. live in the water column of a lake
^{210}Pb	lead-210, a naturally-occurring isotope with a half-life of 20.4 years; used for determining the age of lake sediments
Plankton	an organism that lives in the water column
Root Mean Square Error (RMSE)	a statistical measure of dispersion (standard deviation)
^{34}S	sulfur-34, a naturally-occurring isotope with a natural abundance of 4.2%
Trophic state	a classification of lake productivity ranging from oligotrophic (unproductive) to eutrophic (highly productive)
Tycoplanktonic	organisms that can live in the water column when currents move them from the substrate
Weighted-Averaging (WA)	a regression approach in which values with more information are weighted more heavily in computing the regression statistics

Abbreviations

AI	aluminum
ANC	acid neutralizing capacity (alkalinity)
C	carbon

Ca	calcium
CA	correspondence analysis
C _B	sum of base cations
CCA	canonical correspondence analysis
Cl	chloride
Cu	copper
DOC	dissolved organic carbon
Mg	magnesium
NH ₄	ammonium
NO ₃	nitrate
Pb	lead
pH	a measure of acidity; negative log of [H ⁺]
S	sulfur
Si	silicon
SO ₄	sulfate
Ti	titanium
TP	total phosphorus
V	vanadium
WA	weighted averaging
Zn	zinc

Units

ha	hectares (0.4047 acres)
L	liter (0.264 gallons)
m	meter (0.3048 ft.)
m ³	cubic meter (8.14×10^{-4} acre-ft)
µg/L	microgram (10^{-6} g) per liter
µS	microSiemen (10^{-6}), a unit of conductance
µeq/L	microequivalent per liter

A. INTRODUCTION

Paleolimnology, the study of lake history through analysis of lake sediments, includes a variety of techniques for assessing change. In some cases, changes are inferred on the basis of chemical composition of the sediment. In other approaches, watershed changes may be inferred on the basis of changes in vegetation as determined by pollen or from changes in rates of sediment accumulation. To determine changes in lake water quality, one of the most successful approaches has involved the use of specific algal remains preserved in the sediments. These applications were first developed to assess changes in lake acidification. Atmospheric deposition of sulfur and nitrogen compounds has caused acidification of lakes and streams in the eastern United States, Canada, and northern Europe. Attempts to characterize the status of aquatic ecosystems has been hampered by a paucity of high quality, long-term, water chemistry data. In an attempt to better determine the degree of lake acidification, researchers have noted that the changes in lake chemistry can be inferred from changes in species composition of a class of algae called diatoms.

Diatoms, an important algal group in lakewaters, have cell walls composed of silica. When the cells die, their exteriors (frustules) are often well preserved and incorporated into lake sediments. By sampling the sediments, it is possible to reconstruct changes in lake chemistry based on changes in the relative abundance of the diatoms and known preferences of specific taxa for various ranges of water quality. Early use of diatoms to assess historical changes in acidification employed qualitative approaches that allowed investigators to infer the direction of the pH changes in lakes, but provided little information on the magnitude of the change. More recent advances in the field have made it possible to reconstruct quantitative changes in lake chemistry using a technique in which the downcore changes in diatom taxa are assessed using diatom information collected from the surface sediments of a number of lakes. The lakes from which the surface sediments are collected is referred to as the calibration set. The calibration set makes it possible to develop quantitative estimates of historical water quality changes through the use of multivariate statistical techniques in which a species preference for given water quality conditions is mathematically related to the water quality in the lake and the diatom taxa present in the surface sediments. However,

because taxonomic preferences for specific water quality conditions vary among regions, it is necessary to develop calibration sets that are tailored for a specific region.

Diatom calibration sets have been developed for the Adirondack Mountains in New York (Charles et al. 1987, 1990), Florida (Charles et al. 1986), the Upper Midwest (Kingston et al. 1990), and the Sierra Nevada (Whiting et al. 1989). However, no calibration set was previously available for the Cascade Mountains.

B. BACKGROUND ON DIATOM INFERENCE TECHNIQUES

In the absence of long-term chemical monitoring data, generation of statistical inferences based upon diatom, and in some cases chrysophyte, fossil assemblages preserved in lake sediments is the best technique available to evaluate historical chemical changes (Charles and Norton 1986, Smol 1992). Diatoms (*Bacillariophyta*) and scaled chrysophytes (*Chrysophyceae*, *Synurophyceae*) are single-cell algae composed of siliceous valves and overlapping siliceous scales, respectively. The fossil remains of these organisms are excellent indicators of past lakewater chemistry because: (1) they are common in most lakes, (2) many taxa have rather narrow ecological (water chemistry) tolerances, (3) remains are generally well preserved in sediment, usually in very large numbers, and (4) they can be identified to the species level or below (Smol et al. 1984, 1986; Charles 1985; Charles and Norton 1986; Husar et al. 1991).

Paleolimnological reconstructions of past lakewater chemistry are based on transfer functions derived from relationships between current chemistry and diatom/chrysophyte remains in surface sediments. Predictive equations are developed from these relationships using regional lake data sets to infer past water chemistry. Several techniques have been developed and applied to infer lakewater pH. Calibration equations have also been developed for inferring the concentration of other constituents, including dissolved organic carbon, acid neutralizing capacity, salinity, trophic state, total aluminum, and monomeric aluminum.

Once developed, predictive equations can be applied to diatom assemblage data from lake sediment cores to infer past conditions. Trends within cores can be analyzed statistically to determine if they are significant (Birks et al. 1990). Inferred chemical data can be dated using ^{lead-210} (²¹⁰Pb) activity and compared with stratigraphies of other lake sediment characteristics such as pollen, charcoal, coal and oil carbonaceous particles, polycyclic aromatic hydrocarbons, lead, zinc, copper, vanadium, calcium, magnesium, titanium, sulfur, and others that provide a record of atmospheric inputs of materials associated with the combustion of fossil fuels and of watershed disturbance (Heit et al. 1981, Tan and Heit 1981, Charles and Norton 1986). With these data, in addition to knowledge of watershed events and some historical information on regional atmospheric

emissions of sulfur and nitrogen, it is often possible to assess with reasonable certainty whether lakes have been affected by acidic deposition, and to what extent (Husar et al. 1991, Charles et al. 1989).

Diatom studies have documented the rates and magnitude of acidification of selected lakes in the Adirondack Mountains (Del Prete and Schofield 1981; Charles 1984, 1987; Charles et al. 1986, 1987, 1990; Sullivan et al. 1990; Cumming et al. 1992), New England (Davis et al. 1983), the Upper Midwest (Kingston et al. 1990), Florida (Charles et al. 1986), the Sierra Nevada (Whiting et al. 1989), Ontario (Dixit 1983; Dixit et al. 1987, 1989a,b), Sweden (Renberg and Hellberg 1982, Renberg and Wallin 1985, Renberg et al. 1993), Finland (Tolonen and Jaakkola 1983), Norway (Davis and Anderson 1985, Davis and Berge 1980), Scotland (Flower and Battarbee 1983, Flower et al. 1987), West Germany (Arzet et al. 1986), and the Netherlands (van Dam et al. 1988).

In order to interpret information contained in the sedimentary profile, it is necessary for the signal associated with the sedimentary algal microfossils not to have been modified subsequent to deposition. Approximate timing of sediment interval deposition must also be determined using radioisotope dating techniques (e.g., ^{210}Pb) or by examining changes in other sedimentary constituents, such as pollen or charcoal, that can be correlated with documented major fires, land-clearing activities, etc.

A paleolimnological study by Davidson (1984) at experimentally acidified Lake 223 in the Experimental Lakes Area in Ontario illustrated the rapid response of diatoms to changes in water chemistry. Results from the sediment core showed close agreement with the plankton history measured in the water column. Similarly, paleolimnological inferences of recent changes in lakewater chemistry have been shown to correspond closely with measured values at three lakes near Sudbury, Ontario (Dixit et al. 1987, 1989b). Dixit et al. (1987) reconstructed the pH of Hannah, Lake, near Sudbury, using diatom remains. Between about 1880 and 1975, the inferred lakewater pH declined from about 6.0 to 4.6. After the lake was limed in 1975, its measured pH increased from 4.3 to 7.0. This increase was also reflected in the diatom-inferred values (Dixit et al. 1987). A paleolimnological investigation (Dixit et al. 1989a) of acidification and subsequent recovery of Swan Lake, near Sudbury, indicated that paleolimnological reconstructions of recent chemical change

corresponded well with measured values. Also of significance were the large observed declines in trace metal concentrations coincident with pH recovery. The changes in metal concentrations did not inhibit accurate pH reconstruction. Dixit et al. (1989b) also used diatoms and chrysophytes to reconstruct the recovery of Baby Lake subsequent to the closure of the nearby (1 km distance) Conistan Smelter in 1972. Measured lakewater pH increased from 4.2 in 1972 to 6.5 in 1987. This recovery was closely mirrored by shifts in the diatom and chrysophyte species composition and inferred pH.

A number of techniques have been used to reconstruct lakewater chemistry, particularly pH, from sedimentary diatom remains. Paleolimnology as a quantitative science has evolved extremely rapidly over the past decade. Hustedt (1939) was the first to quantify diatom-pH relationships. Recognizing the strong empirical relationship between diatom distributions and lakewater pH, Hustedt defined the following pH occurrence categories:

- alkalibiotic - occurring at pH values > 7
- alkaliphilous - occurring at pH about 7, with widest distribution at pH > 7
- indifferent - equal occurrences on both sides of pH 7
- acidophilous - occurring at about pH 7, with widest distribution at pH < 7
- acidobiotic - occurring at pH values below 7, with optimum distribution at pH 5.5 and below

Diatom taxa can be assigned to these various categories based on published ecological information or regional patterns of distribution. Changes in the percentages of diatom valves in each pH category within a sediment core can be used to evaluate trends in lakewater pH.

Multiple linear regression analysis of measured lakewater pH with the percentage of diatoms in each pH category has been used to develop predictive equations that typically have smaller standard errors and confidence intervals for prediction of new points than the earlier indices (Charles 1985, Huttunen and Meriläinen 1986). Other multiple regression approaches involve the use of selected taxa, or principal components of taxa (Davis and Anderson 1985, Gasse and Tekaia 1983). The standard error for inferred pH for these techniques ranges between 0.25 and 0.40 pH units (Battarbee 1984, Charles and Norton 1986). Other techniques include detrended correspondence analysis (Huttunen and Meriläinen 1986), canonical correspondence analysis (ter Braak 1986, Stevenson et al. 1989), and a multiple regression technique using both diatoms and

chrysophytes (Charles and Smol 1988). ter Braak and Van Dam (1989) developed new methods using maximum-likelihood calibration based on Gaussian logit response curves of taxa against pH and on weighted averaging (WA). Oksanen et al. (1988) used weighted averaging, least squares, and maximum likelihood to calculate pH optima and tolerance of diatom taxa, and then used these estimates to predict pH of other lakes, using weighted averaging. Birks et al. (1990) suggested using the straightforward but heuristic approach of weighted averaging regression and calibration as a compromise between ecological realism and computational feasibility.

Diatoms and chrysophytes differ somewhat in their ecological characteristics and both can provide useful qualitative and quantitative information regarding temporal trends in the acid-base status of lake water. Chrysophytes are euplanktonic; diatoms are both planktonic and benthic, but the diatom flora of low pH (< 5.5) lakes are dominated by benthic forms (Battarbee 1984, Charles 1985, Charles and Smol 1988). Chrysophytes usually bloom in spring. Although planktonic diatoms are also most abundant in spring, most littoral species are common throughout spring and summer (DeNicola 1986, Jones and Flower 1986, Charles and Smol 1988). Thus, chrysophyte assemblages may provide a better reflection of spring snowmelt pH depressions. Chrysophyte assemblage composition seems to change more rapidly than diatom assemblages along the pH gradient for pH values \leq 5.5, and are, therefore, particularly useful indicators in this range. Chrysophyte scales are not always present in sufficient quantity and variety to be useful, however. We have found chrysophytes to be scarce in several Pacific Northwest lakes. For this reason and also because diatom inference equations have lower root mean square error (Sullivan 1990), we used diatoms for pH reconstructions in Cascade Mountain lakes.

By late 1986 (as the Paleoecological Investigation of Recent Lake Acidification [PIRLA-I] project was ending), category-based, multiple regression techniques were being replaced by theoretically superior gradient analysis techniques. The theory involved has been summarized, elaborated, and implemented primarily by ter Braak (ter Braak 1985, 1986, 1987a, 1988; ter Braak and Barendregt 1986; ter Braak and Looman 1986; ter Braak and Gremmen 1987; ter Braak and Prentice 1988). Gradient analysis theory is based on a species-packing model along environmental gradients, assuming a simple normal distribution of each species' abundance in samples along the

gradient. Although much of ter Braak's research concerns multivariate analysis of several environmental gradients simultaneously (primarily by ordination techniques), the potential for application of these methods to the reconstruction of single environmental variables was recognized as a desirable goal in acidification paleolimnology (Stevenson et al. 1989, ter Braak and van Dam 1989, Birks et al. 1990, Kingston and Birks 1990, Dixit et al., 1989a). With the availability of the computer programs CANOCO (ter Braak 1987b) and WACALIB (Line and Birks 1990) to implement gradient analysis theory, there is now little justification for using the previous category techniques.

Given the unimodal response model, several comparisons have been made between different implementations, primarily comparing maximum likelihood and weighted averaging methods. Although it was expected that maximum likelihood, implemented by Gaussian regression, would be superior to the computationally simpler weighted averaging method, this has not proven to be true (Oksanen et al. 1988, ter Braak and van Dam 1989, Birks et al. 1990, Kingston and Birks 1990). Problems with maximum likelihood implementation include the tendency to find local maxima rather than overall maxima in the data and a significant number of samples failing to converge (the numerical procedure is unable to pick a single most likely value). There was also no indication that there are fewer errors than associated with weighted averaging.

Paleolimnological inferences are performed in two steps, regression and calibration. In the regression step, curves are fitted to each taxon's distribution, and the properties of the estimated optimum (\hat{u}_k) and estimated tolerance (t_k) can be determined. The calibration step may be subdivided further between active samples (with associated environmental data) and passive samples (without associated environmental data). The algal assemblage in each active surface-sediment sample is used to obtain a single estimated value based on the optima, with or without tolerance down-weighting, and the relationship between predicted and observed environmental values emerges for the entire set of active samples. In the next calibration step, that of paleolimnological reconstruction, passive samples (sediment core samples) are used to obtain predicted environmental variable values, based on their taxa having the same characteristics as obtained for the active taxa in the regression step.

C. METHODS

1. Lake Selection

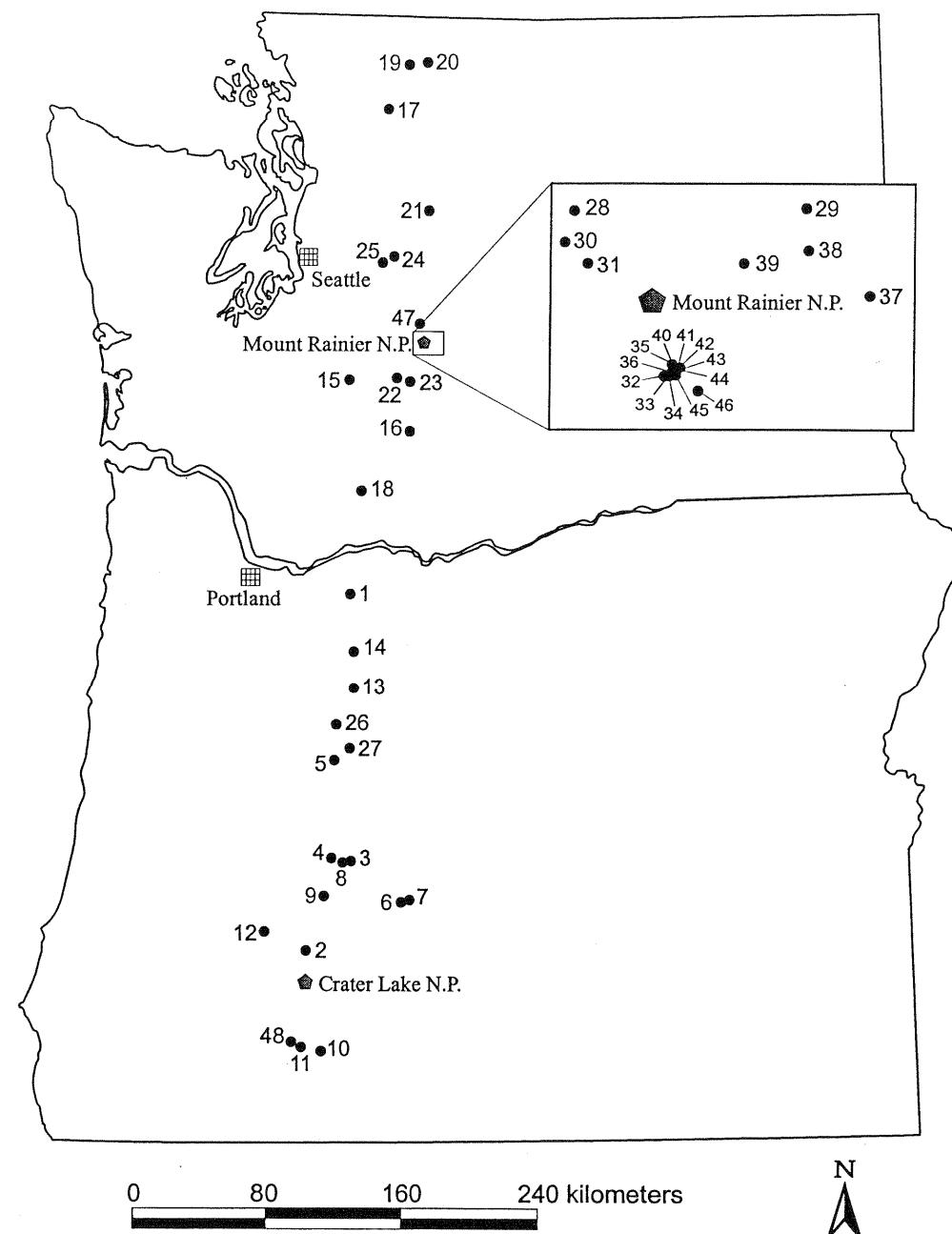
Twenty-five lakes were initially selected in the Oregon and Washington Cascades for sampling. This initial list was modified slightly to accommodate access problems. In addition, several changes in the list were made to provide for better geographical distribution and better representation along chemical gradients. The results from these lakes were combined with previously collected diatom and water chemistry data from other selected lakes in the region including data on 19 lakes and ponds in Mount Rainier National Park and two lakes previously reported for the Oregon Cascades (Eilers et al. 1996a,b), bringing the total number of lakes in the calibration set to 48. The 27 lakes sampled in 1996 were selected to provide adequate spatial distribution along the length of the Cascade Range. Particular emphasis was placed on selecting lakes that represented a broad range in pH (4.95-8.44), base cations (13-8246 $\mu\text{eq/L}$), and nutrients. The final list of study lakes is presented in Table 1 and their locations are shown in Figure 1. The study lakes provide a suitable representation of lakes in the Cascade Range based on cumulative frequency distributions of lakes in the study area (Figures 2-8).

Table 1. Study lakes for the diatom calibration set.

Lake	Core Date	Taxonomy	Reference
A. Newly Sampled Lakes			
Suttle Lake	11/9/96	R. Sweets	This Study
Marion Lake	10/7/96	R. Sweets	This Study
Sphagnum	9/8/96	R. Sweets	This Study
Nordrum	9/8/96	R. Sweets	This Study
Caskey Lake	9/6/96	R. Sweets	This Study
Dumbbell Lake	8/23/96	R. Sweets	This Study
Shellrock Lake	8/23/96	R. Sweets	This Study
Heather Lake	8/22/96	R. Sweets	This Study
Pyramid Lake	8/21/96	R. Sweets	This Study
Middle Thornton	8/20/96	R. Sweets	This Study
Unnamed	8/14/96	R. Sweets	This Study
Gertrude Lake	8/13/96	R. Sweets	This Study
Cora Lake	8/12/96	R. Sweets	This Study
Head Lake	7/30/96	R. Sweets	This Study

Table 1. Continued.

Lake	Core Date	Taxonomy	Reference
Summit Lake, OR	7/30/96	R. Sweets	This Study
Little Twin Lake	7/26/96	R. Sweets	This Study
Agency Lake	7/25/96	R. Sweets	This Study
South Heavenly Twin	7/25/96	R. Sweets	This Study
Lucky Lake	7/24/96	R. Sweets	This Study
Charlton	7/24/96	R. Sweets	This Study
Paulina	7/23/96	R. Sweets	This Study
East Lake	7/23/96	R. Sweets	This Study
Big Lake	7/9/96	R. Sweets	This Study
Lava Lake	6/20/96	R. Sweets	This Study
Mink Lake	6/20/96	R. Sweets	This Study
Diamond Lake	6/14/96	R. Sweets	This Study
Burnt Lake	6/8/96	R. Sweets	This Study
B. Previously Sampled Lakes			
Summit Lake, WA	8/24/95	R. Sweets	Eilers et al. 1996a
Unnamed Lake3	9/3/91	R. Sweets	This Study
Unnamed Lake4	9/3/91	R. Sweets	This Study
Unnamed Lake1	8/20/91	R. Sweets	This Study
Unnamed Lake2	8/16/91	R. Sweets	This Study
Unnamed Lake5	8/14/91	R. Sweets	This Study
Unnamed Lake6	8/14/91	R. Sweets	This Study
Unnamed Lake7	8/14/91	R. Sweets	This Study
Unnamed Lake8	8/13/91	R. Sweets	This Study
Unnamed Lake9	8/13/91	R. Sweets	This Study
Unnamed Lake10	8/12/91	R. Sweets	This Study
Lake Natasha	8/3/91	S. Dixit	Eilers et al. 1996b
Snow Lake	9/27/90	M. Whiting	National Park Service (unpubl. data)
Tipsoo Lake	9/25/90	M. Whiting	National Park Service (unpubl. data)
Shadow Lake	9/25/90	M. Whiting	National Park Service (unpubl. data)
Reflection Lake B	9/12/90	M. Whiting	National Park Service (unpubl. data)
Upper Palisades L	9/5/90	M. Whiting	National Park Service (unpubl. data)
Sunrise Lake	9/4/90	M. Whiting	National Park Service (unpubl. data)
Mowich Lake	8/27/90	M. Whiting	National Park Service (unpubl. data)
Green Lake	8/26/90	M. Whiting	National Park Service (unpubl. data)
Reflection Lake A	8/13/90	M. Whiting	National Park Service (unpubl. data)



1 Burnt	11 South Heavenly Twin	21 Heather	31 Unnamed	41 Unnamed
2 Diamond	12 Little Twin	22 Dumbbell	32 Reflection	42 Unnamed
3 Lava	13 Head	23 Shellrock	33 Unnamed	43 Unnamed
4 Mink	14 Summit, OR	24 Spagnum	34 Reflection	44 Unnamed
5 Big	15 Cora	25 Nordrum	35 Unnamed	45 Unnamed
6 Paulina	16 Gertrude	26 Marion	36 Unnamed	46 Snow
7 East	17 Caskey	27 Suttle	37 Tipsoo	47 Summit
8 Lucky	18 Unnamed	28 Green	38 Sunrise	48 Natasha
9 Charleton	19 Middle Thornton	29 Upper Palisades	39 Shadow	
10 Agency	20 Pyramid	30 Mowich	40 Unnamed	

Figure 1. Location of lakes in the Cascade Diatom Calibration Set. The inset for Mt. Rainier National Park reveals details of lake locations within the park.

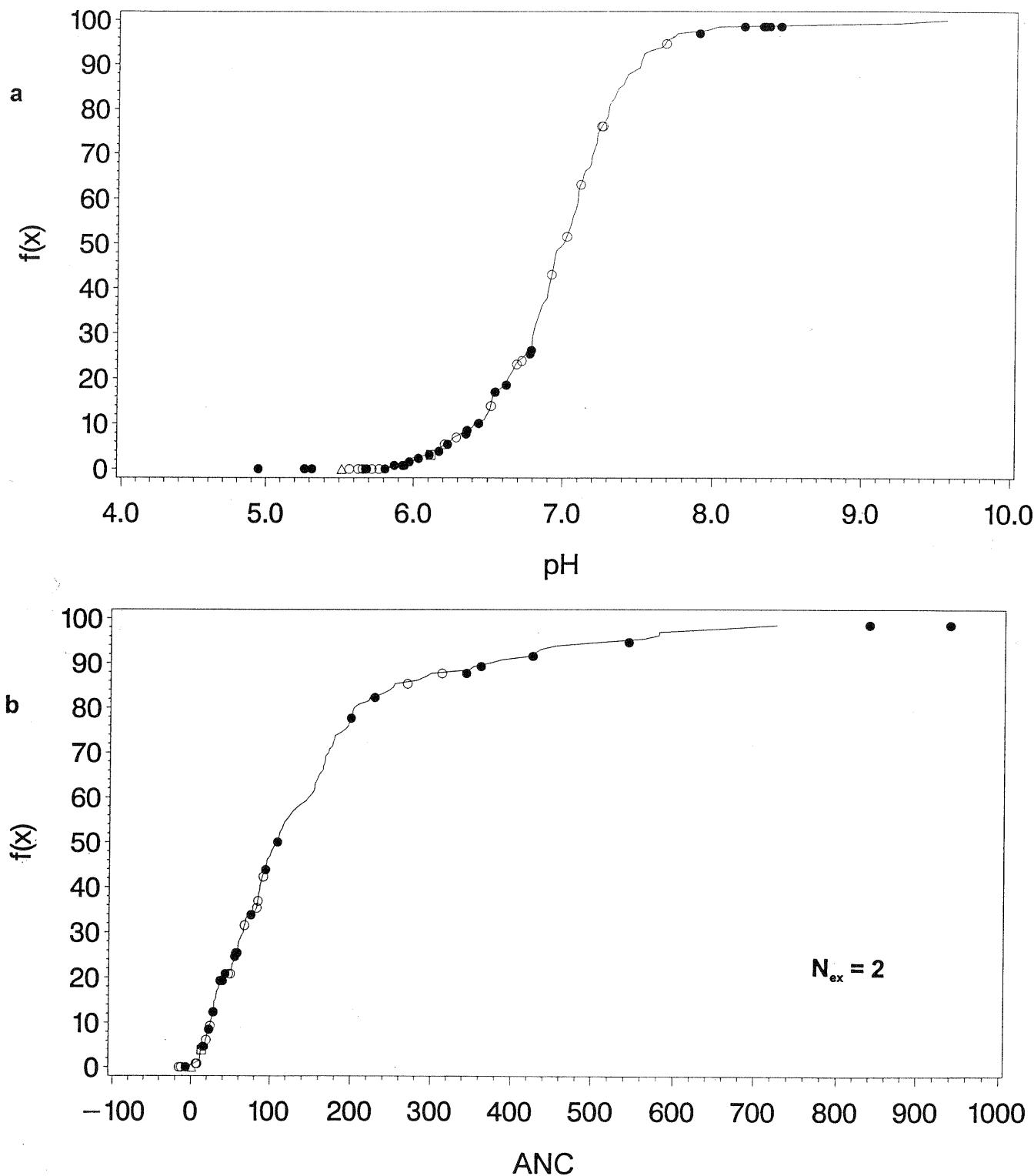


Figure 2. The distribution of Cascade diatom calibration set lakes superimposed over the cumulative frequency distribution of pH (a) and acid neutralizing capacity (ANC; $\mu\text{eq/L}$) (b) for the lake population in the Oregon and Washington Cascades as defined in the Western Lake Survey (Eilers et al. 1987). The solid circles (\bullet) represent lakes sampled in 1996. Lakes sampled prior to 1996 include those in Mount Rainier National Park (\circ) and two lakes sampled by Eilers et al. (1996a [Δ], b [\square]). The distribution for some variables has been truncated. The number of lakes extending beyond the displayed population distributions is indicated as N_{ex} .

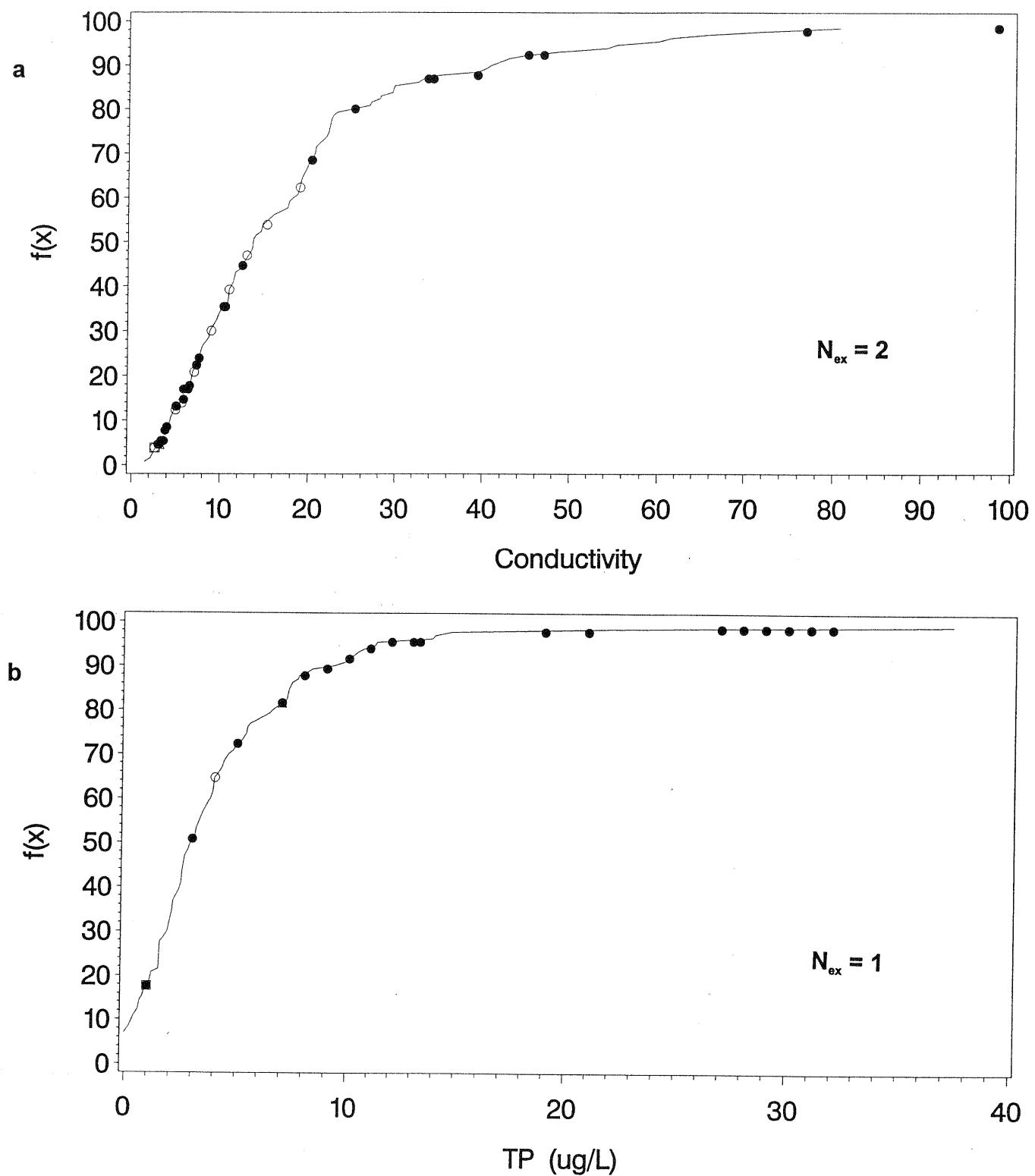


Figure 3. The distribution of Cascade diatom calibration set lakes for a) conductivity (μS); b) total phosphorus (TP; $\mu\text{g/L}$).

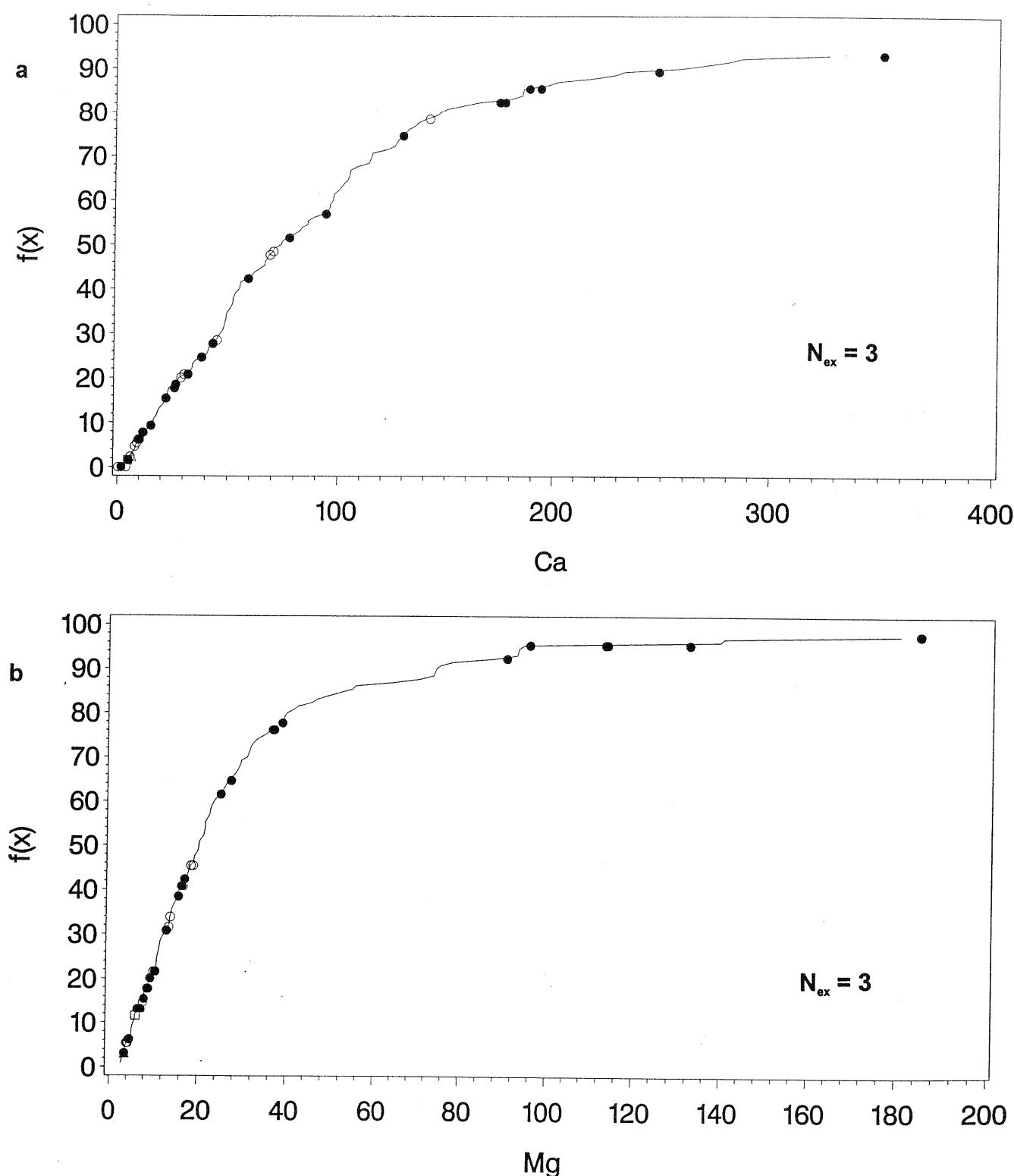


Figure 4. The distribution of Cascade diatom calibration set lakes for a) calcium ($\mu\text{eq/L}$); b) magnesium ($\mu\text{eq/L}$).

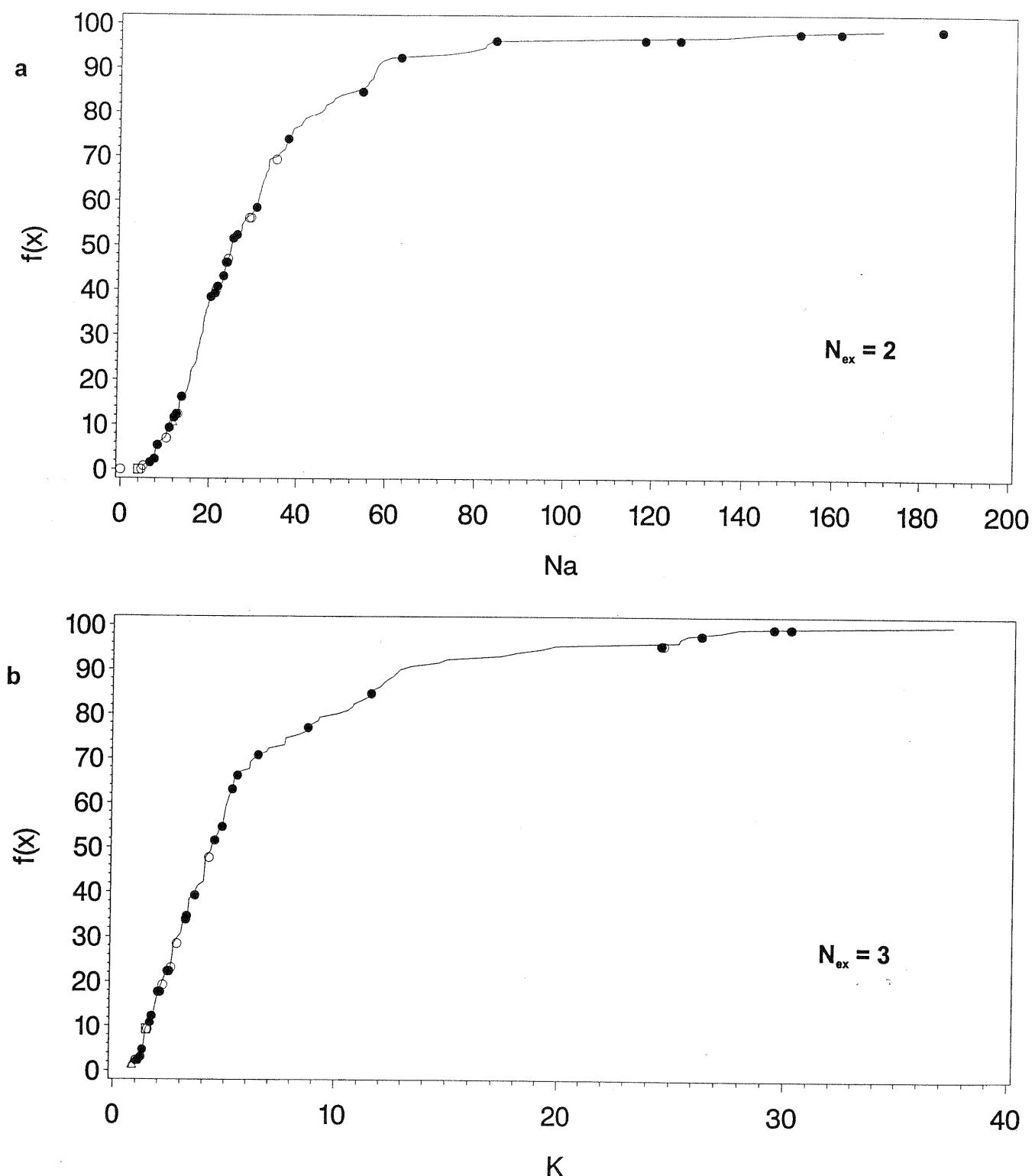


Figure 5. The distribution of Cascade diatom calibration set lakes for a) sodium ($\mu\text{eq/L}$) ; b) potassium ($\mu\text{eq/L}$).

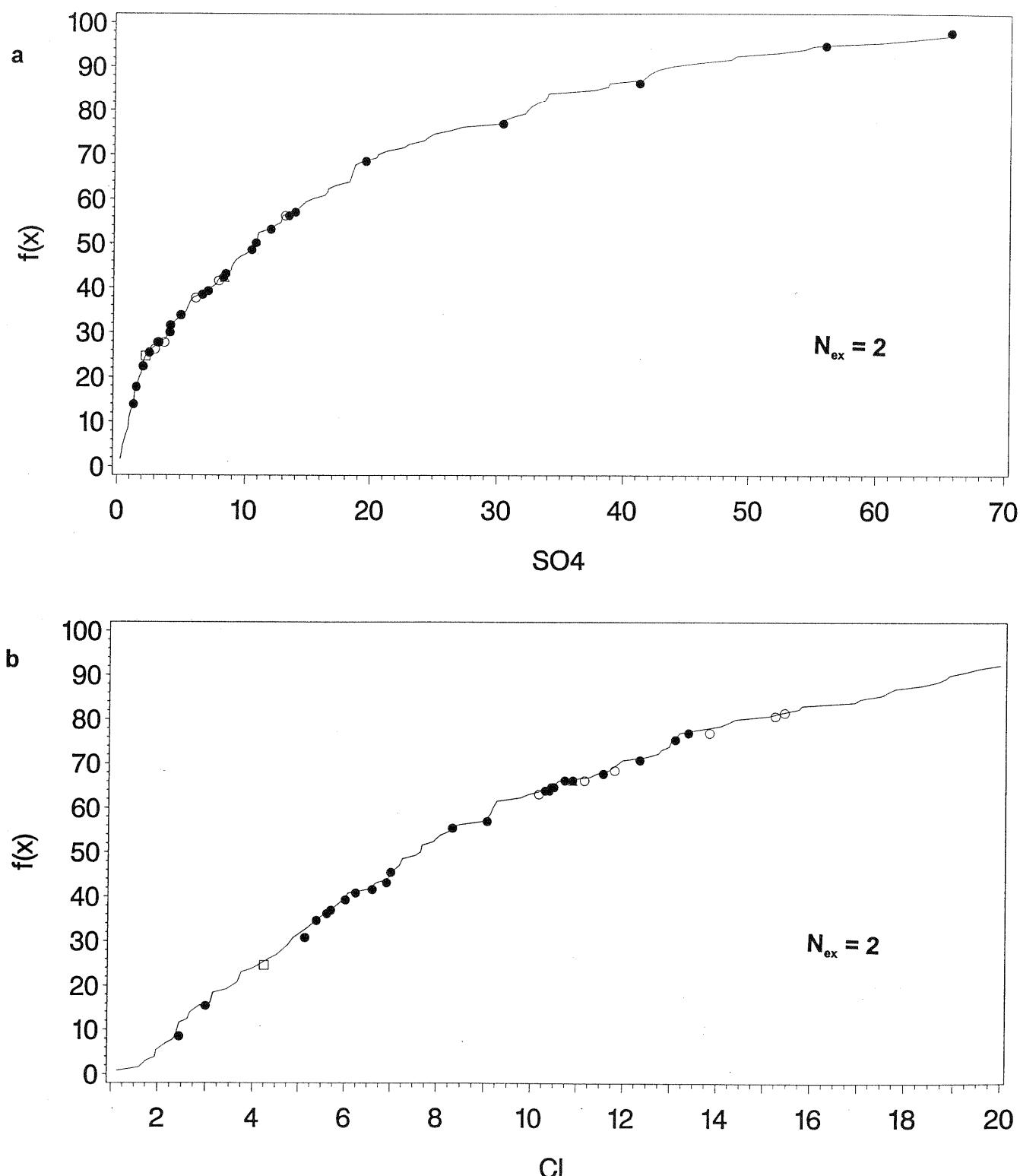


Figure 6. The distribution of Cascade diatom calibration set lakes for a) sulfate ($\mu\text{eq/L}$); b) chloride ($\mu\text{eq/L}$).

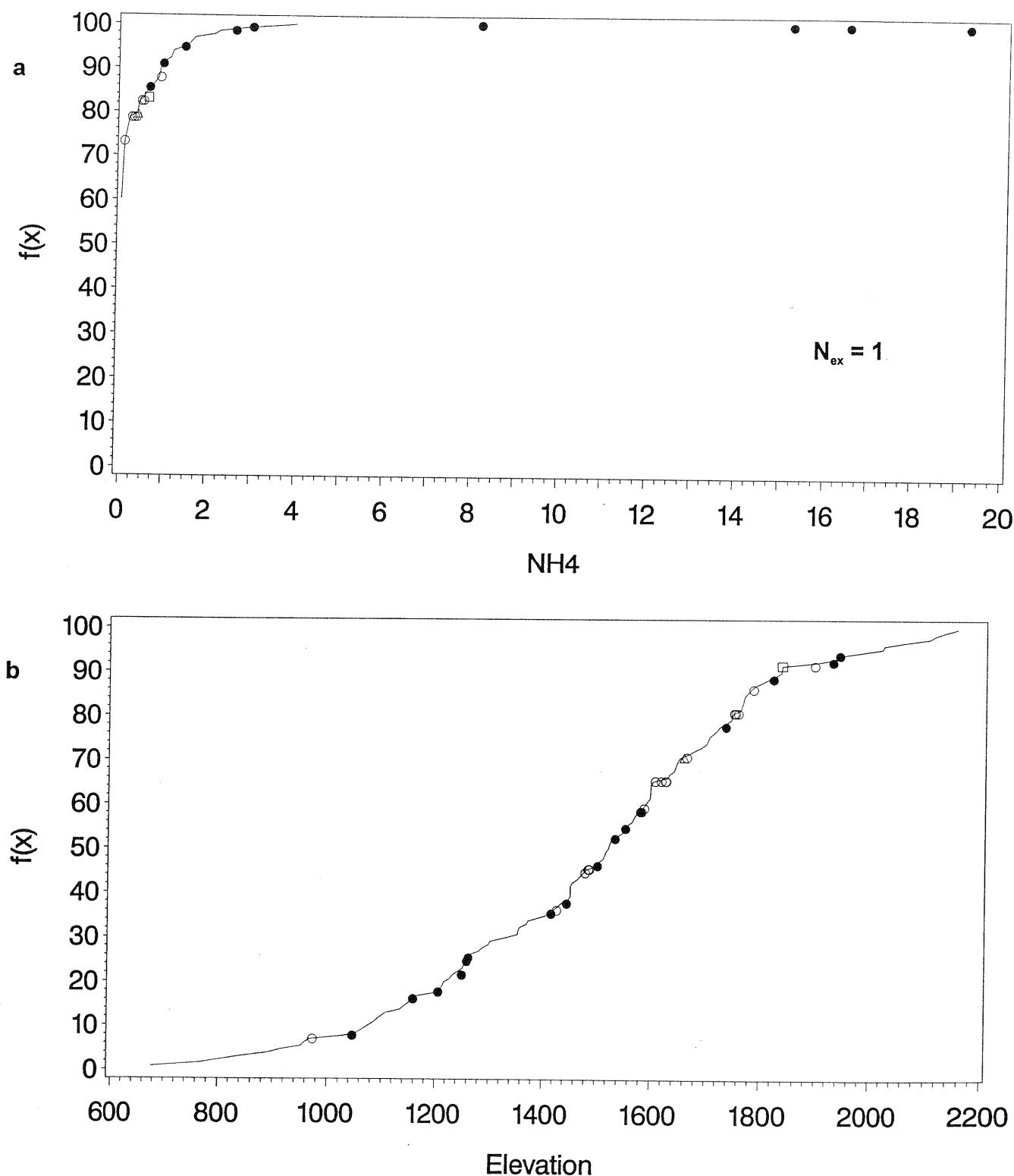


Figure 7. The distribution of Cascade diatom calibration set lakes for a) ammonium ($\mu\text{eq/L}$); b) elevation (m).

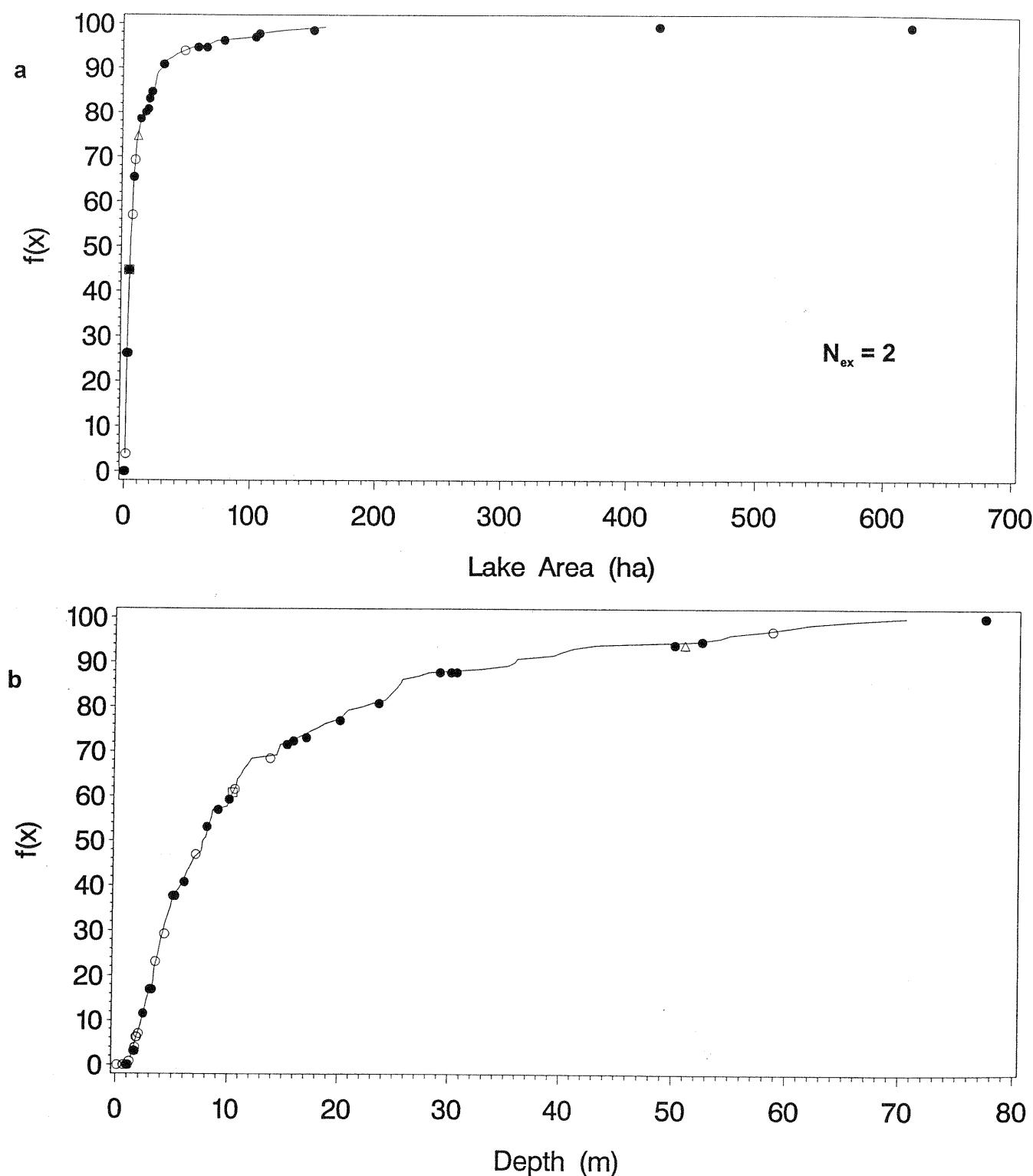


Figure 8. The distribution of Cascade diatom calibration set lakes for a) lake area (ha); b) depth (m).

2. Field and Analytical Methods (1996)

Twenty seven lakes were sampled during the summer and fall of 1996 (Table 1). From each lake, a short sediment core (< 20 cm) was collected from the deeper areas in the lakes using a mini-Glew corer. The cores were extruded on site and a section of sediment was removed from the top 0.5 cm of the core for analysis of diatoms. A 1-L water sample was collected from 0.5 m below the surface and placed in a Nalgene bottle and stored on ice. Four aliquots were collected from each lake for analysis of major ions, phosphorus, chlorophyll a, and phytoplankton. The phosphorus aliquot was preserved with sulfuric acid, the chlorophyll was preserved with magnesium bicarbonate, and the phytoplankton was preserved with Lugol's solution. Phytoplankton samples were archived for possible future analysis. Water samples were shipped via overnight courier to the Rocky Mountain Forest and Range Experiment Station laboratory in Fort Collins, CO for analysis of acid-base chemistry. Water samples were analyzed for pH, ANC, conductivity, calcium, magnesium, sodium, potassium, ammonium, sulfate, nitrate, chloride, total phosphorus, and chlorophyll a. Analytical methods are summarized in Table 2. Separate aliquots were sent to Aquatic Analysts in Portland, OR for analyses of chlorophyll a and phosphorus. The sediment samples were sent to the University of Louisville for analysis of diatom taxonomy. The sediment samples were placed on slides and digested to facilitate counting and identification of diatoms. The

Table 2. Analytical methods for analysis of lake samples collected in 1996 for the diatom calibration set.

Analyte	Method Description	Laboratory
pH	ARAS	USDA - Fort Collins
ANC	ARAS, Gran titration	USDA - Fort Collins
Ca, Mg, Na, K	Atomic absorption	USDA - Fort Collins
SO ₄ , Cl, NO ₃	Ion chromatography (Dionex)	USDA - Fort Collins
Conductivity	Specific conductance meter	USDA - Fort Collins
Total Phosphorus	Ascorbic Acid	Aquatic Analysts
Chlorophyll a	Trichromatic	Aquatic Analysts

The treatment of the diatom data followed the protocols established in the PIRLA program (Charles and Whitehead 1988, Charles and Smol 1990), in which 500 individuals were counted per slide.

The data were entered into a data base and calibration equations were developed for pH and other related variables using weighted averaging calibration techniques as described below.

3. Diatom Taxonomic Identification

Laboratory procedures for diatoms generally followed those used in the PIRLA project (Whitehead et al. 1990). Surface sediment material (typically the top 0.5 cm) was homogenized by gentle working and at least a 1 cc sample extracted for analysis. The organic material was removed by digestion in a strong acid with a catalyst, such as nitric acid and potassium dichromate. The remaining material was almost completely made up of the diatoms' siliceous cell walls. The resulting solution was quantitatively, randomly settled on cover slips using Battarbee trays (Battarbee 1973), and subsequently mounted in Hyrax. Five hundred diatom valves were identified to the lowest possible taxonomic level at 1000X.

For this project, surface sediments from 27 Cascade lakes were collected and the diatoms enumerated by P.R. Sweets. In addition, slides and counts were obtained from an earlier project on lakes from Mt. Rainier conducted by M.C. Whiting as described above. The diatoms from 10 of these lakes were also counted by P.R. Sweets with assistance from K. Manoylova. Nine counts were imported directly from Whiting's data. Two more Northwest lakes (Summit [Eilers et al. 1996a] and Natasha [Eilers et al. 1996b]) were added from previous projects performed by Sweets and Eilers. The melding of these two data sets was performed to enlarge the available calibration data set and to increase the ranges of important environmental parameters.

More than 450 diatom taxa (Appendix 1) were identified from the total calibration set. This large data set was reduced for ordination and modeling. Typically rare taxa are removed, or taxonomic uncertainties are eliminated by 'lumping' some groups of taxa. This latter procedure was done in some cases due to the combining of the Sweets and Whiting datasets. For the Cascades, a minor compression of the data set was performed by choosing important taxa using several criteria:

if the taxon occurred in at least 10% of the sample lakes (5-6) at a frequency of 1% or higher; if the taxon had a maximum percentage in any lake of 10% or greater; and if the taxon occurred in any percentage in 15% of the sampled lakes (10). Taxa fitting any of these three criteria were always admitted to the analytical taxon list (Appendix 2). Each taxon was then examined individually for its occurrence to ensure important taxa were included in the analytical dataset. An example of how a taxon might be introduced into the analytical dataset after the initial screening might be: small taxonomic splits (varieties of species) may result in a low individual frequency in any one lake. However, these species nearly always occur together. In this case, the varieties may be combined to make up a 'lumped' taxon and admitted to the dataset. Or, a lumping procedure may be done purely on the basis of taxonomy or uncertainty, perhaps due to the combination of counts from two individuals.

Many taxonomic references are used, but among the most important are Patrick and Reimer (1966, 1975), Camburn et al. (1986), the series of Krammer and Lange-Bertalot (e.g. 1986), and the many works of F. Hustedt (e.g., 1939). A photographic record is collected of the known and unknown diatom species, digitized, and used for subsequent identifications.

For the Cascade diatom calibration set, taxonomic issues are particularly important, as previous work on these lakes is not described in the literature. A large number of unknown species were identified from the 48 lakes, and taxonomic work is still required to resolve several taxonomic issues. Indeed, more than 60 of the 450 taxa (13.3%) are listed as 'unknown' species. However, in the context of an ordination and modeling project, the crucial taxonomic criterion is that taxa are consistently identified. In most cases, therefore, the fact that taxa are previously undescribed does not affect the resulting statistical tests and models, so long as the taxonomic naming convention is internally consistent. In Appendices 1 and 2, the unknowns are identified by the project in which they were first observed. The many taxa identified as, e.g., '*Cymbella* sp. 6', were first delineated during the PIRLA project (Charles et al. 1986). Another common form is '*Achnanthes* sp. PRS 0A project: Cascades', a form identified by P.R. Sweets during the Cascades project.

When identifications from two different taxonomists at different times are mixed, unknown taxa may become more of a problem. Whiting and Sweets have worked together in the past, however, and in general, these problems are minimized in this project. For example, many of the unknown taxa were described during the PIRLA project and pictures of the taxa in question were reproduced in Camburn et al. (1986). Both Whiting and Sweets were active in this project, so most of these taxa are uniformly identified.

However, there are a subset of unknown taxa seen by Whiting that exist in the lakes from which his counts were directly imported into the dataset (these are typically identified as the highest PIRLA numbered unknowns). For these, the original descriptions were not available, and the slides themselves were only located recently. Further work on harmonizing these unknown taxa or replacement of the Whiting counts with Sweets counts may improve the dataset. The individual cases are discussed in the next section.

4. Inference of Water Quality Characteristics Using Weighted Averaging (WA)

Weighted averaging regression and calibration, robust, computationally-simple, and straightforward methods for reconstructing environmental variables were used to develop calibration equations. They provide more accurate and precise inference than methods based on ecological categories (Birks et al. 1990). This approach is effective primarily because it uses information provided by each individual taxon, and not just abundance-weighted mean, \hat{u}_k , estimated as:

$$\hat{u}_k = \frac{\sum_{i=1}^n y_{ik}x_i}{\sum_{i=1}^n y_{ik}} \quad (1)$$

The fundamental assumption of the WA technique is that the weighted average of a taxon represents the conditions for which this taxon is most abundant. This 'optimum' condition for each taxon can be calculated as the average of mean values for the environmental characteristics (e.g., water chemistry) at the sites in which it is found, weighted by the abundance of the taxon at the sites, namely:

$$\hat{t} = \sqrt{\sum_{j=1}^n y_{ik} (x_j - \hat{u}_k)^2 / \sum_{j=1}^n y_{ik}} \quad (2)$$

Once WA values for an environmental characteristic (e.g., water pH) were calculated for taxa in a calibration data set, the information was used to infer that characteristic from lake samples and consequently to reconstruct past conditions. The first step was to determine the percent abundance of each taxon in the sample assemblages. The taxon abundance was then multiplied by the WA value for that taxon (determined from the calibration data set). These products are summed for all taxa and are standardized by the sum of the relative abundances of the taxa in that sample to obtain an inferred value, namely:

$$\hat{x}_i = \frac{\sum_{k=1}^m y_{ik} \hat{u}_k}{\sum_{k=1}^m y_{ik}} \quad (3)$$

where \hat{x}_i is the water chemistry characteristic being inferred, and m is the number of taxa in the sample assemblage.

An option we also employed in using the WA technique is 'tolerance weighting.' The rationale for this approach is that taxa occurring over a narrow range of an environmental gradient should be better indicators than taxa with broader tolerances. Consequently, taxa with narrower tolerances should be weighted more heavily in the WA calculations. A tolerance-weighted estimate was calculated as:

$$\hat{x}_i = \left(\sum_{k=1}^m y_{ik} \hat{u}_k / t_k^2 \right) / \left(\sum_{k=1}^m y_{ik} / t_k^2 \right) \quad (4)$$

Tolerance weighting has not always worked as well as expected in diatom reconstructions for lakes (Birks et al. 1990), perhaps because calibration data sets have not been large enough to provide sufficiently accurate estimates of tolerance values.

Weighted averaging without tolerance correction has generally proven superior for diatom reconstructions of several environmental variables (Birks et al. 1990, Kingston and Birks 1990), whereas tolerance correction may be superior for chrysophyte data (B.F. Cumming and H.J.B. Birks, personal communication); this may be explained by the higher diversity and greater number of 'zero occurrences' in the diatom data.

Averages are taken twice, once in the regression step and once in the calibration steps. This results in shrinkage of the environmental gradient, which is corrected by a linear deshrinking regression. To minimize this effect, a simple 'classical' or an 'inverse' regression (sometimes called a 'deshrinking' step) can be used (see ter Braak and van Dam 1989 and Birks et al. 1990 for more detailed discussion). For example, the classical deshrinking process performs a simple linear regression to determine the relationship between inferred values (x_{inf}) and actual measured values (x_{meas}):

$$\text{initial } x_{\text{inf}} = a + bx_{\text{meas}} + \epsilon \quad (5)$$

where a is the intercept, b is the slope and ϵ is an error term. The terms from this regression equation are then used to calculate a final 'corrected' or 'deshrunk' value:

$$\text{final } x_{\text{inf}} = (\text{initial } x_{\text{inf}} - a)/b. \quad (6)$$

Inverse deshrinking involves the same process, except that the measured values are regressed on the initial inferred values instead of vice versa. Classical deshrinking moves inferred values farther from the mean than inverse deshrinking, and the former is best if values to be inferred lie near the ends of the environmental gradient. Inverse deshrinking minimizes the root mean square error of the predicted versus measured regression relationships, and therefore may lead to more accurate inferred values over the entire range of the environmental gradient.

In summary, the process of developing and applying a diatom calibration set for the Cascades involved six distinct steps as illustrated schematically in Figure 9.

Process of Developing and Applying a Diatom Calibration Set

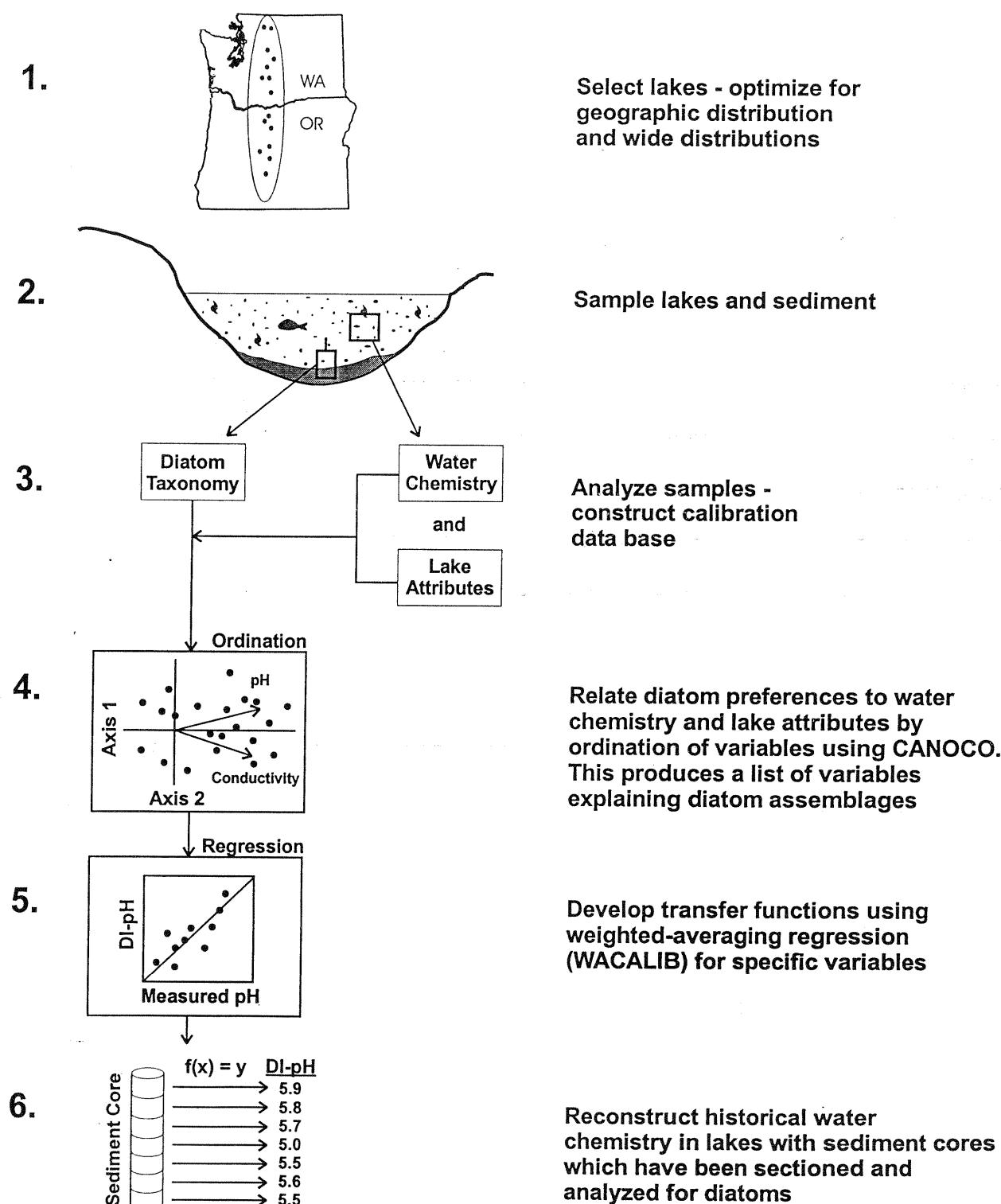


Figure 9. Schematic representation of the process of developing and applying the diatom calibration set for the Cascade Mountain Ecoregion.

5. Diatom Inferences

A standardized technique for quantitative diatom paleolimnological inferences has been established, based largely on research by C.J.F. ter Braak & H.J.B. Birks (e.g., ter Braak & Prentice 1988, ter Braak & van Dam 1989, Birks et al. 1990), and the availability of the computer programs CANOCO (ter Braak 1987b), CALIBRATE (Juggins & ter Braak 1992), and WACALIB 3.1 (Line et al. 1994). Weighted averaging regression and calibration has proven to be an effective technique for explaining the variance in complex taxonomically-rich data sets.

A series of exploratory ordinations of the Cascade calibration lake data was performed using correspondence analysis and canonical correspondence analysis. These analyses use eigenvector techniques to determine which environmental variables explain the greatest amounts of variance within the species assemblages (ter Braak 1987b). Variables that are significantly related to diatom species variance are considered to be good candidates for inferences studies. All variables except pH were \log_{10} transformed for the correspondence analyses.

The entire Cascade calibration set presented challenges in conducting this analysis because of an incomplete data matrix and slight differences in taxonomy for the Mount Rainier lakes. Some of these challenges were presented by melding the 27 Cascade lakes sampled in 1996 with the previously sampled lakes from Mount Rainier National Park (referred to here as the 'Rainier' subset). For instance, taxonomy on the Rainier lakes by Dr. Mark Whiting had to be reconciled with taxonomy on the lakes sampled in 1996 (see Table 1). Of particular importance in conducting the correspondence analyses, however, was the incomplete nature of the environmental data available (particularly with the Rainier lakes). Accurate CA analyses require a full data set without missing values. Therefore, different combinations of lakes were used in the numerous types of analyses performed, depending on the data available. In particular, two main subgroups were used:

Calset A:

34 lakes: eliminates lakes 29-40, 45-47

most environmental variables available: area, depth, conductivity, pH, sulfate, chloride, ANC, total phosphorus, magnesium, calcium, potassium

Calset B:

42 lakes: eliminates lakes 29, 30, 33, 40, 45, 46, 47

sulfate and chloride values unavailable

synthetic conductivity values added for three lakes: 32, 34, 39 (based on pH:conductivity in similar lakes)

Other changes in lake subgroupings are noted individually.

CA maximizes the dispersion or variance obtainable in the diatom data among the sites along a theoretical ('latent') environmental variable. The resulting eigenvalues for axis 1 (λ_1), axis 2 (λ_2), etc., and the total of all 'unconstrained' eigenvalues (λ_{unc}) are measures of the 'explainable' variance in the diatom calibration set. CCA is essentially a restricted correspondence analysis, selecting the linear combination of environmental variables that maximizes the dispersion of the species scores for the first CCA axis. Successive axes are created under the same restraint, but these additional axes also must be uncorrelated to the previous axes. The number of axes is restricted only by the number of available environmental variables, but typically only the first two axes describe meaningful trends in the species assemblages. The eigenvalues calculated for each CCA axis will generally be smaller than that of CA, as the dispersion of diatom taxa is restricted by the variables. A total of all canonical eigenvalues is then calculated (λ_{can}).

Therefore, a comparison of total and axis eigenvalues obtained by CCA with CA (e.g., CCA λ_1 /CA λ_1) is a measure of how well the environmental variables explained the variance in the taxonomic data. A further test of the statistical model is to run a 'constrained' CCA where only one variable is fitted to the first axis. Comparisons of λ_1 can then indicate the amount of variance explained by that individual variable.

D. RESULTS

1. Chemistry and Morphometry of Study Lakes

The chemistry of the individual study lakes (Table 3) span considerable ranges in pH (4.95 to 8.44), ANC (-15 to 7226 µeq/L), conductivity (2.7 to 554 µS), and total phosphorus (1 to 170 µg/L). Two lakes (East and Paulina Lakes) are particularly alkaline and removal of these outliers reduces the ranges of ANC and conductivity considerably, although the ranges in pH and total phosphorus are unaffected by their removal. The upper distribution in total phosphorus is represented by Agency Lake; the next highest total phosphorus value is only 32 µg/L. The chemistry of these systems ranges from precipitation-dominated lakes such as Natasha and Summit (WA) to lakes such as Paulina and East which receive major geothermal inputs.

The distribution for pH is nearly normal with a skewness of only 0.66 (Table 4). Most other distributions are highly skewed. The Rainier subset of lakes exhibits a much more restricted range of chemistry compared to non-Rainier lakes. Consequently, the distributions for the lake chemistry among the Rainier lakes exhibit a lower degree of skewness.

The physical properties of the study lakes also exhibit a wide range (Tables 5 and 6). The study sites range from small, shallow ponds that refill with every storm event to moderately large, deep lakes that have hydraulic residence times of multiple decades. Most of the study lakes occurred at elevations of about 1400 to 1700 m (~ 4600 to 5600 ft). The depths of the lakes varies considerably (0.1 to 77 m) with a central tendency of 8 to 14 m.

Table 3. Water chemistry for calibration lakes.

Lake	pH	ANC ^a	Cond ^b µS µeq/L	TP ^c µg/L	Chloro ^d µg/L	Ca ^e µg/L	Mg ^f µg/L	Na ^g µg/L	K ^h µg/L	SO ₄ ⁱ µeq/L	NO ₃ ^j µeq/L	NH ₄ ^k µeq/L	Cl ^l µeq/L
A. Newly Sampled Lakes													
Marion Lake	8.35	425	39.3	32	11.2	187.9	113.3	125.4	30.2	3.2	< dl	15.24	10.89
Sphagnum	5.81	37	7.3	13	0.5	43.2	9.0	22.9	1.7	10.5	< dl	< dl	10.72
Nordrum	6.44	40	6.4	5	0.4	32.1	9.5	20.1	2.0	10.8	< dl	< dl	10.29
Caskey Lake	6.17	109	12.5	19	0.2	77.5	36.6	37.5	3.2	13.9	< dl	< dl	11.55
Dumbbell Lake	6.55	55	7.6	12	0.3	22.3	16.4	25.1	4.5	6.7	< dl	0.94	5.70
Shellrock Lake	5.69	39	5.9	13	0.2	15.6	15.6	21.6	3.3	7.1	< dl	< dl	6.23
Heather Lake	6.23	94	10.6	10	0.2	59.0	27.2	23.8	8.7	13.9	< dl	< dl	5.13
Pyramid Lake	6.78	839	76.8	5	1.3	753.9	132.4	54.2	11.5	41.1	< dl	< dl	5.39
Middle Thornton	6.36	40	5.1	3	0.3	26.1	4.7	7.8	3.6	8.4	< dl	< dl	2.45
No Name	6.03	43	6.6	12	1.7	12.2	10.6	30.4	6.4	8.3	< dl	< dl	13.08
Gertrude Lake	6.62	75	10.4	13.3	2.2	38.2	24.9	26.0	5.3	12.0	0.06	0.64	6.60
Cora Lake	6.36	228	20.3	1	6.5	176.6	38.7	62.8	2.4	19.6	< dl	1.44	8.32
Head Lake	5.93	17	3.4	21	2.1	4.9	6.5	11.1	1.1	2.1	< dl	< dl	6.02
Summit Lake, OR	5.94	15	4.0	27	1.8	1.8	3.6	12.7	4.8	1.3	< dl	< dl	9.05
Little Twin Lake	4.95	-7	45.0	11	0.1	193.3	90.3	117.6	5.5	319.6	< dl	2.96	10.48
Agency Lake	8.37	939	98.5	170	23.4	351.1	300.7	520.6	47.3	55.9	7.56	16.53	115.99
South Heavenly Twin	5.32	17	3.2	9	0.9	11.7	8.7	6.8	1.3	2.1	< dl	< dl	3.01
Lucky Lake	5.27	16	3.1	11	0.3	9.6	8.0	8.5	2.1	5.0	< dl	< dl	5.61
Chariton	5.87	58	5.9	8	0.3	26.6	16.9	21.0	3.2	1.5	< dl	< dl	5.14
Paulina	8.44	7226	553.8	30	0.3	1567.1	4107.7	2438.5	132.4	65.7	< dl	< dl	30.72
East Lake	7.90	2161	342.0	19	0.9	1398.4	1199.9	83.8	91.5	1367.6	< dl	< dl	10.89
Big Lake	5.97	28	3.7	7	0.2	10.3	13.0	12.2	2.5	4.2	< dl	< dl	6.91
Lava Lake	7.90	342	34.3	29	2	94.1	113.8	184.3	24.5	13.4	< dl	8.17	12.33
Mink Lake	6.11	23	3.8	5	0.3	10.3	7.2	13.9	1.2	2.6	< dl	< dl	7.00
Diamond Lake	8.20	361	33.7	28	5.6	129.5	95.6	161.2	26.3	3.3	< dl	19.24	10.42
Burnt Lake	6.79	199	25.1	3	0.8	174.2	36.9	54.1	1.7	30.3	< dl	2.58	10.38
Suttle Lake	8.33	543	46.8	31	15.7	247.5	185.0	151.9	29.4	4.2	< dl	30.94	13.37

Table 3. Continued

Lake	pH	ANC ^a	Cond ^b	TP ^c	Chloro ^d	Ca ^e	Mg ^f	Na ^g	K ^h	SO ₄ ⁱ	NO ₃ ^j	NH ₄ ^k	Cl ^l
		µeq/L	µS	µg/L	µg/L					µeq/L			
B. Previously Sampled Lakes													
Unnamed Lake3	6.21	-12	2.7										
Unnamed Lake4	5.72	8		27		10.5	18.9	0.0	2.6			0.02	0.50
Unnamed Lake	5.66	8	7.1										
Unnamed Lake	5.68	-15	5.1	8		0.5	4.4		2.6			0.03	0.89
Unnamed Lake5	5.57	24											
Unnamed Lake6	5.63	6	10			4.0	4.6	5.2	2.2			0.02	0.06
Unnamed Lake7	6.52	7	10			6.0	4.1	4.8	1.5			0.06	
Unnamed Lake8	5.77	28	5.7	9		8.0	7.8	10.4	1.0			0.02	0.06
Unnamed Lake9	6.62	56	10.4	7		30.4	13.5	28.7	2.0			0.00	0.22
Unnamed Lake10	7.11	47	5.0	13		44.9	18.3	29.1	24.6			0.02	0.44
Snow Lake	7.26	67	9.0	9		44.9	7.3	21.3	2.0			0.13	10.15
Tipsoo Lake	7.68	312	15.3										
Shadow Lake	6.72	49	5.1										
Reflection Lake B	6.69	82	11.0	9		28.9	10.2	23.9	2.6	3.0		0.02	11.14
Upper Palisades L	7.25	84	13.0	11		68.9	16.6	34.8	4.2	13.1		0.02	15.43
Sunrise Lake	6.92	6.0											
Mowich Lake	7.11	91	11.0	4		70.4	13.8	23.5	1.5	3.7		0.00	13.82
Green Lake	7.02	268	19.0	7		141.7	16.3	34.8	2.8	7.9		0.13	15.23
Reflection Lake A	6.29	19	5.9	3		9.0	4.0	13.0	1.5			0.02	
Summit Lake	5.52	1	3.3	7		1.8	6.6	3.6	11.8	0.8	8.3	0.01	10.89
Lake Natasha	6.12	14	2.7	1		1.2	4.9	6.0	4.1	1.5	2.3	< dl	0.61
													4.25

^a acid neutralizing capacity^b conductivity^c total phosphorus^d chlorophyll a^e calcium^f magnesium^g sodium^h potassiumⁱ sulfate^j nitrate^k ammonium^l chloride

Table 4. Summary statistics for calibration lakes and major subsets of study lakes.

	pH ^a	Cond ^b	ANC ^c	C _b ^d	C _a ^e	Mg ^f	Na ^g	K ^h	NH ₄ ⁱ	SO ₄ ^j	Cl ^k	NO ₃ ^l	TP ^m	Chloro ⁿ
	µS												µg/L	µg/L
All N=48	48	44	47	42	42	42	42	41	35	35	41	42	29	0.1
Min	4.95	2.7	-15	13	0.5	3.6	0.04	0.9	0	1.3	2.5	0	1	23.4
Max	8.44	554	7226	8246	1567	4108	2439	132	31	1368	116	7.6	170	0.9
Median	6.36	7.5	47	71	31	15	24	2.7	0.06	7.9	10.4	0	10	0.9
Mean	6.54	35.3	321	427	147	161	107	12	2.5	60	12.8	0.2	16.2	2.9
sd	0.91	95.9	1090	1324	330	652	379	25	6.4	234	18.6	1.2	25.8	5.3
Skewness	0.66	4.63	5.87	5.41	3.55	5.78	6.00	3.62	3.2	5.49	5.30	1.2	5.38	2.88
Cascade N=34	34	34	34	34	34	34	34	34	34	34	34	34	34	29
Min	4.95	2.7	-6.5	16.5	1.8	3.6	4.1	0.8	0	1.3	2.5	0	1	0.1
Max	8.44	554	7226	8246	1567	4108	2439	132	30.9	1368	116	7.6	170	23.4
Median	6.40	10.5	71	86	44	16	25	3.2	0	8.1	10.3	0	12	0.9
Mean	6.63	43.6	428	519	178	197	130	13.9	3.7	61	12.8	0.3	20.3	2.9
sd	0.98	108.0	1269	1460	361	722	419	27.6	7.7	237	18.9	1.5	31.4	5.3
Skewness	0.51	4.04	5.00	4.87	3.16	5.20	5.41	3.29	2.4	5.40	5.22	5.0	4.46	2.88
Rainier N=19	19	15	18	13	16	16	13	16	12	6	6	12	13	No Data
Min	5.57	2.7	-15	13	0.5	4	0.04	1.0	0.06	3.0	10.1	0	3	27
Max	7.68	19	312	196	142	19	35	25	0.9	13.1	15.4	0.1	9	9
Median	6.62	7.1	37	66	29	10.2	21	2.2	0.3	5.0	12.8	0.02	0.04	9.8
Mean	6.50	8.7	63	69	36	10.7	18	3.9	0.3	6.3	12.9	0.04	0.05	5.8
sd	0.67	4.5	89	55	40	5.7	12.2	6.3	0.2	3.8	2.2	0.05	0.05	5.8
Skewness	0.04	0.91	2.10	1.01	1.71	0.12	-0.04	3.50	1.37	1.38	0	1.9	2.82	
Non-Rainier	29	29	29	29	29	29	29	29	29	29	29	29	29	29
Min	4.95	2.7	-6.5	16.5	1.8	3.6	4.1	0.9	0	1.3	2.5	0	1	0.1
Max	8.44	594	7226	8246	1567	4108	2439	132	30.9	1368	116	7.6	170	23.4
Median	6.23	7.6	55	77	38	17	25	3.6	0	8.3	9.1	0	12	0.9
Mean	6.56	49	482	588	196	229	147	15.8	3.4	71	12.7	0.3	19.1	2.9
sd	1.04	116	1371	1574	388	779	452	29	7.5	256	20.5	1.4	30.5	5.3
Skewness	0.71	3.71	4.62	4.49	2.88	4.80	5.00	3.01	2.5	4.99	4.89	5.18	4.57	2.89

^a pH^b conductivity^c acid neutralizing capacity^d sum of base cations^e sum of acid anions^f magnesium^g sodium^h potassiumⁱ ammonium^j sulfate^k chloride^l nitrate^m total phosphorusⁿ chlorophyll a

Table 5. Location and physical characteristics of study lakes.

Lake	State	Wilderness	Latitude/ Longitude	Area (ha)	Elevation (m)	Depth (m)
A. Newly Sampled Lakes						
Agency Lake	OR		42 30 00 N 122 00 00 W	3763	1262	3.0
Big Lake	OR		44 22 10 N 121 52 10 W	77	1416	23.5
Burnt Lake	OR	Mt Hood	45 21 03 N 121 48 05 W	3	1250	8.0
Charlton	OR		43 44 43 N 121 57 51 W	63	1735	29.0
Diamond Lake	OR		43 11 02 N 122 09 57 W	1301	1580	15.8
East Lake	OR		43 43 04 N 121 12 53 W	423	1942	52.4
Head Lake	OR		44 49 07 N 121 47 43 W	3	1525 ^e	2.4
Lava Lake	OR		43 54 53 N 121 45 52 W	149	1445	10.0
Little Twin Lake	OR		43 13 41 N 122 35 47 W	3	1540	9.1
Lucky Lake	OR		43 54 26 N 121 47 49 W	20	1585 ^e	15.2
Marion Lake	OR	Mt. Jefferson	44 33 41 N 121 52 20 W	105	1259	50.0
Mink Lake	OR	Three Sisters	43 55 29 N 121 55 17 W	56	1534	17.0
Paulina	OR		43 42 47 N 121 16 26 W	620	1930	77.4
South Heavenly Twin	OR	Sky Lakes	42 31 01 N 122 11 41 W	2	1821	5.2
Summit Lake	OR		45 01 52 N 121 47 20 W	2.5	1280 ^e	1.0
Suttle Lake	OR		44 25 33 N 121 43 23 W	102	1047	20.0
Caskey Lake	WA		48 24 07 N 121 34 25 W	15	250 ^e	5.0
Cora Lake	WA		46 41 20 N 121 53 19 W	11	1159	15.8
Dumbbell Lake	WA	William O. Douglas	46 41 30 N 121 22 45 W	18	1553	3.0
Gertrude Lake	WA	Goat Rocks	46 23 29 N 121 23 57 W	3.3	1748	6.0
Heather Lake	WA	Henry Jackson	47 51 30 N 121 07 35 W	29	1206	30.0
Middle Thornton	WA	North Cascades	48 41 18 N 121 20 11 W	3.1	1450 ^e	30.5

Table 5. Continued

Lake	State	Wilderness	Latitude/ Longitude	Area (ha)	Elevation (m)	Depth (m)
Unnamed	WA	Indian Heaven	46 01 19 N 121 49 51 W	1.7	1260	1.7
Nordrum	WA	Alpine	47 33 23 N 121 26 14 W	17	1118 ^e	9.0
Pyramid Lake	WA	North Cascades	48 40 59 N 121 08 38 W	0.9	800 ^e	8.0
Shellrock Lake	WA	William O. Douglas	46 41 05 N 121 20 40 W	6	1502	3.2
Sphagnum	WA	Alpine	47 33 01 N 121 26 26 W	0.02	1180 ^e	1.0
B. Previously Sampled Lakes						
Lake Natasha	OR	Sky Lakes	42 34 05 N 122 12 08 W	2.70	1836	10.3
Green Lake	WA	Mt. Rainier NP	46 58 42 N 121 51 48 W	4.98	974	29.0
Mowich Lake	WA	Mt. Rainier NP	46 56 42 N 121 52 12 W	45.42	1502	58.6
Reflection Lake A	WA	Mt. Rainier NP	46 46 00 N 121 43 24 W	0.70	1487	3.5
Reflection Lake B	WA	Mt. Rainier NP	46 46 00 N 121 43 48 W	6.74	1479	10.0
Shadow Lake	WA	Mt. Rainier NP	46 55 06 N 121 39 36 W	0.89	1896	4.3
Snow Lake	WA	Mt. Rainier NP	46 45 48 N 121 41 36 W	2.39	1426	10.5
Summit Lake	WA	Clearwater	47 02 29 N 121 49 57 W	8.82	1658	50.9
Sunrise Lake	WA	Mt. Rainier NP	46 55 00 N 121 35 48 W	1.50	1750	7.0
Tipsoo Lake	WA	Mt. Rainier NP	46 52 54 N 121 31 36 W	1.45	1617	1.7
Unnamed Lake	WA	Mt. Rainier NP	46 55 06 N 121 50 30 W	0.11	1758	1.0
Unnamed Lake	WA	Mt. Rainier NP	46 46 42 N 121 43 00 W	0.21	1626	1.2
Unnamed Lake3	WA	Mt. Rainier NP	46 46 54 N 121 43 24 W	0.17	1663	1.8
Unnamed Lake4	WA	Mt. Rainier NP	46 46 48 N 121 43 00 W	0.05	1625	0.7
Unnamed Lake5	WA	Mt. Rainier NP	46 46 36 N 121 43 54 W	0.03	1606	0.1
Unnamed Lake6	WA	Mt. Rainier NP	46 46 36 N 121 43 42 W	0.19	1582	2.0
Unnamed Lake7	WA	Mt. Rainier NP	46 46 30 N 121 43 42 W	0.13	1586	1.9

Table 5. Continued

Lake	State	Wilderness	Latitude/ Longitude	Area (ha)	Elevation (m)	Depth (m)
Unnamed Lake8	WA	Mt. Rainier NP	46 46 00 N 121 43 36 W	0.12	1485	1.1
Unnamed Lake9	WA	Mt. Rainier NP	46 46 06 N 121 43 30 W	0.21	1487	1.6
Unnamed Lake10	WA	Mt. Rainier NP	46 46 06 N 121 44 06 W	0.04	1485	1.0
Upper Palisades L	WA	Mt. Rainier NP	46 57 06 N 121 35 12 W	0.78	1785	15.3

^e estimated from topographic maps

Table 6. Summary statistics for physical attributes of study lakes.

Data Set	Elevation (m)	Lake Area (ha)	Lake Depth (m)
All N =	38	48	48
Min	250	0.02	0.14
Max	1942	3763	77.4
Median	1514	3.0	8.0
Mean	1473	143	13.9
sd	309	574	17.5
Skewness	-1.50	3.75	1.95
Cascade N =	23	33	34
Min	250	0.02	-1.0
Max	1942	3763	77.4
Median	1465	11	10.4
Mean	1413	208	18.7
sd	342	686	19.0
Skewness	-1.15	4.76	1.54
Rainier N =	16	19	19
Min	974	0.03	0.14
Max	1896	45.4	58.6
Median	1586	0.46	22.2
Mean	1569	3.4	8.3
sd	191	10.0	13.8
Skewness	-1.42	4.25	2.94
Non-Rainier N =	29	28	29
Min	.250	0.02	1.0
Max	1942	3763	77.4
Median	1450	16	10.0
Mean	1409	243	17.7
sd	355	741	18.9
Skewness	-1.17	4.37	1.67

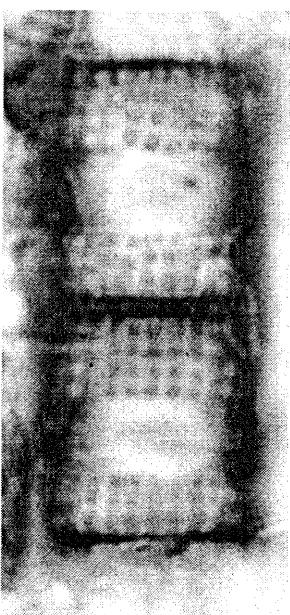


Figure 10.
Aulacoseira distans
from Big Lake, OR.

2. Diatom Taxonomy

The most widespread diatom in the data set is the centric *Aulacoseira distans* and its several varieties (Figure 10). It occurred in 40 lakes and was often observed at very high abundances in the sediments. The genus *Aulacoseira* is generally well represented in the Cascade lakes. The species *A. distans* is likely tycoplanktonic, rising into the water column as conditions permit, and resting in the sediments for long periods, often as resting spores. Its weighted average (or optimum) environmental tolerances in the Cascades (Table 7) indicates a species that prefers slightly acidic waters in mildly oligotrophic conditions.

Unfortunately, the *Aulacoseira distans* complex is also the most problematic taxonomic group in the Cascades. Although well known throughout the world, several new forms were seen in the Cascade counts in abundant numbers (*Aulacoseira* sp. 0A, 0B, 0C, 0D; and from the Whiting counts, *Melosira* sp. 4 and 8 PIRLA). This problem was exacerbated by an unclear conception of where the Whiting unknowns may fit in with the Sweets unknowns. Furthermore, *A. distans* itself was most abundant in the Rainier lakes, where it is often the dominant taxon (>30%). Upon examination of the Whiting slides, many of the *A. distans* varieties and the Whiting unknowns were combined in the analysis (*Aulacoseira distans* varieties), but the Sweets unknowns were retained. This group needs additional work, including electron microscopy analysis to fully appreciate the taxonomic differences in the Cascades. It is likely that the analysis might be improved with effort in resolving taxonomic uncertainties.

Another important diatom was *Navicula tenuicephala*. This taxon is one of the best-known strongly acidophilic diatoms in the world and was widely distributed in the Cascade lakes (36). It was present in the most acidic lakes, particularly the small, shallow lakes from Mt. Rainier, and Cascade lakes such as such as Big, Lucky, and Head. For that reason, it had a relatively low pH optimum (5.86).

Table 7. Occurrence and optima (abundance weighted mean) for taxa and taxa groups used in ordinations and inference models.

Genus	Species	Maximum % Abundance	Mean % Abundance	Sites	Optimum pH	Optimum Conductivity ($\mu\text{S}/\text{cm}$)	Optimum ANC ($\mu\text{eq}/\text{L}$)	Optimum Mg (mg/L)	Optimum Depth (m)	Optimum TP ($\mu\text{g}/\text{L}$)
Achnanthes	sp. PRS 0A	16.07	5.82	3.00	6.36	5.3	41	0.06	29.3	3.3
Achnanthes	sp. 11 PIRLA	16.93	5.59	7.00	7.15	9.8	100	0.11	23.1	7.3
Achnanthes	bioreti	5.36	1.20	15.00	5.99	9.8	26	0.65	12.0	7.6
Achnanthes	curtissima	5.00	1.95	5.00	6.53	12.4	107	0.27	13.7	8.8
Achnanthes	detha	4.82	2.53	5.00	7.28	11.0	196	0.10	15.1	8.0
Achnanthes	lanceolata	4.60	0.80	9.00	6.78	63.3	624	1.46	11.1	5.8
Achnanthes	linearis	4.00	1.62	8.00	6.56	34.7	324	0.86	16.5	11.1
Achnanthes	marginulata	28.16	5.58	44.00	6.15	6.7	42	0.14	13.2	13.3
Achnanthes	minutissima	6.45	1.62	14.00	6.71	18.6	255	0.35	21.3	8.4
Achnanthes	pusilla	8.39	2.10	5.00	7.13	12.1	123	0.16	50.3	4.4
Achnanthes	subatomoides	6.20	2.11	6.00	6.53	11.7	105	0.29	15.9	8.6
Achnanthes	suechlandti	6.21	3.25	2.00	7.09	11.0	90	0.16	56.4	4.2
Amphora	ovalis	1.00	0.44	10.00	7.31	49.8	527	1.46	13.4	11.8
Amphora	perpusilla	1.67	0.59	6.00	8.21	344.9	419	27.31	53.9	26.0
Anomoeoneis	brachysira	4.20	1.10	27.00	6.27	8.2	62	0.18	10.3	8.4
Anomoeoneis	vitrea	4.20	0.81	9.00	6.36	18.6	203	0.44	24.7	9.4
Asterionella	formosa	35.50	10.09	7.00	8.25	142.3	441	8.08	45.7	27.5
Aulacoseira	sp. PRS 0A	6.60	3.50	2.00	6.38	20.6	226	0.45	15.4	1.1
Aulacoseira	sp. PRS 0B	37.65	5.38	20.00	6.09	7.2	49	0.18	2.9	11.4
Aulacoseira	sp. PRS 0C	6.40	1.84	8.00	6.19	7.0	49	0.16	3.6	11.8
Aulacoseira	sp. PRS 0D	7.25	2.33	10.00	6.22	6.8	36	0.14	1.8	9.0
Aulacoseira	ambigua	9.00	2.94	7.00	6.66	22.3	201	0.65	7.7	10.9
Aulacoseira	distsans	57.67	17.45	42.00	6.12	7.4	34	0.12	6.8	10.9

Table 7. Continued.

Genus	Species	Maximum % Abundance	Mean % Abundance	Sites	Optimum pH	Optimum Conductivity ($\mu\text{S}/\text{cm}$)	Optimum ANC (meq/L)	Optimum Mg (mg/L)	Optimum Depth (m)	Optimum TP ($\mu\text{g/L}$)
Aulacoseira	granulata	2.80	0.97	9.00	7.11	24.4	249	0.70	12.8	12.3
Aulacoseira	italica	5.33	1.76	11.00	7.59	43.3	353	2.04	20.5	21.8
Aulacoseira	paffiana	65.33	9.60	16.00	5.52	22.8	17	0.53	5.3	13.0
Cocconeis	placentula	1.80	0.76	6.00	8.21	160.7	727	7.00	24.4	21.3
Cymbella	sp. PRS OA	28.03	4.97	22.00	5.90	4.4	18	0.09	5.7	11.8
Cymbella	sp. 18 PIRLA	3.48	1.27	5.00	6.81	5.9	48	0.09	6.7	8.6
Cymbella	sp. 6 PIRLA	16.00	5.50	10.00	5.61	4.1	22	0.09	6.6	15.3
Cymbella	aequalis	5.89	1.44	21.00	5.85	14.8	44	0.37	8.8	10.9
Cymbella	amphicephala	8.93	8.93	1.00	5.52	3.3		0.04	50.9	27.0
Cymbella	brehmii	6.60	1.15	15.00	6.06	4.8	25	0.11	11.2	8.6
Cymbella	falaisensis	7.66	2.77	12.00	5.82	8.2	26	0.13	3.5	11.1
Cymbella	gaeumannii	11.20	2.20	20.00	6.34	5.8	43	0.13	10.7	8.7
Cymbella	hebridica	3.20	0.98	24.00	5.87	7.2	34	0.14	5.9	13.0
Cymbella	lunata	5.86	1.35	23.00	6.50	7.9	72	0.18	9.5	11.5
Cymbella	minuta	13.02	1.51	18.00	7.23	12.8	217	0.25	10.5	10.1
Cymbella	perpusilla	2.19	1.46	3.00	6.46	8.1	39	0.11	2.4	5.5
Cyclotella	ocellata	6.40	3.40	2.00	6.23	11.1	101	0.32	29.2	9.5
Cyclotella	pseudostelligera	40.24	25.57	2.00	7.02	19.0	268	0.19	25.7	7.0
Cyclotella	stelligera	80.49	16.23	14.00	6.40	16.4	179	0.33	27.0	7.3
Eunotia	bilunaris	3.20	0.69	10.00	6.15	10.9	51	0.20	6.5	11.9
Eunotia	bilunaris	1.80	0.62	19.00	6.29	8.5	58	0.19	10.8	10.3
Eunotia	minor	4.47	1.36	8.00	5.97	6.4	50	0.10	35.1	14.6
Eunotia	paludosa	1.97	0.90	8.00	6.01	9.9	44	0.15	5.0	10.2

Table 7. Continued.

Genus	Species	Maximum % Abundance	Mean % Abundance	Sites	Optimum pH	Optimum Conductivity ($\mu\text{S}/\text{cm}$)	Optimum ANC (meq/L)	Optimum Mg (mg/L)	Optimum Depth (m)	Optimum TP ($\mu\text{g}/\text{L}$)
Eunotia	rhomboidea	4.27	0.65	11.00	5.75	4.6	29	0.07	33.4	20.9
Eunotia	subarcuatoidea	3.80	1.00	9.00	6.24	6.6	39	0.12	13.9	9.0
Eunotia	vanhurckii	3.37	0.93	5.00	6.31	6.2	38	0.09	24.5	4.0
Fragilaria	sp. PRS 0A	4.60	1.44	5.00	6.06	6.8	42	0.12	3.8	10.6
Fragilaria	sp. 11 PIRLA	2.17	1.38	3.00	7.23	13.2	207	0.11	5.7	9.0
Fragilaria	sp. 9 PIRLA	13.02	6.74	2.00	7.66	15.2	304	0.16	3.7	4.0
Fragilaria	brevistriata	33.40	9.64	19.00	7.28	105.3	339	7.02	25.2	12.4
Fragilaria	capucina	2.20	0.80	7.00	7.03	54.4	392	1.98	15.2	9.6
Fragilaria	crotontensis	34.40	10.46	7.00	8.23	104.2	480	5.07	32.1	24.3
Fragilaria	construens	28.60	8.84	17.00	7.32	37.2	361	1.35	10.0	13.9
Fragilaria	exigua	3.40	0.64	9.00	7.72	61.9	634	2.24	7.4	10.4
Fragilaria	pinnata	43.00	8.73	9.00	7.78	67.6	334	4.04	14.8	22.4
Fragilaria	pinnata	32.50	9.82	17.00	7.55	52.0	463	2.01	13.3	17.7
Fragilaria	pseudoconstruens	6.80	2.86	3.00	6.86	14.7	129	0.43	8.1	13.4
Frustulia	rhomboides	4.96	1.23	21.00	6.26	5.8	39	0.11	14.3	7.1
Frustulia	rhomboides	7.80	1.49	31.00	5.98	6.0	34	0.13	8.5	11.4
Gomphonema	angustatum	1.45	0.66	8.00	7.06	10.4	154	0.14	14.3	6.8
Gomphonema	gracile	1.40	0.39	14.00	6.21	18.8	149	0.51	13.5	8.9
Melosira	sp. 13 PIRLA	4.38	2.04	3.00	7.10	8.3	58	0.08	9.2	9.0
Navicula	sp. 0E	3.80	1.40	4.00	6.36	24.5	267	0.55	14.4	4.6
Navicula	sp. 20 PIRLA	3.53	1.41	4.00	7.16	8.6	82	0.08	12.3	9.0
Navicula	sp. 24 PIRLA	11.17	1.89	16.00	6.23	7.9	26	0.10	4.3	11.9
Navicula	sp. 25 PIRLA	1.69	0.63	7.00	7.00	9.8	96	0.12	18.4	7.5

Table 7. Continued.

Genus	Species	Maximum % Abundance	Mean % Abundance	Sites	Optimum pH	Optimum Conductivity ($\mu\text{S}/\text{cm}$)	Optimum ANC (meq/L)	Optimum Mg (mg/L)	Optimum Depth (m)	Optimum TP (ug/L)
Navicula	sp. 45 PIRLA	2.42	0.91	4.00	6.82	6.3	53	0.09	6.0	9.0
Navicula	cryptocephala	1.00	0.45	11.00	6.63	43.5	122	2.39	10.4	9.5
Navicula	cryptotenella	2.38	0.74	16.00	7.03	14.1	176	0.49	15.2	11.2
Navicula	laevissima	2.40	0.75	10.00	6.94	35.7	414	0.98	12.5	7.6
Navicula	leptostrata	5.40	1.66	21.00	5.93	6.2	38	0.15	6.4	13.5
Navicula	mediocris	10.80	1.08	27.00	6.18	7.1	51	0.16	11.1	8.3
Navicula	pupula	6.52	1.02	13.00	6.68	24.3	210	0.57	10.8	8.8
Navicula	radiosa	3.10	0.82	12.00	7.15	21.3	221	0.72	16.4	9.5
Navicula	seminuloides	5.40	1.51	7.00	6.46	16.8	168	0.58	8.5	14.7
Navicula	seminulum	2.60	0.78	11.00	6.67	37.6	244	1.54	10.8	12.3
Navicula	subatomoides	2.20	0.59	9.00	6.56	28.8	165	1.45	10.9	13.1
Navicula	submuralis	2.40	0.85	8.00	6.62	19.9	195	0.60	8.9	13.6
Navicula	subtilissima	16.00	1.97	27.00	5.71	5.1	19	0.11	9.4	11.5
Navicula	tenuicephala	49.11	15.93	36.00	5.86	5.8	25	0.12	6.5	13.0
Neidium	affine	3.48	0.86	12.00	6.41	6.3	45	0.13	9.7	12.9
Neidium	alpinum	1.76	0.64	18.00	6.13	6.4	35	0.11	6.5	10.9
Neidium	iridis	2.67	0.75	27.00	5.89	11.5	27	0.24	8.5	12.5
Nitzschia	fonticola	14.73	1.69	14.00	7.63	32.9	319	3.26	9.4	18.7
Nitzschia	frustulum	10.39	3.36	5.00	7.36	12.8	259	0.11	4.4	9.0
Nitzschia	gracilis	6.03	1.34	17.00	6.21	27.9	121	0.62	7.0	11.1
Nitzschia	perminuta	5.10	1.45	6.00	6.44	8.6	78	0.25	3.8	10.8
Nitzschia	tropica	1.57	0.56	5.00	7.90	289.3	460	15.77	43.9	21.1
Pinnularia	sp. PRS 0C	4.20	1.52	4.00	5.79	4.5	15	0.07	4.1	23.0

Table 7. Continued.

Genus	Species	Maximum % Abundance	Mean % Abundance	Sites	Optimum pH	Optimum Conductivity ($\mu\text{S}/\text{cm}$)	Optimum ANC ($\mu\text{eq/L}$)	Optimum Mg (mg/L)	Optimum Depth (m)	Optimum TP ($\mu\text{g/L}$)
Pinnularia	sp. 28 PIRLA	14.98	4.12	7.00	6.79	5.7	47	0.08	7.0	8.5
Pinnularia	biceps	13.83	3.37	24.00	6.09	6.5	33	0.11	7.8	12.0
Pinnularia	braunii	9.20	3.43	11.00	6.14	6.1	43	0.14	9.2	10.4
Pinnularia	brebissonii	6.69	3.89	2.00	6.23	3.2	14	0.07	9.8	8.0
Pinnularia	gibba	3.33	0.68	12.00	6.62	9.7	71	0.18	8.3	10.4
Pinnularia	subcapitata	5.24	5.24	1.00	5.52	3.3		0.04	50.9	27.0
Stenopterobia	intermedia	0.92	0.39	14.00	6.42	7.0	64	0.17	11.2	7.9
Stauroneis	anceps	3.40	0.92	17.00	6.57	7.8	59	0.18	7.7	11.4
Stephanodiscus	hantzschii	2.67	1.00	5.00	8.34	350.9	509	27.88	60.6	28.7
Stephanodiscus	medius	18.67	5.88	5.00	8.35	57.3	573	2.04	34.5	30.9
Stephanodiscus	minutus	56.71	15.76	8.00	8.16	387.7	516	26.30	57.7	24.2
Stephanodiscus	niagarae	6.80	2.40	4.00	8.15	36.2	384	1.29	14.8	21.4
Stephanodiscus	parvus	3.33	1.83	7.00	8.20	215.7	536	15.31	39.6	28.2
Surirella	delicatissima	3.00	0.78	11.00	5.85	4.7	26	0.09	7.4	16.6
Surirella	delicatissima	2.98	0.92	28.00	6.03	9.5	27	0.20	5.2	11.0
Tabellaria	spp.	0.80	0.32	11.00	6.13	9.4	82	0.23	11.5	9.1

^a conductivity^b acid neutralizing capacity^c magnesium^d total phosphorus

Achnanthes marginulata was widespread (37 lakes) and often occurred at very high percentages (e.g., 28% in Summit Lake). It also is most common in the small shallow Rainier lakes, perhaps because it is often found as an epiphyte. Environmental optima are similar to that of *Aulacoseira distans*. Because of some uncertainties in melding the two data sets, the percentages were combined with those of *Achnanthes levanderi*, but this combination, although not taxonomically precise, probably had little effect on the statistical analysis. The common planktonic diatom *Cyclotella stelligera* (Figure 11) occurred in only 14 lakes, but was abundant in these lakes. It appears to prefer relatively unproductive lakes that have sufficient area and depth to support a vigorous planktonic community such as the Cascade lakes Charlton (>80%) and Heather and the small but deep Green Lake from Mt. Rainier. Its pH optima is 6.41. *Stephanodiscus minutulus* is also planktonic, and like *C. stelligera* is found in a few lakes (8), occasionally at high abundances (over 50% in Paulina and East Lake). It is found in larger, deeper lakes of higher productivity, that are typically quite alkaline.

The many small *Fragilaria* taxa that make up the species *F. pinnata*, *F. construens*, and *F. brevistriata* also are very abundant in the Cascade lakes. Although very similar in appearance under the light microscope, some authorities now separate them into different genera. Nevertheless, they often occur together in differing numbers, and can dominate the surface sediment assemblages of alkaline lakes such as Lava, Caskey and Gertrude. A very different important diatom species is the well known eutrophic plankter *F. crotonensis* which dominated Suttle and Marion lakes. *F. crotonensis*, *Asterionella formosa*, and *S. parvus* were among the species with the highest TP optima.

A number of diatoms probably need further taxonomic effort to resolve their differences in the Cascades. Among these is a very complex group of *Cymbella* (commonly epiphytic) species of

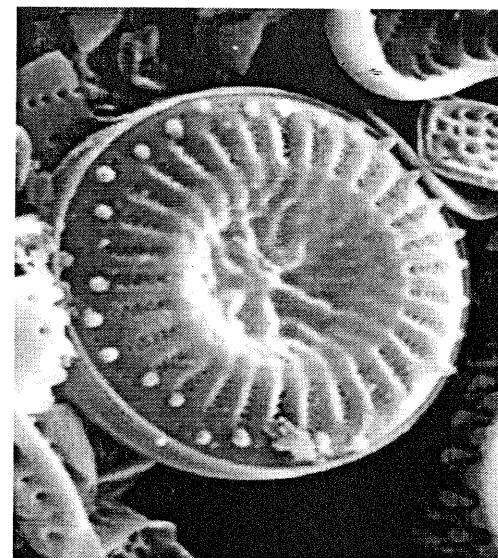


Figure 11. Electron micrograph of *Cyclotella stelligera*.

which the important and widespread taxa *Cymbella* sp. 6 PIRLA, *Cymbella* sp. 0A CASCades

(Figure 12), and *Cymbella brehmii* may all be closely related.

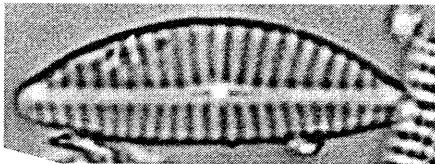


Figure 12. *Cymbella* sp. 6 from Lucky Lake, OR

The *Achnanthes* genus is also well represented in the Cascade calibration set and some careful taxonomic work, with electron microscopical analysis, could also benefit this difficult-to-identify group.

In summary, the Cascade diatom assemblages are typified by larger, more productive lakes being dominated by assorted planktonic species, and smaller, shallower lakes containing a diverse mix of epiphytic and benthic species. The resulting weighted average optima for the many well-known species are consistent with values found in the literature. The most abundant taxon, *Aulacoseira distans* and its varieties, is also the most complex and the most in need of further taxonomic study in the Cascade lakes.

3. Statistical Diatom Ordination and Regression

To assess the importance of the fullest possible suite of environmental variables, analyses were first run on the smaller calibration set, Calset A. For both transformed and untransformed data, the first axis was most strongly related to interrelated components of acid-base chemistry such as pH, ANC, conductivity, and the individual base cations. The second axis was most strongly related to lake depth. The first axis was significant as determined by a Monte Carlo permutations test (99 unrestricted permutations, $P \leq 0.05$). The CANOCO program also enables individual variables to undergo forward selection to ascertain their importance to the first axis of ordination. These tests indicated that the variables that best explain the diatom variance were one of the three major metrics of ionic constituency (pH, conductivity, and ANC) with one of the base cations also being important. Sulfate and chloride concentrations (the major variables missing in the Mt. Rainier lakes) were not strongly correlated to diatom assemblages. This was fortunate because it meant that subsequent statistical analysis of the diatoms could be conducted for those lakes in the data set

for which sulfate and chloride were not measured. It was decided that the larger calibration set of 42 lakes (Calset B) could be employed for more detailed analyses.

CANOCO also assists in determining correlations between environmental variables (Table 8). It is desirable to eliminate strongly correlated variables. Many variables may be candidates for diatom inferences, but reconstructions of highly correlated variables cannot be considered to be independent. Most of the available variables in the Cascade calibration set are related to major ion chemistry. Lake depth, area, and total phosphorus are not strongly correlated with other variables. It was decided in the second phase of calibration set ordination to eliminate all base cations except Mg (which was the most important in the transformed data), and to explore the three major acid-base variables (pH, ANC, conductivity) in detail. Water depth, area, and total phosphorus would also be retained.

The percent of theoretical variance that may be explained by the diatom assemblages in the 35 lake set by CA approaches 50%, with 12.9% ($\lambda_1 = 0.864$) ordinating along the first canonical axis (usually the only significant axis) and 23.5% on the first two axes combined (Table 9). This number compares favorably with other diatom calibration set investigations. More importantly, the CCA for the same lake set has a $\lambda_1 = 0.782$. The ratio of the first axis eigenvalues of CA and CCA implies that the measured environmental variables capture 90% of the theoretical variance along the first axis. This demonstrates that we have collected the most important environmental data for representing the diatom communities.

The amount of variance explained by the other axes declines abruptly from CA to CCA and as correlated variables are eliminated. However, it appears that although we captured the most important variables, minor variables that may not be represented could explain some of the variance in the diatom communities. Such variables may include metrics for transparency, morphometry, or other chemical attributes such as silica.

The total unconstrained eigenvalues (or inertia) sum to 6.674; this value is high and reflects the heterogeneity within the diatom data set. A measure of this is found by running CA on just the 27 Cascade lakes sampled for this study. The inertia falls to 5.342 and the variance explained by the

Table 8. Correlation matrix for 12 environmental variables, 35 lakes.

	Area	Depth	Cond ^a	pH	lgSO ₄ ^b	lgCl ^c	lgANC ^d	lgTP ^e	lgNa ^f	lgK ^g	lgCa ^h
Area	1.0000										
Depth	.5099	1.0000									
Cond	.5018	.3556	1.0000								
pH	.6084	.3719	.7368	1.0000							
lgSO ₄	.1779	.1900	.7345	.2243	1.0000						
lgCl	.4250	.0313	.5943	.5933	.3735	1.0000					
lgANC	.5617	.3830	.8764	.8573	.4536	.5280	1.0000				
lgTP	.3956	.1659	.2964	.3709	.0843	.5365	.2519	1.0000			
lgNa	.5729	.2492	.9044	.7407	.5411	.7311	.7999	.4223	1.0000		
lgK	.6424	.2740	.8621	.7721	.5167	.5044	.8224	.5319	.8355	1.0000	
lgCa	.4002	.4262	.9415	.6841	.7224	.5021	.8387	.0887	.8224	.7271	1.0000
lgMg	.6084	.3701	.9603	.7042	.6804	.5295	.8653	.3621	.9017	.8949	.8881
	Area	Depth	Cond	pH	lgSO ₄	lgCL	lgANC	lgTP	lgNa	lgK	lgCa

^a Conductivity^b log sulfate^c log chloride^d log acid neutralizing capacity^e log total phosphorus^f log sodium^g log potassium^h log calcium

Table 9. Eigenvectors calculated in CCA ordination analyses for Cascade Lake subset.				
Subset (variables)	λ_1	λ_2	λ_{can}	λ_{unc}
35 lakes, 12 vars. CA CCA	0.864 0.782	0.706 0.497	- 30.274	60.674 60.674
42 lakes, 7 vars. CCA	0.775	0.308	10.981	60.803
42 lakes, 5 vars. CA CCA (5 w/pH) - outliers CCA (5 w/cond) CCA (5 w/ANC)	0.877 0.775 0.764 0.771 0.751	0.709 0.291 0.305 0.294 0.301	- 10.709 10.739 10.563 10.589	60.762 60.803 60.803 60.803 60.803
Cascade Lakes (27) CA	0.881	0.621	-	50.342
single vars., 42 lakes CCA - pH CCA - conductivity CCA - ANC CCA - Mg CCA - TP CCA - depth	0.652 0.701 0.660 0.713 0.256 0.431	0.707 0.716 0.709 0.721 0.843 0.748	- - - - - -	- - - - - -

first axis accordingly increases to over 20% with the exclusion of the Rainier lakes. Therefore, the melding of the two data sets (Cascade and Rainier) is a trade-off of greater sample size versus the increased heterogeneity of the merged data. This test merely reflects the differences in diatom assemblages and does not take into account the different physico-chemical status of the two lake sets.

Exploration of important environmental variables

The exploration of the datasets continued by running CA and CCA analyses on different sets of transformed variables using the 42 lake set Calset B described above. First, the eigenvalues and correlations were examined with seven environmental variables (all three major ionic variables) and then substituting just one of either pH, conductivity, or ANC in a run with six environmental variables

(pH/ANC/conductivity, total phosphorus, depth, area). The eigenvalues (Table 9) are very similar for the first axis in all the CCA analyses (0.751-0.775) explaining a little more than 11% of the diatom variance, and still capturing a significant amount of the first axis theoretical variance.

In each of these cases, the variables most closely related to the first axis are Mg and one of the three major variables (pH/ANC/conductivity). Depth is most strongly correlated to the second axis. The importance of these five variables can be further tested by constraining the individual variable to the first axis of ordination — in effect, the vector for the variable becomes the first axis and can be tested for its power to explain the variance in the diatom assemblages. In particular, important variables should demonstrate a ratio of close to 1 for $\lambda_1:\lambda_2$. Table 9 demonstrates that this criterion is met for all three of the ionic summary variables as well as magnesium, each of these explaining as much as 10% of the variance.

The CANOCO program also identifies those lakes that have an undue influence on the ordination. These lakes are typically identified from a casual review of the environmental variables. For instance, East and Paulina Lakes have extreme values of conductivity and ANC and Agency Lake has a very high total phosphorus value (Table 3). Eliminating these extreme observations does not greatly affect the CCA results (see Table 9), but the outliers can be very important in assessing the standard errors of any lake reconstruction derived from these calibrations.

A final biplot of both site scores (Figure 13) and sample scores (Figure 14) with the vectors of the retained environmental variables demonstrates the strength of the ionic constituents in describing the first axis of ordination. In particular, the Cascade lakes are well dispersed along the first axis, providing the basic line along which the diatoms differentiate. The Rainier lakes are more concentrated in the low pH (and conductivity and ANC) end of the continuum and are distributed across the poorly defined second axis of ordination. The Rainier lakes also include many of the 40 most important species to a lower abundance weighted mean of the target variables.

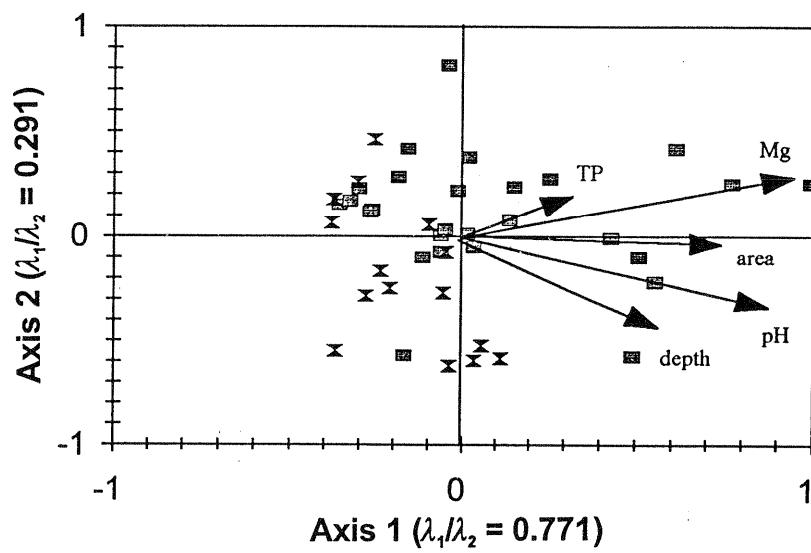


Figure 13. CCA ordination diagrams of sediment diatoms. Biplot of environmental variables with site scores are presented as linear combinations of environmental variables. Cascade lakes are squares and Rainier lakes are double triangles.

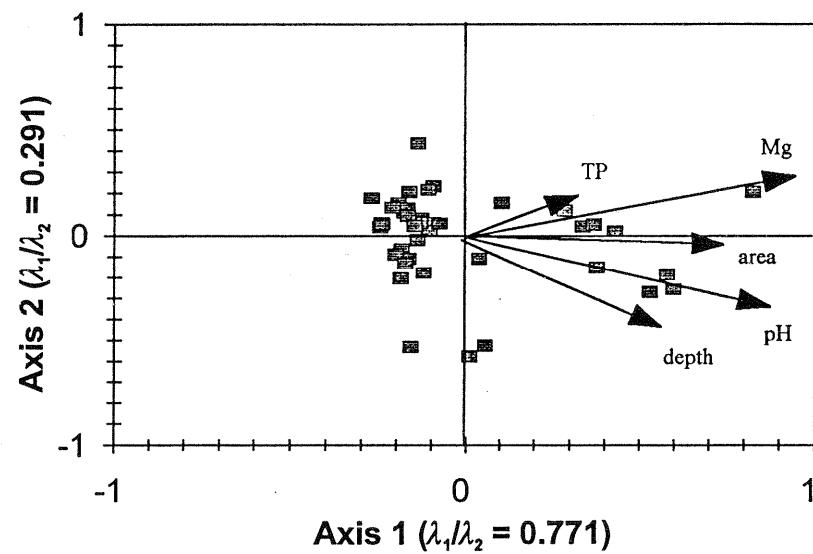


Figure 14. CCA ordination of species scores with environmental variables for Cascade lakes.

Choosing variables as candidates for inference equations

Transfer functions were derived for pH. The computer program WACALIB 3.3 calculates transfer functions using WA regression and calibration. A coefficient is assigned to each diatom based on its optimum (weighted average mean pH, etc.) within the calibration set. The final model may be based on either a simple weighted average (WA) or weighted average with a tolerance or range factor built in (WA-tol). Comparison of the inferred versus observed values will give the absolute residual and what is now referred to as the apparent r^2 and apparent standard error (RMSE). Bootstrapping techniques can be used to eliminate the circularity that can affect these statistics. Bootstrapping randomly selects with replacement a subset of lakes to be used as the calibration set and the remainder make up the test set (Birks et al. 1990). By running numerous (e.g., 1000) bootstrap cycles, an error estimation can be determined eliminating the previous circularity. Bootstrapping error is referred to as the RMSEP_{boot} or the standard error of prediction using bootstrapping. In an actual water quality historical reconstruction for a lake, an individual prediction error can be calculated for each modern sample and an estimated prediction error derived for each inferred sample.

Furthermore, outliers can be identified as observations whose absolute residual (observed-inferred) is greater than the standard deviation (Jones & Juggins 1995). The calibration model was run on a series of environmental variables and the resulting inference statistics are summarized in Table 10. Although the table contains statistics from log-transformed reconstructions, in fact such reconstructions can be difficult to use and evaluate. Therefore, we will primarily discuss the statistics based on untransformed values.

pH. Inference statistics for pH (based on 48 lakes) indicate a high coefficient of determination ($r^2 = 0.84$), with tolerable standard errors (apparent $s=0.36$, RMSEP_{boot} = 0.52). Statistics less amenable for use in inference equations often will show a large difference between the two types of standard errors, and the standard errors is well within the range of the metric standard deviation. A graph of the observed versus diatom-inferred pH (Figure 15) shows that most values are closely inferred. Two lakes from the subset of the calibration set had very high absolute residuals

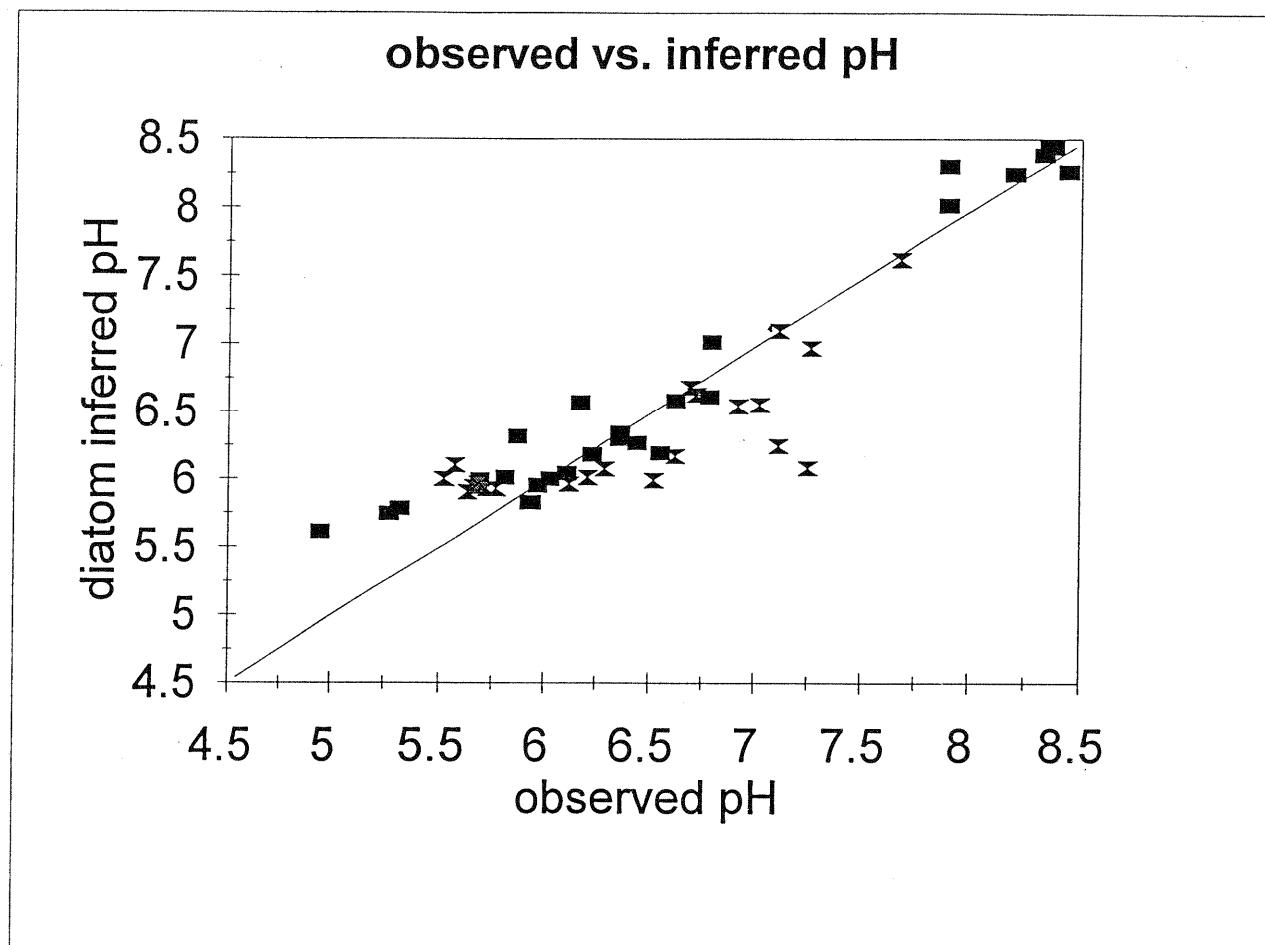


Figure 15. Measured pH versus diatom-inferred pH for all lakes in the Cascade diatom calibration set. Cascade lakes are squares and Rainier lakes are double triangles.

Table 10: Summary of inference statistics from weighted averaging regression of selected environmental variables against the diatom assemblages. Calculated using WACALIB and CALIBRATE

Environmental Variable	WA		WA-tol		Range of metric		RMSE bootstrap	
	r ²	RMSE	r ²	RMSE	Min /Max	sd	WA	WA-tol
ANC	0.661	635	0.859	409	-15/7226	1079	1118	992
ANC (-extreme)	0.668	123	0.601	134	-15/939	207	166	185
log (ANC +16)	0.765	0.32	0.801	0.30	0/3.86	0.66	0.39	0.41
conductivity	0.817	41.0	0.933	24.8	2.7/553.8	94.8	80.8	84.3
log conductivity	0.851	0.20	0.861	0.20	.43/2.74	0.52	0.30	0.33
depth	0.712	9.4	0.746	8.8	0.1/77	17.3	14.8	16.6
log depth	0.571	0.40	0.641	0.37	-1/1.9	0.61	.516	0.54
magnesium	0.657	4.5	0.938	1.9	0.4/48	7.5	8.2	7.5
log magnesium	0.861	0.25	0.877	0.24	-1.4/1.7	0.67	.37	0.39
total phosphorus	0.355	20.6	0.390	20.0	1/170	25.3	31.2	26.7
TP (-outliers)	0.530	5.7	0.620	5.1	1/32	8.20	7.8	7.5
log TP	0.455	0.27	0.621	0.23	0/2.23	0.37	.402	0.35
pH	0.786	0.42	0.845	0.36	4.9/8.4	0.90	.54	0.52
pH (-outliers)	0.837	0.31	0.887	0.31	4.9/8.4	0.90	.48	0.46

(Figure 16) and were removed from the model for a second run. The removal of these outliers has a moderate effect on the inference statistics ($r^2 = 0.89$, $s=0.31$; Table 10) that may not be readily apparent from Figure 17. This demonstrates that the removal of some sites can improve a diatom calibration model if used with caution. Such outliers probably arise most frequently when non-measured environmental variables are controlling diatom populations. In summary, pH is an excellent candidate for inference studies in the Cascades.

The diatom-inferred pH results and plot of residuals, although acceptable on statistical grounds, raise several questions that warrant further examination: (1) What is the effect of the high pH lakes on the predictive capability of the equation? (2) What is the effect of the Rainier lakes? and (3) What is the effect of the lakes for which reconstructions are being prepared? Historically, the primary motivation for creating a diatom-inferred pH calibration has been to assess changes in lake chemistry associated with acidification from atmospheric deposition (e.g., Charles and Smol 1988), although other applications for pH reconstructions are now becoming recognized (Smol 1992).

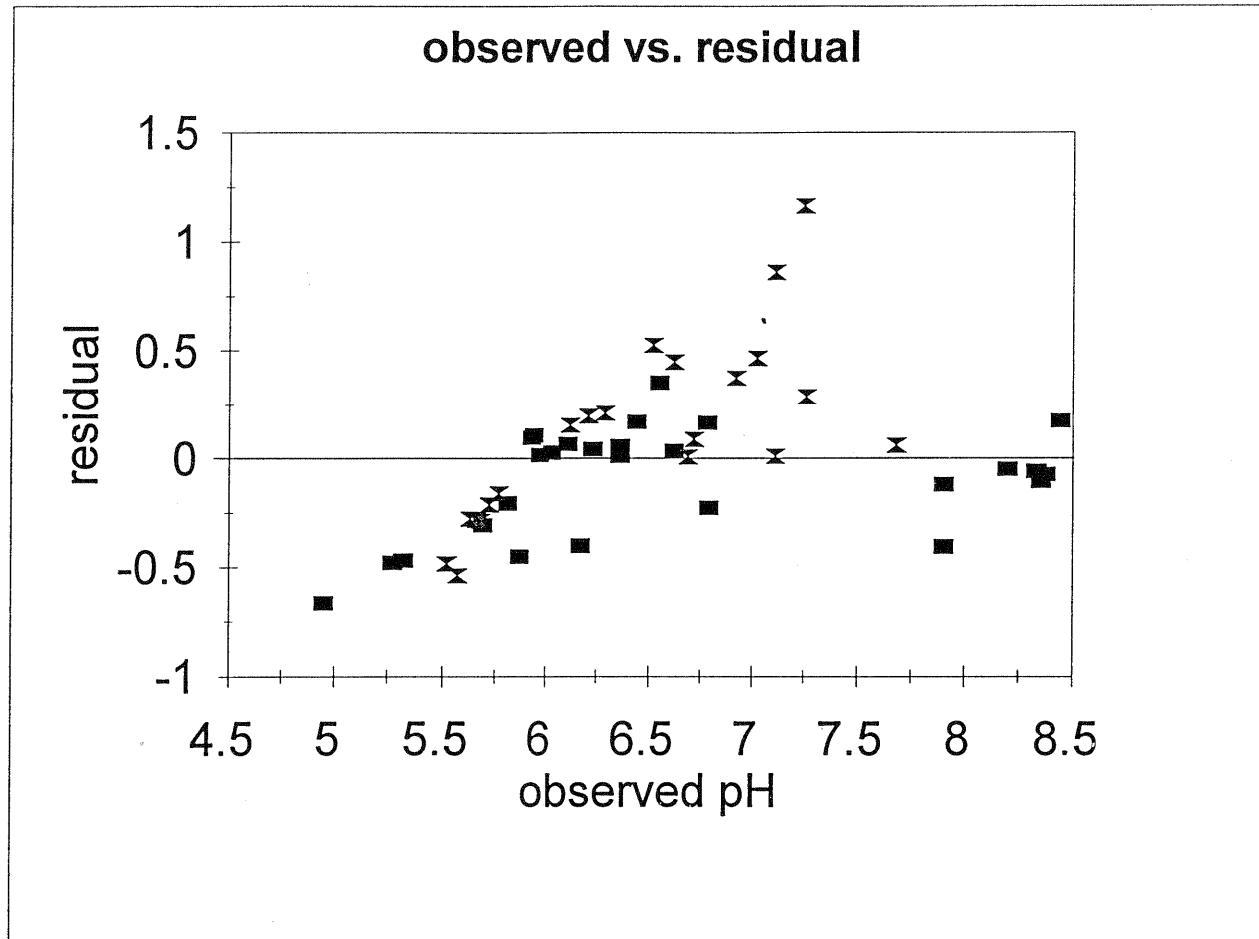


Figure 16. Measured pH versus the residual (from Figure 15) of measured and diatom-inferred pH. Cascade lakes are squares and Rainier lakes are double triangles.

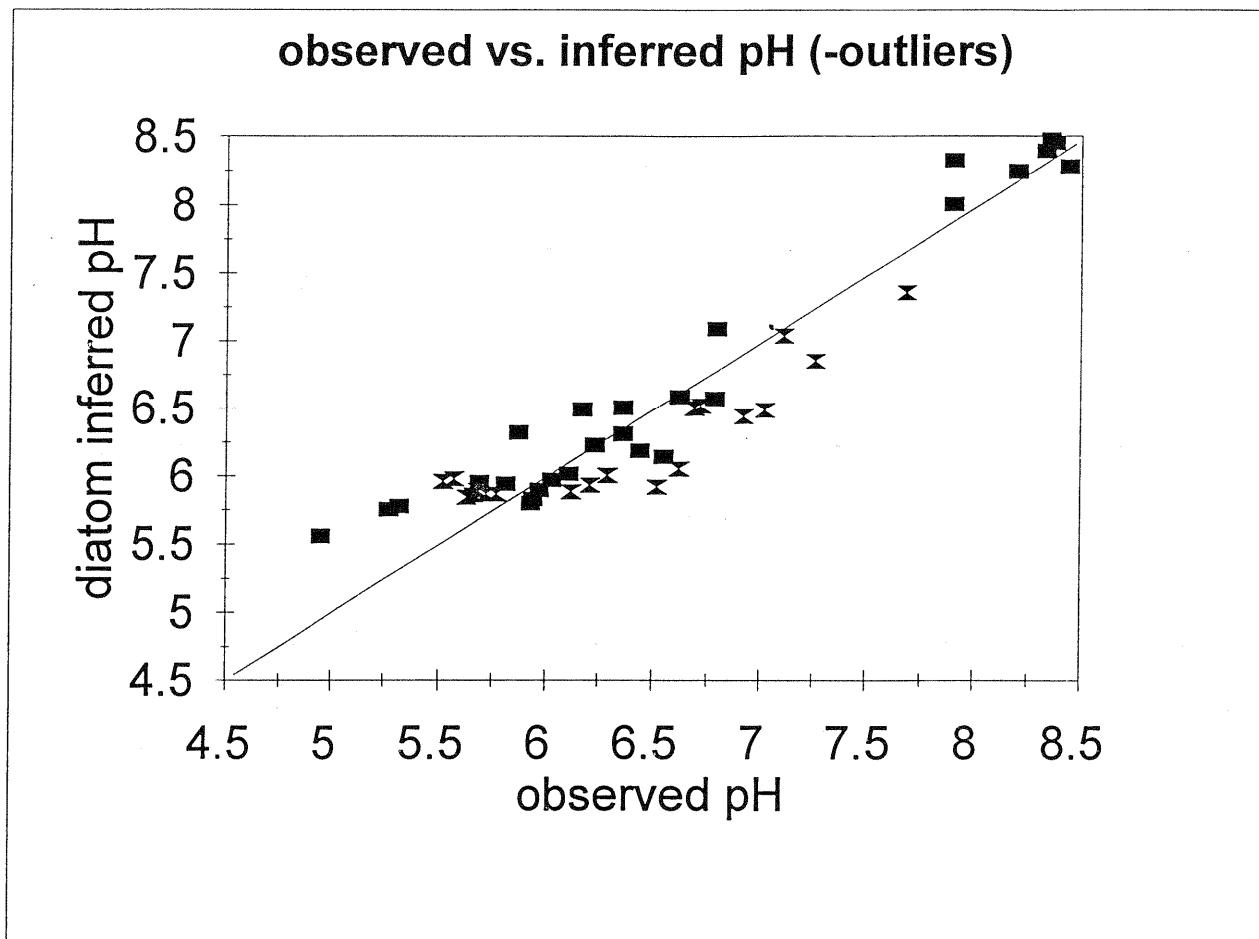


Figure 17. Measured pH versus diatom-inferred pH with two outliers removed. Cascade lakes are squares and Rainier lakes are double triangles.

If the objective of creating a pH reconstruction is to analyze potential acidification issues in low pH lakes, then there is no need or advantage in including high pH lakes in the calibration set because high pH lakes are not susceptible to acidification. To assess the effect of the high pH lakes on the calibration equation, high pH lakes were successively removed from the calibration set starting with lakes with pH greater than 7.5. The resulting calibration set of 38 lakes yields an equation with higher variance ($r^2=0.76$) and lower standard error ($s=0.27$) (Figure 18). The calibration set was further restricted to lakes with pH less than 6.5 (n=26 lakes) to yield an equation with even greater variance ($r^2=0.58$) and lower standard error of prediction ($s=0.24$) (Figure 19).

The plot of pH residuals illustrates that the Rainier subset of lakes predicts somewhat differently than the non-Rainier (i.e. Cascade subset) lakes (Figure 16). The effect of the Rainier lakes was evaluated by removing these lakes from the calibration set and re-running the weighted averaging (with tolerance) equation (Figure 20). The results indicate that removing the Rainier lakes results in reduced unexplained variance ($r^2=0.94$) and a lower standard error of prediction ($s=0.25$). The subset of Cascade lakes (n=29) may yield improved results with the Rainier lakes excluded because of several factors including (1) exclusion of the very shallow Rainier ponds which are dominated by benthic diatom taxa; (2) consistent taxonomy of the Cascade subset, or (3) more consistent water chemistry data in the Cascade subset.

Finally, there was concern that Summit and Notasha Lakes which will utilize the pH calibration results in the pH reconstructions unduly influences the calibration equations. The effect that these two lakes exert on the process was evaluated by re-running the Cascade calibration subset shown above, but this time excluding Summit and Notasha Lakes. The results show a very minimal effect of these two lakes on the calibration equation ($r^2=0.95$, $s=0.24$). Subsequent analyses for other parameters in the remainder of this report report the results with both Summit and Notasha Lakes included in the calibration equations. The effect of the various permutations of lake subsets on the pH calibration statistics are presented in Table 11.

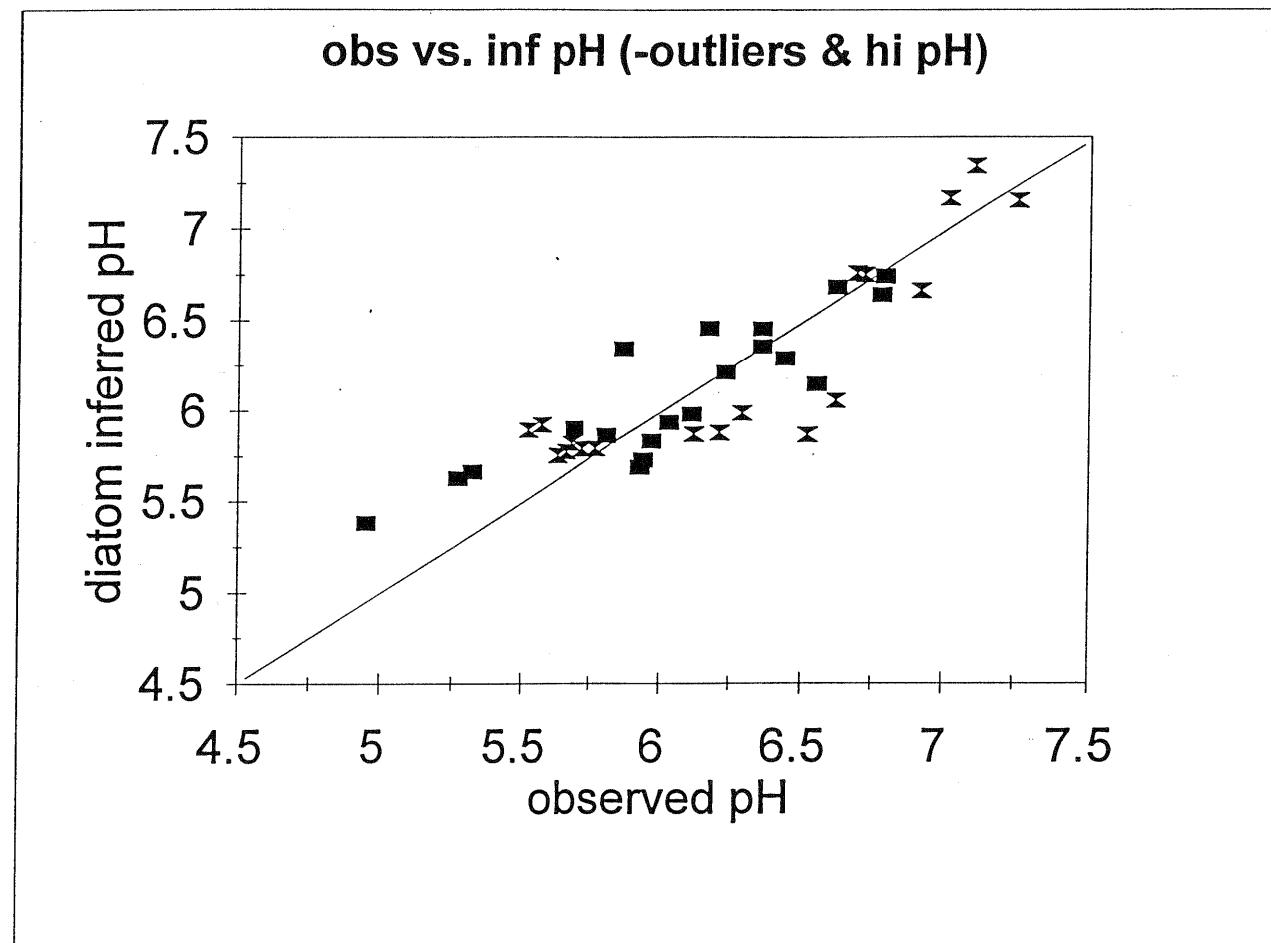


Figure 18. Measured pH versus diatom-inferred pH for lakes with measured pH less than 7.5. Cascade lakes are squares and Rainier lakes are double triangles. Two outliers described in the previous figure are also omitted.

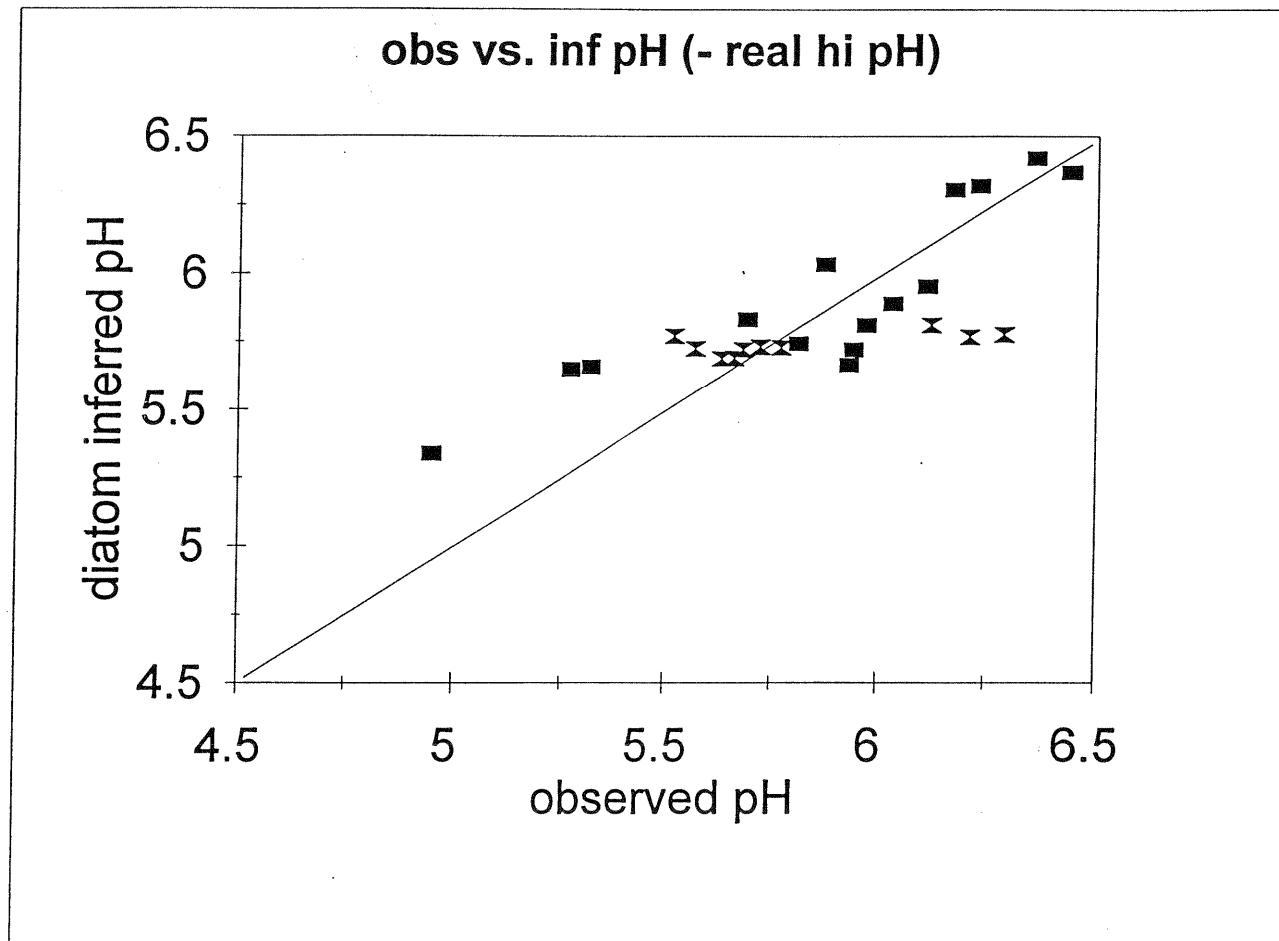


Figure 19. Measured pH versus diatom-inferred pH for lakes with measured pH less than 6.5. Cascade lakes are squares and Rainier lakes are double triangles. Two outliers described in the previous figure are also omitted.

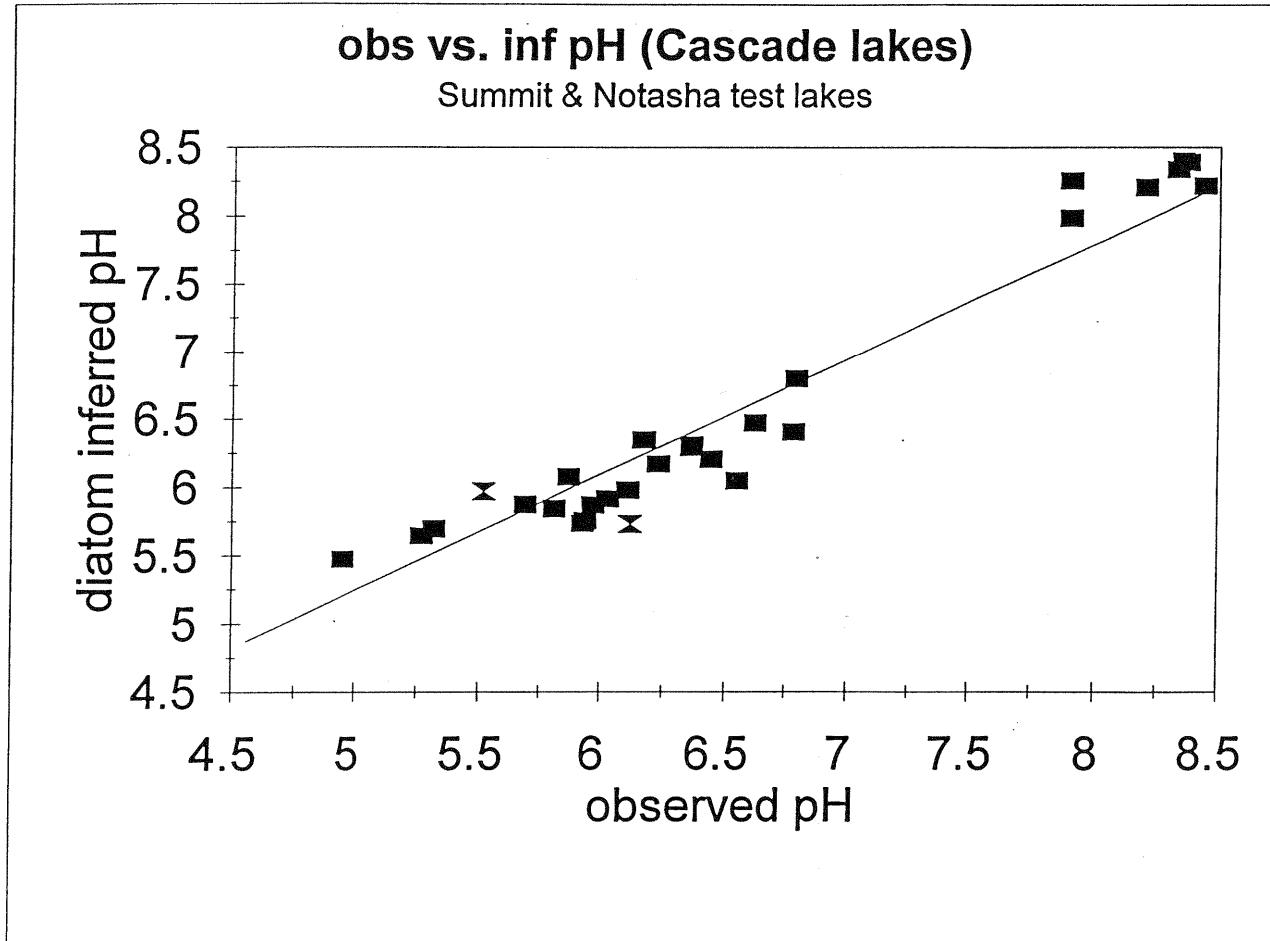


Figure 20. Measured pH versus diatom-inferred pH for lakes in the Cascade-only subset (i.e., all Rainier lakes omitted). Cascade lakes are squares. Summit and Natasha Lakes are plotted on the figure as double triangles, but are not included in the calibration equation.

Table 11. pH calibration regression statistics (weighted-averaging, with tolerance) for the Cascade calibration set.

	Population Subset					
	All	All-outliers ^a	pH < 7.5	pH < 6.5	Non-Rainier	Non-Rainier - S & N ^b
N	48	46	38	26	29	27
r ²	0.84	0.89	0.76	0.58	0.94	0.95
se	0.36	0.31	0.27	0.24	0.25	0.24

^a excludes Unnamed Mt. Rainier Lake (Park Service code ln23, our code RAIN38) and Upper Paradise Lake (Park Service code ln14, our code RAIN28)
^b excludes Summit and Natasha Lakes

Conductivity. Inference statistics for specific conductance indicate that this variable is a strong candidate for inference using diatom assemblages. Using a suite of 44 lakes, WA(tol) equations have a very high r² (0.93), and a high standard error (24.8). However, RMSEP_{boot} values are significantly higher (84.3) than the WA(tol) results. It is likely that the two lakes with extreme values for conductivity (East and Paulina Lakes are greater than 300 µS whereas all others are generally below 100 µS) may artificially inflate these statistics. However, the graph of observed versus inferred values demonstrates close agreement with the 1:1 line (Figure 21). The initial conductivity inference equation based on all available data yields a high r² value, but the standard error of the prediction is also high. The standard error from this equation is too large to have any predictive value. The weighted-averaging regression was re-run first excluding the two lakes with conductivity greater than 100 µS and again excluding the two lakes with conductivity between 50 and 100 µS. The resulting regression statistics illustrate the improvement in the smaller standard errors (Table 12). The improvement in the regression statistics comes at the price of a reduced range in which diatom inferences can be made, but a conductivity range of 0 to 50 µS still includes approximately 85% of the lakes in the study area (Eilers et al. 1987).

Acid Neutralizing Capacity (ANC). The possibility of reconstructing ANC is somewhat more problematic. Initially based on all lakes, r² of the WA(tol) inferred versus observed is high (0.86), but the bootstrap values are very high — greater than the standard deviation of the range of observed

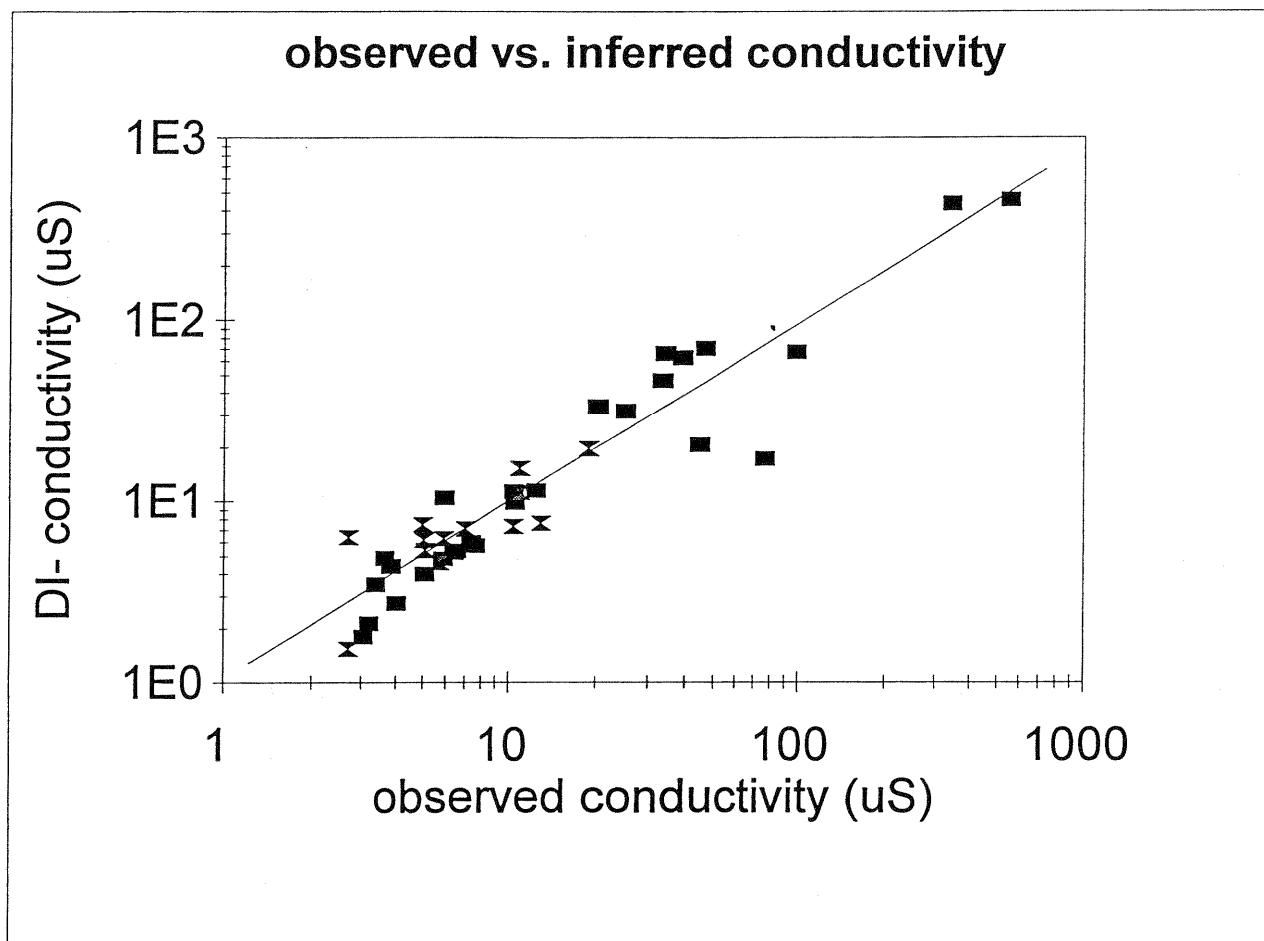


Figure 21. Measured conductivity versus diatom-inferred (DI) conductivity for lakes in the Cascade diatom calibration set. Cascade lakes are squares and Rainier lakes are double triangles.

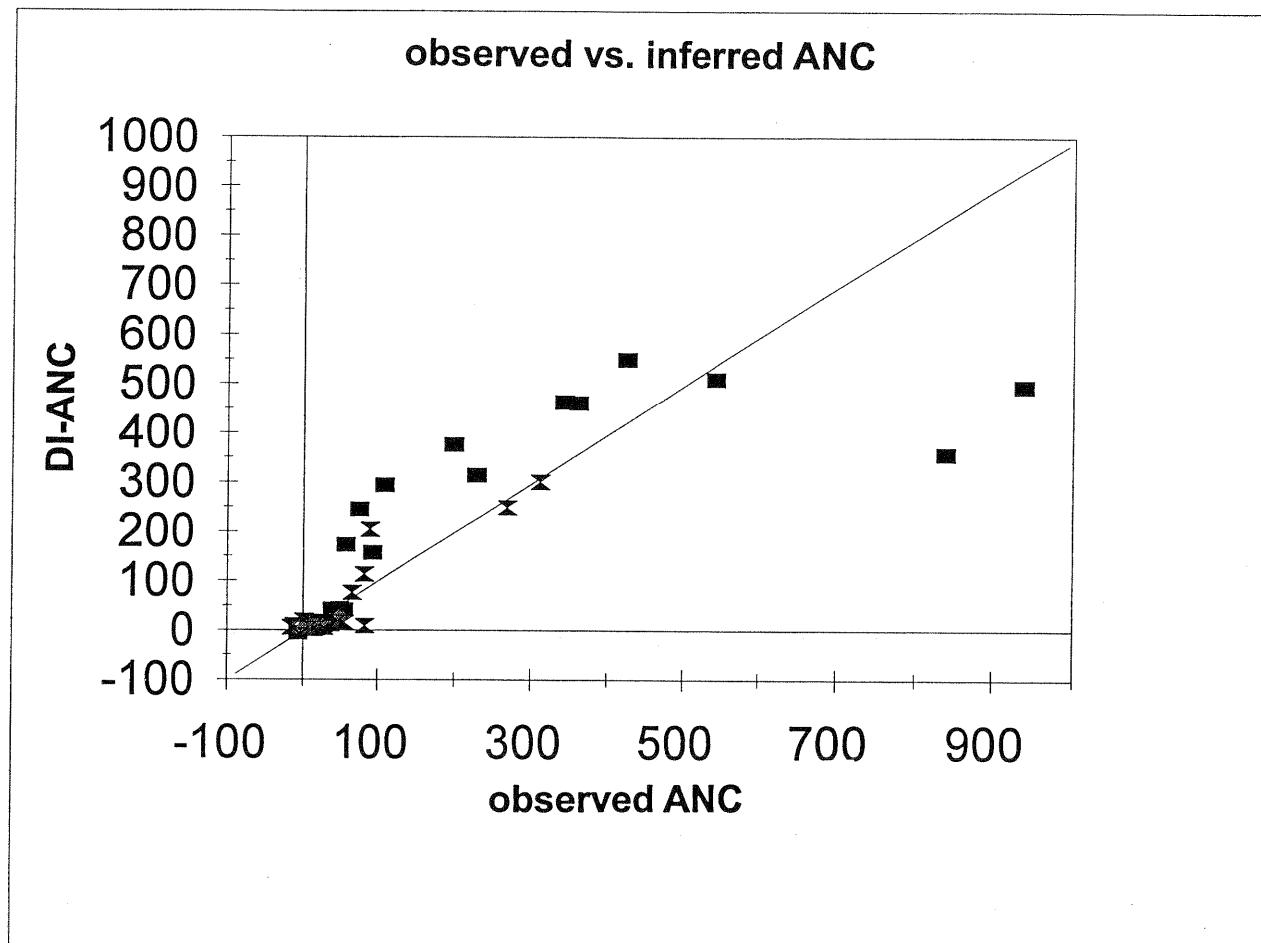
Table 12. Weighted-averaging regression statistics for diatom-inferred conductivity (μS) in the Cascade calibration set.

Conductivity Range	2.7-553	2.7-98	2.7-49
N	44	42	40
r^2	0.933	0.61	0.83
RMSE	24.8	12.5	4.9
RMSE _{bootstrap}	84.3	17.0	7.9

values. There are two extreme ANC values in the dataset (East and Paulina Lakes are an order of magnitude above other values). As the inference equation is judged by a regression line between observed and inferred, such disparate values can generate spurious r^2 values by drawing a line between two groups of points that are widely separated. The log transformations can assist with these problems, but ANC values can have negative values, and obtaining a normal distribution of points through transformation required adding a constant to each value before converting to log values. This obscures the object of this analysis, which is determining how well diatom assemblages can predict measured ANC values that scientists can easily interpret.

We removed the extreme untransformed values and again plotted the observed values against diatom-inferred values (Figure 22). The resulting r^2 (0.67 for WA) is lower than observed for WA(tol), but the equation is improved when considering the standard error. Unfortunately, a standard error of 123 $\mu\text{eq/L}$ is too great to provide useful predictions. It is possible that the removal of the next two extreme values which are also outliers with respect to poor prediction by diatoms may improve this statistic. However, ANC clearly is of limited use as a potential candidate for diatom inference equations in the Cascades.

Lake Depth. Lake depth was strongly related to the second axis of the CCA ordination. The r^2 of the WA(tol) equation is 0.75 and the standard error is 8.8 m. This is well within the range of the standard deviation of the observed values and represents around 10% of the range of lake depth. Unlike the above environmental values, water depth appears to be a weak independent variable in the diatom prediction. It is more likely that depth is inversely related to ionic strength or related



through altitude to ionic strength. Figure 23 demonstrates a large spread in the values with many outliers and a well-defined group of values representing shallow lakes.

Total Phosphorus (TP). There was no indication in either the CCA or the WACALIB analyses that total phosphorus is an important predictor of diatom variance. WACALIB calibration model results. The total phosphorus results from the 42 available lakes suggests that diatoms cannot be used to predict past TP concentrations with sufficient accuracy at this time (Table 10). However, utilizing the log TP or removing one extreme value (Agency Lake) improves the statistic considerably (r^2 of WA(tol) = 0.62, apparent $s=5.1 \mu\text{g/L}$). Figure 24 shows that the higher TP (e.g., TP > 15 $\mu\text{g/L}$) Cascade lakes do not predict well. Careful screening for outliers or resampling of some lakes may improve the inference equation, but at present, TP calibration results are not yet adequate to provide reliable diatom reconstructions for the Cascades.

4. Summit Lake Reconstruction

The calibration set was applied to a sediment core collected from Summit Lake, located in the Clearwater Wilderness of the Mt. Baker-Snoqualmie National Forest. Physical and chemical attributes of Summit Lake are summarized in Tables 13 and 14. Principal diatom taxa in Summit Lake include *Achnanthes marginulata*, *Aulacoseira distans*, and *Navicula tenuicephala* (Figure 25). Summit Lake is located in relatively close proximity to the Puget Sound urban/industrial corridor and there was some concern that the lake pH may have been reduced as a consequence of atmospheric deposition of sulfur. The reconstructed pH and conductivity for Summit Lake show no statistically significant change over the last 3150 years (Figure 26). Diatom-inferred pH ranged

Table 13. Morphometry of Summit Lake, WA (Source: Eilers et al. 1996a)

Parameter	Units	Metric	English
Elevation	m/ft	1658	5439
Surface Area	ha/ac	8.82	21.8
Total Volume	$\text{m}^3/\text{ac-ft}$	1.87×10^6	1521
Mean Depth	m/ft	21.2	69.6
Maximum Depth	m/ft	50.9	167.0

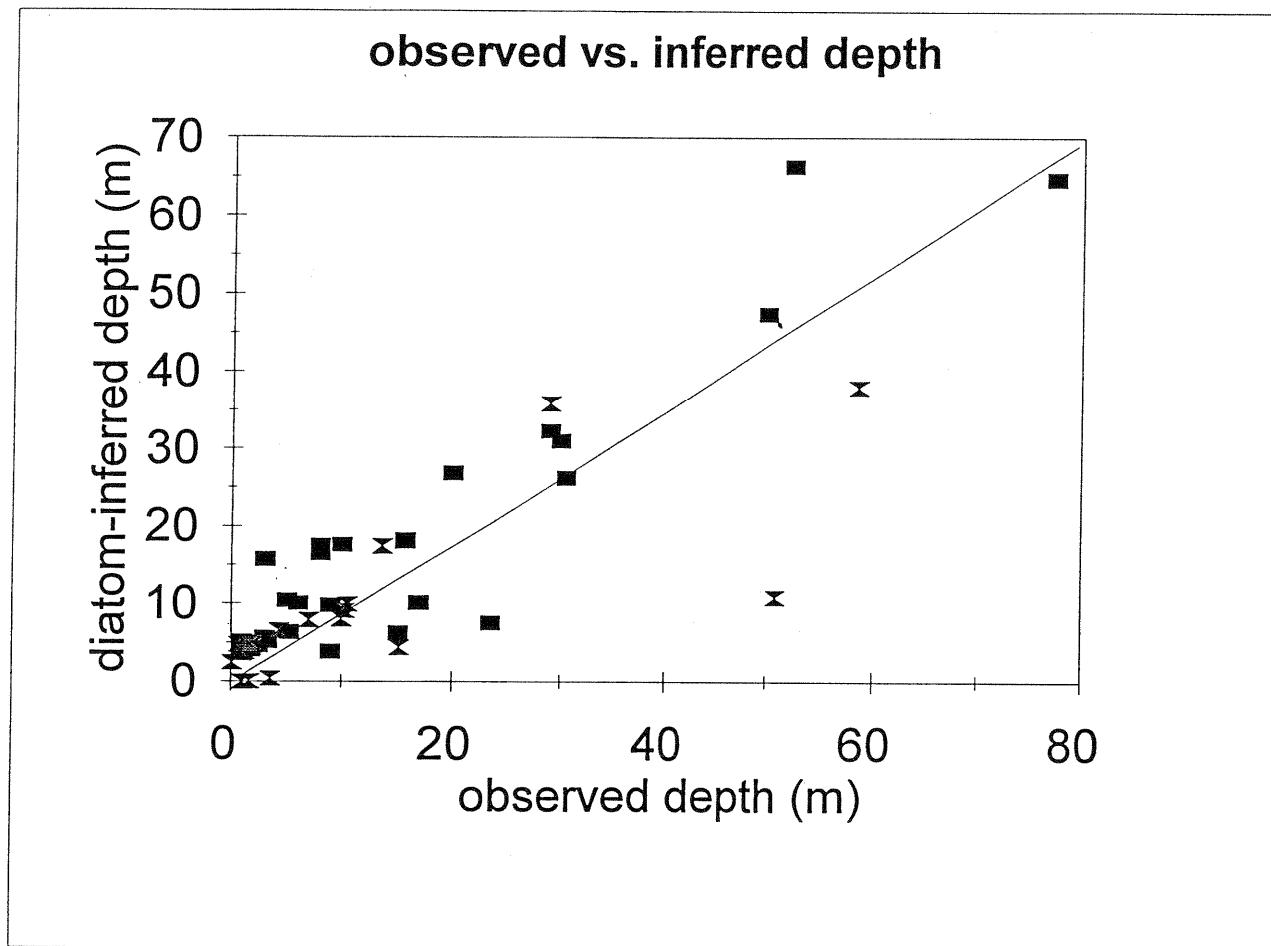


Figure 23. Measured depth versus diatom-inferred depth for lakes in the Cascade diatom calibration set. Cascade lakes are squares and Rainier lakes are double triangles.

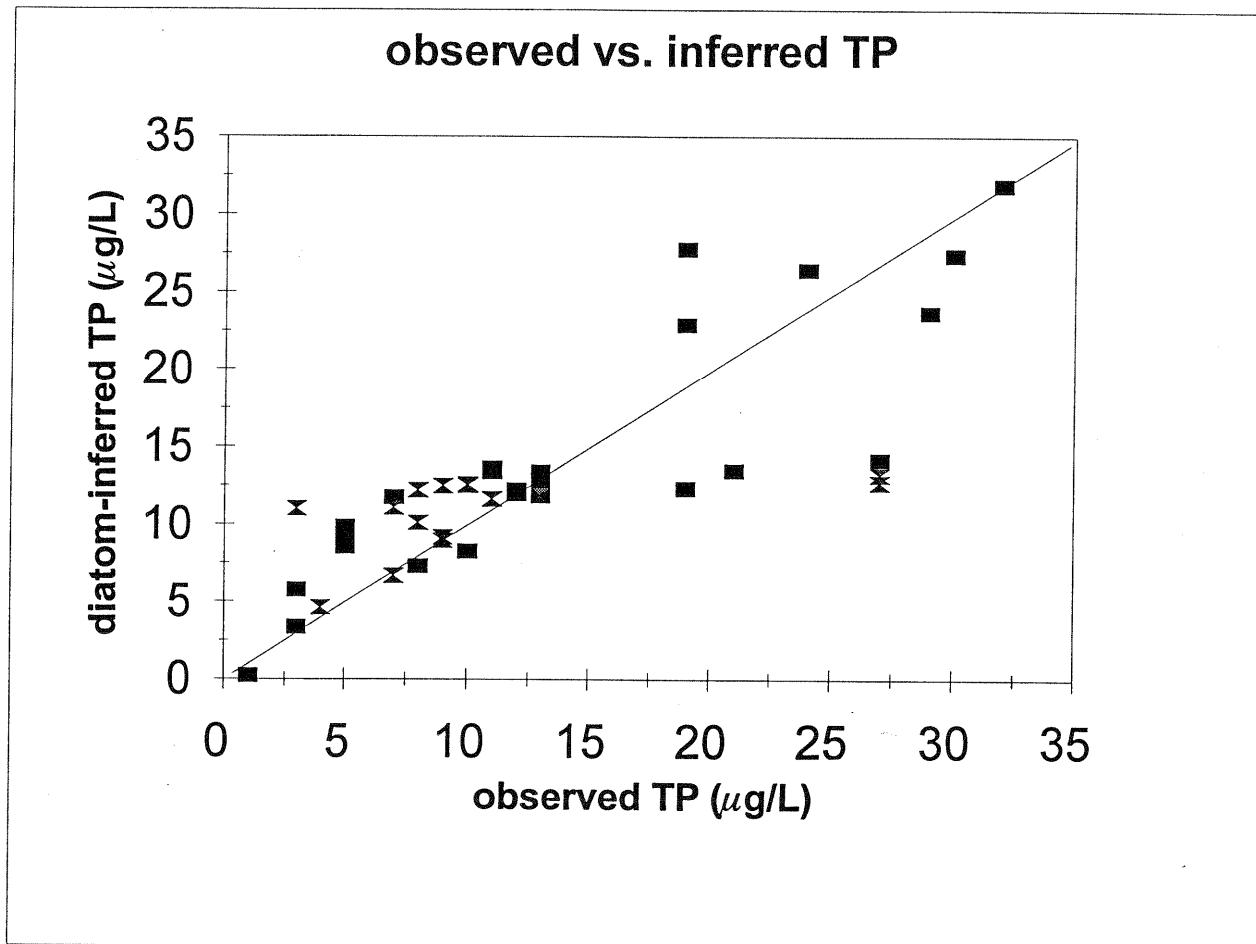


Figure 24. Measured total phosphorus (TP) versus diatom-inferred total phosphorus for lakes in the Cascade diatom calibration set. Cascade lakes are squares and Rainier lakes are double triangles.

Table 14. Surface water chemistry of Summit Lake, WA and adjacent pond. (Source: Ellers et al. 1996a)

	7/17/73 ^a	9/21/85 ^b	11/20/93	4/16/94	7/11/95	Routine	Split ^c	Lake	Pond	10/26/95	3/21/96	Duplicate	6/27/96
pH													
ANC (μeq/L)	5.94	5.74	5.53	5.24	5.37	5.95	5.30	5.29	5.27	5.28	5.30	5.30	
Conductivity (μS)	3.9	0.8	1.3	-2.6	2.3	5.8	-0.6	-2.3	-1.2	-1.0	-0.5		
Base Cations (μeq/L)	4	2.6	3.8	4.8	3.7	3.5	4.1	3.6	3.8	3.96	3.95	4.6	
Ca	6.9	5.0	12.4	8.9	4.1	5.3	4.6	4.6	5.3	5.7	5.8		
Mg	3.9	3.4	8.1	4.9	1.7	3.3	3.5	3.4	3.8	4.1	3.4		
Na	11.4	14.1	12.4	12.4	11.7	10.1	10.9	9.8	13.8	14.1	12.0		
K	0.8	1.0	0.9	0.8	1.0	0.6	0.7	2.3	0.9	0.9	0.4		
NH ₄	0	0	1.3	1.4	0	<0.7	0.5	0.6	0.6	0.7	0		
Acid Anions (μeq/L)													
SO ₄	9.2	8.6	11.5	7.7	8.2	8.7	7.5	7.2	9.2	9.6	7.8		
Cl	12.5	12.2	18.5	10.4	9.8	10.8	11.3	9.9	9.8	9.8	10.1		
NO ₃	0.1	0	1	0	0	<0.06	0	0	0.6	0.9	0		
Miscellaneous													
TP (μg/L)	6	3.8	0.24				<10						
SiO ₂ (mg/L)			0.34				0.71						
DOC (mg/L)							<1.0						
Chlorophyll a													
QA Checks													
pH _{cal}	6.01	5.74	5.79	5.41	5.88	6.14	5.60	5.44	5.54	5.56	5.61		
ANC _{cal} (μeq/L)	1.2	2.7	3.8	8.9	0.5	-0.2	0.9	3.0	4.8	4.5	3.7		
Conductivity _{cal} (μS)	3.8	3.6	5.4	5.0	4.0	3.2	4.3	4.7	4.8	4.8	4.4		
Ion Balance (+/-)	0.89	1.17	1.24	1.89	1.12	0.81	1.34	1.51	1.52	1.52	1.49		
(w/o RCOO ⁻)													

^a Bortleson et al. (1976)^b Source: Ellers et al. (1987)^c Split sample with ESE, Gainesville, FL

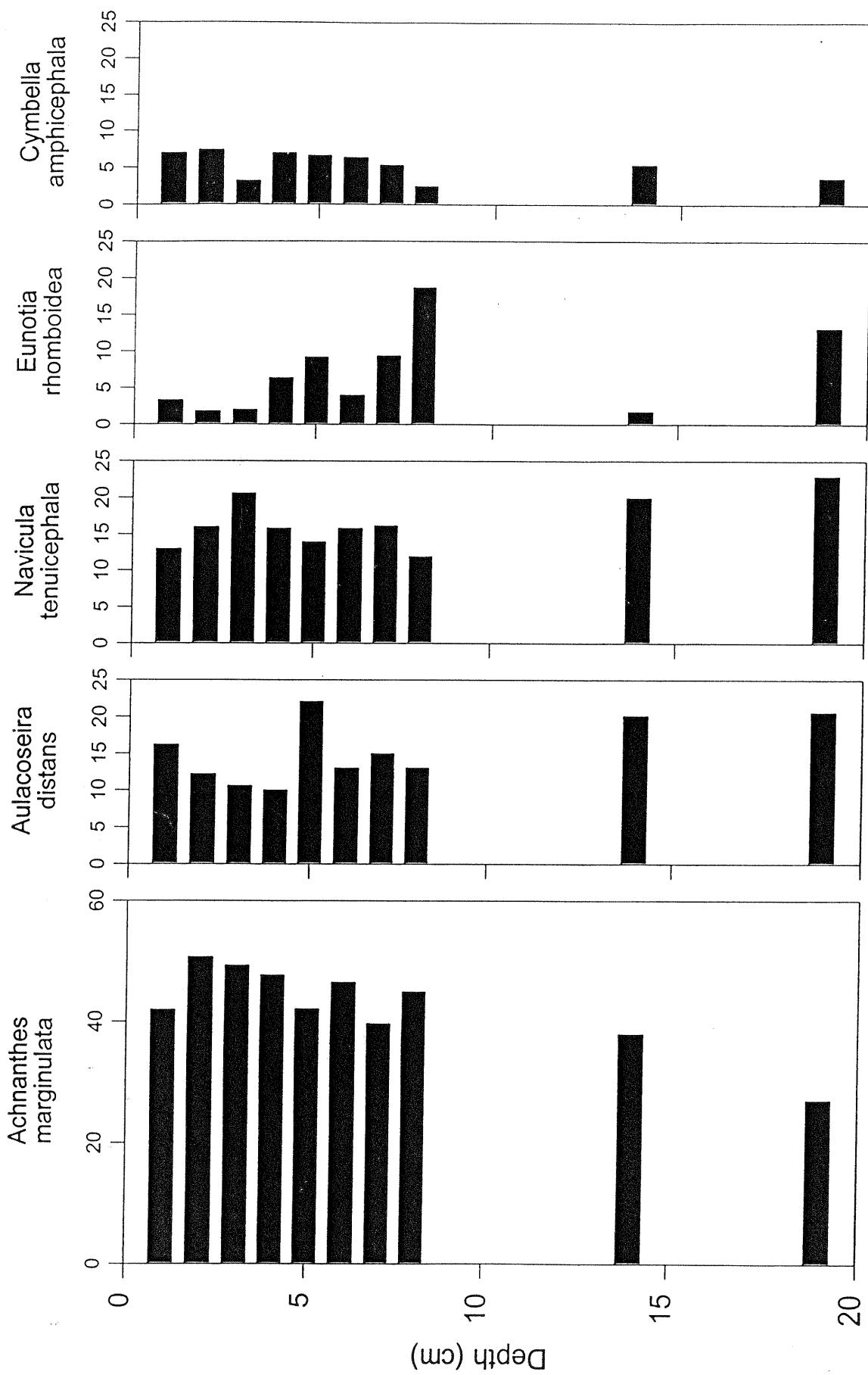


Figure 25. Relative abundance (%) of the five most abundant diatom taxa present in the sediments of Summit Lake. (Source: Eilers et al. 1996a)

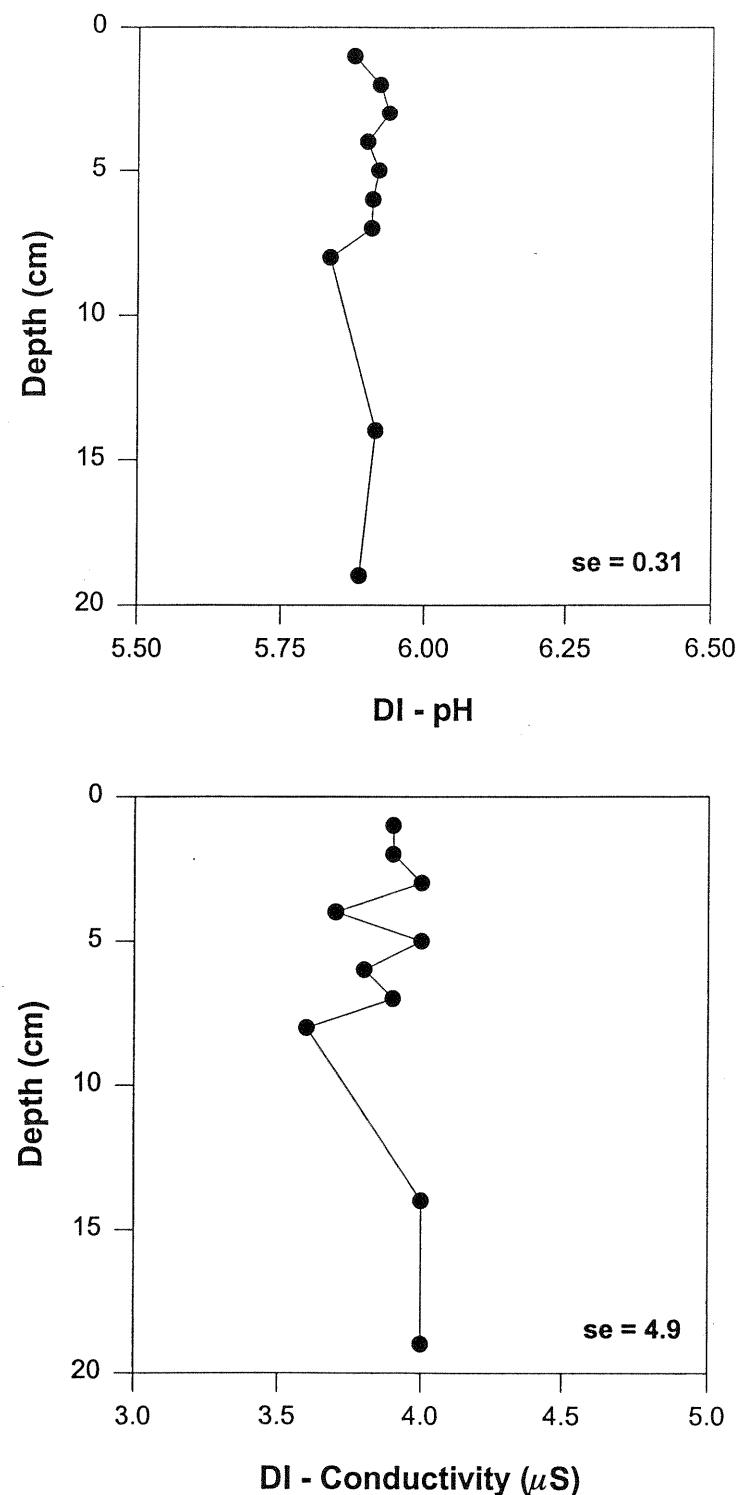


Figure 26. Diatom-inferred (DI) pH (top) and diatom-inferred conductivity (bottom) for sediments from Summit Lake, WA generated by applying the calibration equations developed in this study to diatom data reported in Eilers et al. (1996a).

between 5.83 and 5.94 among the 11 samples analyzed. Although the diatom community composition exhibited some minor changes, the pH optimum of the various taxa was remarkably constant. The diatom-inferred pH of 5.8 to 5.9 matched the pH values measured in the lake on two dates, although measured values from the Forest Service laboratory show pH values between 5.3-5.7 (Table 15). The diatom-inferred conductivity also was very stable with a range of only 3.6 to 4.0 μS . The actual measured range for conductivity in the lake was 2.6 to 4.8 μS (Table 15).

The results from the diatom-inferred reconstruction of pH and conductivity in Summit Lake illustrates that the lake chemistry has been remarkably stable for the previous three thousand years. This is particularly surprising given the inputs of volcanic ash during two events in this period. Factors contributing to this stability may include: (1) depth of the lake (>50 m); (2) small watershed contributing area; and (3) low weathering rates in the basin. The estimated hydraulic residence time is long ($\tau = 9$ to 14 yr) which would act to dampen fluctuations in climate and other factors. The considerable depth of the lake minimizes recycling from the sediments and promotes permanent burial of settling particles into the sediments. Summit Lake, as its name implies, is located near the crest of a ridge called the Rooster Comb which would minimize watershed inputs from both surface and groundwater. Weathering of the andesite in the watershed is extremely low as shown in the base cation concentrations (sea salt-corrected $C_B^* = 8.5 \mu\text{eq/L}$). Thus, inputs from the watershed, including base cations and nutrients, is extraordinarily small.

Another unusual feature of Summit Lake that may have a bearing on its acid-base chemistry is the aquatic bryophyte (moss) present on the lake bottom. Samples of bryophytes were collected from depths of over 35 m in Summit Lake and appear to cover a significant portion of the substrate. Most of the biomass in the lake is probably represented by the bryophytes. These aquatic plants have the capability to act as ion exchangers by sequestering base cations (esp. Ca^{2+}) and releasing hydrogen ion. The ability of some bryophytes (e.g., *Sphagnum*) to acidify their environment is well documented (e.g., Clymo 1967, Glime et al. 1982). Whether the bryophytes affect the chemistry of Summit Lake is unknown.

Table 15. Diatom-inferred reconstruction of lake pH and conductivity (μS) for Summit Lake, WA using the Cascade calibration set from this study and the Sierra Nevada (Whiting et al. 1989) calibration set.

Sediment Interval (cm)	pH		Conductivity	
	Cascades	Sierra Nevada	Cascades	Sierra Nevada
0-1	5.88	6.05	6.0	6.7
1-2	5.92	6.04	6.1	6.5
2-3	5.94	6.10	5.9	6.8
3-4	5.90	6.08	5.1	6.1
4-5	5.92	6.02	5.2	6.6
5-6	5.91	6.05	5.7	6.6
6-7	5.91	6.09	4.9	6.4
7-8	5.83	6.09	3.6	5.3
13-14	5.91	6.08	6.5	7.6
18-19	5.89	6.16	4.4	7.2
r^2	0.836	0.784	0.928	0.673
s	0.31	0.385	4.9	13.2

The question remains, "Why is the ANC near 0 $\mu\text{eq/L}$ ($\bar{x} = 1.0 \mu\text{eq/L}$, $n=9$) when C_B^* is ~9 $\mu\text{eq/L}$?" The current sulfate concentration (8.7 $\mu\text{eq/L}$; $\text{SO}_4^* = 7.6$) may be a recent phenomenon leading to a gradual depletion of ANC. For example, if historical ANC¹ were 8 $\mu\text{eq/L}$ then the historical pH would have been about 6.0. The calculated pH if ANC declined from its present 1 $\mu\text{eq/L}$ to -7 $\mu\text{eq/L}$ (equal to the possible historical maximum decline in ANC from 8 $\mu\text{eq/L}$ to 1 $\mu\text{eq/L}$) would be about 5.6.

The reconstruction of Summit Lake reveals that the use of the new Cascades calibration set provides an improvement in accuracy over the Sierra Nevada calibration set for inferring environmental variables from high altitude lakes in the Pacific Northwest (Table 15). Both pH and conductivity inferred from surface sediments are closer to the observed values. The inferred values from the Cascade calibration set are more accurate for the Cascade lakes than use of the calibration set from the Sierra Nevada. The core assemblages vary greatly in major taxa such as

¹ Organic anions are not included in these calculations because: (1) the measured DOC was 0.3 mg/L (Eilers et al. 1987); (2) the anion deficit ($\sum \text{cations} - \sum \text{anions}$ was small ($\bar{x}=1.3 \mu\text{eq/L}$, $n=10$); and Secchi disk transparency was high (21 m).

Navicula tenuicephala, *Achnanthes marginulata*, and *Aulacoseira distans*, however, the optima for these species are not greatly different and so the stratigraphies are remarkably stable (Figure 26).

It is not surprising that the shape of the stratigraphy does not change, even though the accuracy is improved. Sweets *et al.* (1990) noted that Florida diatom pH optima were significantly different from those found in the other regions of the PIRLA project, sometimes by as much as a full pH unit. However, the relative position of the pH optima of the species was largely unchanged. That this holds true between the Sierra Nevada and Cascade data sets is a testament to the robustness of the diatom technique, while also highlighting the importance of using regional diatom calibration sets to improve accuracy in lake reconstructions.

E. DISCUSSION AND CONCLUSIONS

The results indicate that the Cascade calibration set is a valuable new tool for reconstructing historical water quality chemistry in Cascade lakes. Among the variables examined, pH shows the strongest statistical relationship with the diatom assemblages and exhibits an acceptable standard error of prediction. Conductivity also is strongly related to the diatom assemblages, but it exhibits a high standard error which currently limits its utility for reconstructions to low conductivity (< 50 µS) lakes. Other variables also were strongly related to the diatom assemblages including ANC, individual base cations, total phosphorus, and lake depth. At present, the standard errors of prediction for these variables are still too high to reconstruct historical lake attributes with confidence, but they have considerable potential for future application in the Cascades. Some variables such as total phosphorus are currently hampered by high variability associated with the measured lake concentrations. The annual variability of total phosphorus increases with increasing TP concentrations which means that relatively few samples are required to characterize TP in oligotrophic lakes, but a large number of samples are required to characterize TP in eutrophic lakes assuming equal estimates of precision are required. Additional lakewater chemistry data would likely improve the TP calibrations.

Base cations such as calcium, magnesium, and sodium were highly correlated with conductivity. It is likely that these ions, either individually or collectively, could also be used to develop inferences to the diatom assemblages. However, because both ANC and base cations are so highly correlated with conductivity ($r^2=0.87$ for ANC, for individual base cations all $r^2 > 0.86$) in these lakes, there is likely to be little advantage in further exploring use of base cations and ANC. Individual ions such as sodium, however, could potentially be of interest for lakes with known anthropogenic inputs of sodium-based substances.

Surprisingly, the diatom assemblages showed strong relationships with physical variables such as lake depth. The range in lake depths (77 m) provided a relatively high r^2 value (0.75), but again the standard error of the prediction (8.8 m) was so great that there is little value in refining this relationship at this time. It is likely that this relationship was heavily influenced by a number of very

shallow lakes and ponds from Mt. Rainier National Park. In general, the Cascade diatom assemblages were dominated by either large percentages of planktonic diatoms in the larger, more productive lakes or by complex groups of benthic diatoms in small, shallow oligotrophic lakes. Other physical variables were not explored in detail although Secchi disk transparency, elevation, and hydraulic residence time may help to resolve some of the unexplained variance in the data set.

The taxonomic challenges in the Cascade calibration set were considerable. Over 13% of the taxa remain undescribed as species. Much of the taxonomic uncertainty is associated with the *Aulacoseira distans* complex, *Achnanthes* spp., and *Cymbella* spp. These taxa are generally associated with low nutrient, clearwater lakes that are slightly acidic (pH optimum near 5.5). Further advances in the use of paleolimnology in the Cascades will probably require resolution of the taxonomic uncertainty of these three genera.

As with any calibration data set, the errors in the diatom-inferred predication often can be improved by adding additional lakes to the data set. The distributions in Figures 2-8 should be helpful in deciding the types of lakes that would be most useful for better representing the population of Cascade lakes. In addition, some consideration should be given to geographic distribution associated with sampling additional data for this calibration set. The current calibration set is heavily weighted towards lakes in the Washington Cascades, particularly those in Mt. Rainier National Park (MORA). Fortunately, the MORA lakes exhibit no particular chemical extremes that unduly influenced the regression equations. However, the geographic distribution could be improved by adding several lakes from the northern Washington Cascades and additional lakes from the Oregon Cascades (Figure 1). Note that only 17 of the 48 study lakes were located in Oregon.

The pH reconstruction of Summit Lake was helpful in resolving some of the uncertainty in the current lake chemistry. The low pH of the lake and its proximity to emission sources raised concern that the lake had begun to acidify from atmospheric deposition. The pH reconstruction showed that Summit Lake has been stable for the last 3150 years. This finding does not exclude the possibility that Summit Lake could acidify in the future, but it clarifies the historical status of the lake. As noted

earlier, given its virtual lack of acid neutralizing capacity, Summit Lake could exhibit significant declines in pH with further inputs of acid sulfate ions. It would be desirable to test the calibration set on one or more lakes in the Cascades that have experienced changes in the 20th century. In particular, some of the lakes moderately impacted by ashfall from Mount St. Helens would be excellent candidates for diatom reconstructions. Additionally, lakes that are in national forests subject to the effects of timber harvest or recreational development might be suitable candidates for reconstruction.

F. LITERATURE CITED

- Arzet, K., D. Krause-Dellin, and C. Steinberg. 1986. Acidification of four lakes in the Federal Republic of Germany as reflected by diatom assemblages, cladoceran remains, and sediment chemistry, pp. 227-250. In: J.P. Smol, R.W. Battarbee, R.B. Davis, and J. Meriläinen, eds. *Diatoms and Lake Acidity*. Dr. W. Junk, Dordrecht, The Netherlands.
- Battarbee, R.W. 1973. A new method for the estimation of absolute microfossil numbers, with special reference to diatoms. *Limnol. Oceanogr.* 18:647-653.
- Battarbee, R.W. 1984. Diatom analysis and the acidification of lakes. *Phil. Trans. R. Soc. Lond. B*, 305:451-477.
- Birks, H.J.B., J.M. Line, S. Juggins, A.C. Stevenson, and C.J.F. ter Braak. 1990. Diatoms and pH reconstruction. *Phil. Trans. R. Soc. Lond. B* 327:263-278.
- Camburn, K.E., J.C. Kingston, & D.F. Charles. 1986. PIRLA diatom iconograph. PIRLA unpublished report series, Report No. 3. Indiana University Biology Dept. Bloomington, IN, USA.
- Charles, D.F. 1984. Recent pH history of Big Moose Lake (Adirondack Mountains, New York, USA) inferred from sediment diatom assemblages. *Verh. Internat. Verein. Limnol.* 22:559-566.
- Charles, D.F. 1985. Relationships between surface sediment diatom assemblages and lakewater characteristics in Adirondack lakes. *Ecology* 66:994-1011.
- Charles, D.F. 1987. Diatom counts of Adirondack lake surface sediment samples. PIRLA Unpublished Report Series, Report number 12, Department of Biology, Indiana University, Bloomington, IN. 393 pp.
- Charles, D.F., and S.A. Norton. 1986. Paleolimnological evidence for trends in atmospheric deposition of acids and metals. pp. 335-435. In: Acid deposition: Long-term Trends. Committee on monitoring and assessment of trends in acid deposition. National Academy Press, Washington, D.C.
- Charles, D.F., and J.P. Smol. 1988. New methods for using diatoms and chrysophytes to infer past pH of low-alkalinity lakes. *Limnol. Oceanogr.* 33:1451-1462.
- Charles, D.F. and J.P. Smol. 1990. The PIRLA II project: regional assessment of lake acidification trends. *Verh. Internat. Verein. Limnol.* 24:474-480.
- Charles, D.F., and D.R. Whitehead (Eds.). 1988. *Paleoecological Investigation of Recent Lake Acidification. Developments in Hydrobiology*. Dr. W. Junk, Dordrecht, Netherlands.
- Charles, D.F., D. R. Whitehead, D.S. Anderson, R. Bienert, K.E. Camburn, R.B. Cook, T.L. Crisman, R.B. Davis, B.D. Fry, R.A. Hites, J.S. Kahl, J.C. Kingston, R.G. Kreis, Jr., M.J. Whiting, and R.J. Wise. 1986. The PIRLA project (Paleoecological Investigation of Recent Lake Acidification): preliminary results for the Adirondacks, New England, N. Great Lake States, and N. Florida. *Water Air Soil Pollut.* 30:355-365.
- Charles, D.F., D.R. Whitehead, D.R. Engstrom, B.D. Fry, R.A. Hites, S.A. Norton, J.S. Owen, L.A. Roll, S.C. Schindler, J.P. Smol, A.J. Uutala, J.R. White, and R.J. Wise. 1987. Paleolimnological evidence for recent acidification of Big Moose Lake, Adirondack Mountains, N.Y. (USA). *Biogeochemistry* 3:267-296.

Charles, D.F., R.W. Battarbee, I. Renberg, H. van Dam, and J.P. Smol. 1989. Paleoecological analysis of lake acidification trends in North America and Europe using diatoms and chrysophytes. In: Norton, S.A., S.E. Lindberg, and A.L. Page (eds.). Acid Precipitation. Vol. 4. Soils, Aquatic Processes, and Lake Acidification. Springer-Verlag, New York. pp. 208-276.

Charles, D.F., M.W. Binford, E.T. Furlong, R.A. Hites, M.J. Mitchell, S.A. Norton, F. Oldfield, M.J. Paterson, J.P. Smol, A.J. Uutala, J.R. White, D.R. Whitehead, and R.J. Wise. 1990. Paleoecological investigation of recent lake acidification in the Adirondack Mountains, N.Y. J. Paleolimnol. 3:195-241.

Clymo, R.S. 1967. Control of cation concentration, and in particular of pH, in *Sphagnum* dominated communities. In Golterman, J.S. and R.S. Glymo (eds.). Chemical Environment in the Aquatic Habitat. Amsterdam, N.V. Noord-Hollandsche Uitgevers Maatschappij. pp. 273-284.

Cumming, B.F., J.P. Smol, J.C. Kingston, D.F. Charles, H.J.B. Birks, K.E. Camburn, S.S. Dixit, A.J. Uutala, and A.R. Selle. 1992. How much acidification has occurred in Adirondack region (New York, USA) lakes since pre-industrial times? Can. J. Fish. Aquat. Sci. 49:128-141.

Davidson, G.A. 1984. Paleolimnological reconstruction of the acidification history of an experimentally acidified lake. Master's Thesis, University of Manitoba. 186 pp.

Davis, R.B., and D.S. Anderson. 1985. Methods of pH calibration of sedimentary diatom remains for reconstructing history of pH in lakes. Hydrobiologia 120:69-87.

Davis, R.B., and F. Berge. 1980. Atmospheric deposition in Norway during the last 300 years as recorded in SNSF lake sediments II. Diatom stratigraphy and inferred pH, pp. 270-271. In: D. Drablos and A. Tolland, eds. Ecological impact of acid precipitation. Proceedings of an International Conference, Sandefjord, Norway, SNSF Project, Oslo, Norway.

Davis, R.B., S.A. Norton, C.T. Hess, and D.F. Brakke. 1983. Paleolimnological reconstruction of the effects of atmospheric deposition of acids and heavy metals on the chemistry and biology of lakes in New England and Norway. Hydrobiologia 103:113-123.

Del Prete, A., and C. Schofield. 1981. The utility of diatom analysis of lake sediments for evaluating acid precipitation effects on dilute lakes. Archiv. Hydrobiol. 91:332-340.

DeNicola, D.M. 1986. The representation of living diatom communities in deep-water sedimentary diatom assemblages in two Maine (U.S.A.) lakes, pp. 73-85. In: J.P. Smol et al., eds. Diatoms and Lake Acidity. Dr. W. Junk Publishers, Dordrecht, The Netherlands.

Dixit, S.S. 1983. The utility of sedimentary diatoms as a measure of historical lake pH. Master's Thesis, Brock University, Ontario.

Dixit, S.S., A.S. Dixit, and R.D. Evans. 1987. Paleolimnology evidence of recent acidification in two Sudbury (Canada) lakes. Sci. Total Environ. 67:53-67.

Dixit, S.S., A.S. Dixit, and J.P. Smol. 1989a. Lake acidification recovery can be monitored using chrysophycean microfossils. Can. J. Fish. Aquat. Sci. 46:1309-1312.

Dixit, S.S., A.S. Dixit, and J.P. Smol. 1989b. Acidification and recovery of Baby Lake, Sudbury, Ontario, Canada. Presented at Vth Intern. Symp. on Paleolimnology, Cumbria, U.K. Sept. 1-6, 1989.

Eilers, J.M., C.P. Gubala, and P.R. Sweets. 1996a. Limnology of Summit Lake, WA with special emphasis on its acid-base chemistry and sensitivity to impacts from atmospheric deposition. Final Report submitted to Mt. Baker-Snoqualmie National Forest. E&S Environmental Chemistry, Inc., Corvallis, OR.

Eilers, J.M. J.A. Bernert, S. S. Dixit, C. P. Gubala, and P. R. Sweets. 1996b. Processes influencing water quality in a sub-alpine Cascade Mountain lake. *Northwest Science* 70:59-70.

Eilers, J.M., P. Kanciruk, R.A. McCord, W.S. Overton, L. Hook, D.J. Blick, D.F. Brakke, P.E. Kellar, M.S. DeHaan, M.E. Silverstein, and D.H. Landers. 1987. Characteristics of lakes in the western United States. Volume II. Data compendium for selected physical and chemical variables. EPA-600/3-86/054b, U.S. Environmental Protection Agency, Washington, D.C. 492 pp.

Flower, R.J., and R.W. Battarbee. 1983. Diatom evidence for recent acidification of two Scottish Lochs. *Nature* 305:130-133.

Flower, R.J., R.W. Battarbee, and P.G. Appleby. 1987. The recent paleolimnology of acid lakes in Galloway, south-west Scotland: diatom analysis, pH trends, and the role of afforestation. *J. Ecol.* 75:797-824.

Gasse, F., and F. Tekaia. 1983. Transfer functions for estimating paleoecological conditions (pH) from East African diatoms. *Hydrobiologia* 103:85-90.

Glime, J.M., R.G. Wetzel, and B.J. Kennedy. 1982. the effects of bryophytes on succession from alkaline marsh to *Sphagnum* bog. *Amer. Midland Nat.* 108:209-223.

Heit, M., Y.L. Tan, C. Klusek, and J.C. Burke. 1981. Anthropogenic trace elements and polycyclic aromatic hydrocarbon levels in sediment cores from two lakes in the Adirondack acid lake region. *Water Air Soil Pollut.* 15:441-464.

Husar, R.B., T.J. Sullivan, and D.F. Charles. 1991. Historical trends in atmospheric sulfur deposition and methods for assessing long-term trends in surface water chemistry. pp. 65-82, In: Charles, D.F. (ed.). Acidic Deposition and Aquatic Ecosystems. Regional Case Studies. Springer-Verlag, New York.

Hustedt, F. 1939. Systematische und okologische Untersuchungen über die Diatomeen-Flora von Java, Bali, und Sumatra nach dem Material der Deutschen Limnologischen Sunda-Expedition III. Die Okologischen Factor in und ihr Einfluss auf die Diatomeenflora. *Archiv. für Hydrobiol. Suppl.* 16:274-394.

Huttenen, P., and J. Meriläinen. 1986. Diatom response to pH and humic matter of the water. pp. 47-54. In: J. Simola, ed. Proc. Finnish-Soviet Symp. Methods in Paleoecology and the Nordic Meeting of Diatomologists, Karelian Institute, Joensuu, Finland.

Jones, V.J., and R.J. Flower. 1986. Spatial and temporal variability in periphytic diatom communities: Paleoecological significance in an acidified lake, pp. 87-94. In: J.P. Smol et al., eds. Diatoms and Lake Acidity. Dr. W. Junk Publishers, Dordrecht, The Netherlands.

Jones, V.J. and S. Juggins. 1995. The construction of a diatom-based chlorophyll a transfer function and its application at three lakes on Siyny Island (Maritime Antarctic) subject to differing degrees of nutrient enrichment. *Freshw. Biol.* 34:433-445.

Juggins, S. & C.J.F. ter Braak. 1992. CALIBRATE - a program for species-environment calibration by [weighted averaging] partial least squares regression. Environmental Change Research Center, University College, London.

- Kingston, J.C., and H.J.B. Birks. 1990. Dissolved organic carbon reconstruction from diatom assemblages in PIRLA project lakes, North America. *Phil. Trans. R. Soc. Lond. B* 327:279-288.
- Kingston, J.C., R.B. Cook, R.G. Kreis, K.E. Camburn, S.A. Norton, P.R. Sweets, M.W. Binford, M.J. Mitchell, S.C. Schindler, L. Shane, and G. King. 1990. Paleoecological investigation of recent lake acidification in the northern Great Lakes States. In: Charles, D.F. and D.W. Whitehead (eds.). *Paleoecological investigation of Recent Lake Acidification*. Kluwer Academic Press, Dordrecht, The Netherlands.
- Krammer, K. & H. Lange-Bertalot. 1991. *Bacillariophyceae 3. Teil: Centrales, Fragilariaeae, Eunotiaceae*. Gustav Fischer, Stuttgart.
- Line, J.M., and H.J.B. Birks. 1990. WACALIB version 2.1 - a computer program to reconstruct environmental variables from fossil assemblages by weighted averaging. *J. Paleolimnol.* (in press).
- Line, J.M., C.J.F. ter Braak, & H.J.B. Birks. 1994. WACALIB version 3.3 - a computer program to reconstruct environmental variables from fossil assemblages by weighted averaging and to derive sample-specific errors of prediction. *J. Paleolim.* 10: 147-152.
- Oksanen, J., E. Läära, P. Huttunen, and J. Meriläinen. 1988. Estimation of pH optima and tolerances of diatoms in lake sediments by the methods of weighted averaging, least squares, and maximum likelihood, and their use for the prediction of lake acidity. *J. Paleolimnol.* 1:39-49.
- Patrick, R. & C.W. Reimer. 1966. *The Diatoms of the United States Exclusive of Alaska and Hawaii*. Academy of the Natural Sciences of Philadelphia, Philadelphia, PA.
- Patrick, R. & C.W., Reimer. 1975. *The Diatoms of the United States Exclusive of Alaska and Hawaii*. Academy of Natural Sciences of Philadelphia, Philadelphia, PA.
- Renberg, I., and T. Hellberg. 1982. The pH history of lakes in southwestern Sweden, as calculated from the subfossil diatom flora of the sediments. *Ambio* 11:30-33.
- Renberg, I., and J.E. Wallin. 1985. The history of the acidification of Lake Gärdsjön as deduced from diatoms and Sphagnum leaves in the sediment, pp. 47-52. In: F. Andersson and B. Olsson, eds. *Lake Gärdsjön: An Acid Forest and Its Catchment*. Ecological Bulletin No. 37.
- Renberg, I., T. Korsman, and H.J.B. Birks. 1993. Prehistoric increases in the pH of acid-sensitive Swedish lakes caused by land-use changes. *Nature* 362:824-826.
- Smol, J.P. 1992. Paleolimnology: an important tool for effective ecosystem management. *J. Aquat. Ecosystem Health* 1:49-58.
- Smol, J.P., D.F. Charles, and D.R. Whitehead. 1984. Mallomonadacean (Chrysophyceae) assemblages and their relationships with limnological characteristics in 38 Adirondack (New York) lakes. *Can. J. of Bot.* 62:911-923.
- Smol, J.P., R.W. Battarbee, R.B. Davis, and J. Meriläinen, Eds. 1986. *Diatoms and Lake Acidity*. Dr. W. Junk, Dordrecht.
- Stevenson, A.C., H.J.B. Birks, R.J. Flower, R.W. Battarbee. 1989. Diatom-based pH reconstruction of lake acidification using Canonical Correspondence Analysis. *Ambio* 18:228-233.
- Sullivan, T.J. 1990. Historical Changes in Surface Water Acid-Base Chemistry in Response to Acidic Deposition. State of the Science, SOS/T 11, National Acid Precipitation Assessment Program. 212 pp.

- Sullivan, T.J., D.L. Kugler, M.J. Small, C.B. Johnson, D.H. Landers, B.J. Rosenbaum, W.S. Overton, W.A. Kretser, and J. Gallagher. 1990. Variation in Adirondack, New York, lakewater chemistry as a function of surface area. *Water Resour. Bull.* 26:167-176.
- Sweets, P.R., R.W. Bienert, T.L. Crisman, & M.W. Binford. 1990. Paleoecological investigations of recent lake acidification in northern Florida. *J. Paleolim.* 4: 103-137.
- Tan, Y.L., and M. Heit. 1981. Biogenic and abiogenic polynuclear aromatic hydrocarbons in sediments from two remote Adirondack lakes. *Geochim. Cosmochim. Acta* 45:2267-2279.
- ter Braak, C.J.F. 1985. Correspondence analysis of incidence and abundance data: properties in terms of a unimodal response model. *Biometrics* 41:859-873.
- ter Braak, C.J.F. 1986. Canonical correspondence analysis: a new eigenvector technique for multivariate direct gradient analysis. *Ecology* 67:1167-1179.
- ter Braak, C.J.F. 1987a. The analysis of vegetation-environment relationship by Canonical correspondence analysis. *Vegetatio* 69:79-87.
- ter Braak, C.J.F. 1987b. CANOCO - a FORTRAN program for canonical community ordination by partial detrended canonical correspondence analysis, principal components analysis and redundancy analysis (version 2.0). TNO Institute of Applied computer Science, Statistical Department Wageningen, 6700 AC Wageningen, The Netherlands.
- ter Braak, C.J.F. 1988. Partial canonical correspondence analysis, pp. 551-558. In H.H. Bock, ed. *Classification and Related Methods of Data Analysis*. North-Holland:Amsterdam.
- ter Braak, C.J.F., and L.G. Barendregt. 1986. Weighted averaging of species indicator values: its efficiency in environmental calibration. *Mathemat. Biosci.* 78:57-72.
- ter Braak, C.J.F., and N.J.M. Gremmen. 1987. Ecological amplitudes of plant species and the internal consistency of Ellenberg's indicator values for moisture. *Vegetatio* 69:79-87.
- ter Braak, C.J.F., and C.W.N. Loosman. 1986. Weighted averaging, logistic regression and the Gaussian response model. *Vegetatio* 65:3-11.
- ter Braak, C.J.F., and I.C. Prentice. 1988. The theory of gradient analysis. *Advan. Ecolog. Res.* 18:271-317.
- ter Braak, C.J.F., and H. van Dam. 1989. Inferring pH from diatoms: a comparison of old and new calibration methods. *Hydrobiologia* 178:209-223.
- Tolonen, K., and T. Jaakkola. 1983. History of lake acidification and air pollution studied on sediments in South Finland. *Annales Botanici Fennici* 20:57-78.
- van Dam, H., B. van Geel, A. van der Wijk, J.F.M. Geelen, R. van der Heijden, and M.D. Dickman. 1988. Palaeolimnological and documented evidence for alkalization and acidification of two Moorland pools (The Netherlands). *Rev. Paleobot. Palynol.* 55:273-316.
- Whitehead, D.R., D.F. Charles, & R.A. Goldstein, 1990. The PIRLA project (Paleoecological Investigation of Recent Lake Acidification): an introduction to the synthesis of the project. *J. Paleolim.* 3: 187-194.
- Whiting, M.C., D.R. Whitehead, R.B. Holmes, and S.A. Norton. 1989. Paleolimnological reconstruction of recent acidity changes in four Sierra Nevada lakes. *J. Paleolimnol.* 2:285-304.

G. APPENDICES

1. Diatom Counts for the Cascade Diatom Calibration Set
2. Compressed Diatom Taxa List Used in Ordination and Modeling for the Cascade Diatom Calibration Set

APPENDIX 1

Diatom Counts for the Cascade Diatom Calibration Set

Diatom Counts

Long name	TAXANAME	Division	Genus	Species	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta	Achnanthes	sp. PRS 0A				
Achn sp. 1 ?	AC1?	Chrysophyta	Achnanthes	sp. 1 ?				
Achn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA				
Achn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA				
Achn sp. 13	AC13PRL	Chrysophyta	Achnanthes	sp. 13 PIRLA				
Achn sp. 14	AC14PRL	Chrysophyta	Achnanthes	sp. 14 PIRLA				
Achn sp. 16	AC16PRL	Chrysophyta	Achnanthes	sp. 16 PIRLA				
Achn sp. 19	AC19PRL	Chrysophyta	Achnanthes	sp. 19 PIRLA				
Achn sp 2 Mt. Rain.	AC2MRNP	Chrysophyta	Achnanthes	sp. 2				
Achn acares	ACACARES	Chrysophyta	Achnanthes	acares				
Achn altaica	ACALTAIC	Chrysophyta	Achnanthes	altaica				
Achn austriaca	ACaustrI	Chrysophyta	Achnanthes	austriaca				
Achn bioreti	ACBIORET	Chrysophyta	Achnanthes	bioreti				
Achn calcar	ACCALCAR	Chrysophyta	Achnanthes	calcar				
Achn chilidanos	ACCHLIDA	Chrysophyta	Achnanthes	chilidanos				
Achn clevei	ACCLEVEI	Chrysophyta	Achnanthes	clevei				
Achn curtissima	ACCURTIS	Chrysophyta	Achnanthes	curtissima				
Achn dauti alaskaensi	ACdauala	Chrysophyta	Achnanthes	dauti				
Achn detha	ACdetha	Chrysophyta	Achnanthes	detha				
Achn didyma	ACDIDYMA	Chrysophyta	Achnanthes	didyma				
Achn exigua	ACEXIGUA	Chrysophyta	Achnanthes	exigua				
Achn flexella	ACTflexel	Chrysophyta	Achnanthes	flexella				
Achn grana	ACGRANA	Chrysophyta	Achnanthes	grana				
Achn grischuna	ACGRISCH	Chrysophyta	Achnanthes	grischuna				
Achn helvetica	ACHelvet	Chrysophyta	Achnanthes	helvetica				
Achn holstii	ACHOLSTI	Chrysophyta	Achnanthes	holstii				
Achn hungarica	ACHUNGAR	Chrysophyta	Achnanthes	hungarica				
Achn kuelbsii	ACKUELBS	Chrysophyta	Achnanthes	kuelbsii				
Achn lanceolata	ACLANCEO	Chrysophyta	Achnanthes	lanceolata				
Achn lanceolat dubia	ACLANDUB	Chrysophyta	Achnanthes	lanceolata				
Achn lan ssp. frequ	ACLANFRE	Chrysophyta	Achnanthes	lanceolata				
Achn laterostrata	ACLATERO	Chrysophyta	Achnanthes	laterostrata				
Achn levanderi	ACLEVAND	Chrysophyta	Achnanthes	levanderi				
Achn levan helvetica	ACLevhel	Chrysophyta	Achnanthes	levanderi				
Achn linearis curta	ACLINCUR	Chrysophyta	Achnanthes	linearis				
Achn linearis	ACLINEAR	Chrysophyta	Achnanthes	linearis				
Achn marginulata	ACMARGIN	Chrysophyta	Achnanthes	marginulata				

Diatom Counts

Long name	TAXANAME	Division	Genus	Species	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta	Achnanthes	sp. PRS 0A	0.00400			
Achn sp. 1 ?	AC1?	Chrysophyta	Achnanthes	sp. 1 ?				
Achn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA				
Achn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA				
Achn minutissima	ACMINUTI	Chrysophyta	Achnanthes	minutissima				
Achn nodosa	ACNODOSA	Chrysophyta	Achnanthes	nodosa				
Achn pusilla	ACPUSILL	Chrysophyta	Achnanthes	pusilla				
Achn spp.	ACSPP	Chrysophyta	Achnanthes	spp.				
Achn subatomoides	ACSUBATO	Chrysophyta	Achnanthes	subatomoides				
Achn suchlandii	ACsuchlia	Chrysophyta	Achnanthes	suchlandii				
Amph sp. A Cascades	AM0Arscs	Chrysophyta	Amphora	sp. PRS 0A				
Amph libyca	AMLIBYCA	Chrysophyta	Amphora	libyca				
Amph ovalis	AMOVALIS	Chrysophyta	Amphora	ovalis				
Amph ovali pediculus	AMOvaped	Chrysophyta	Amphora	ovalis				
Amph perpusilla	AMPERPUS	Chrysophyta	Amphora	perpusilla				
Anom brachysira	ANBRACHY	Chrysophyta	Anomeoneis	brachysira				
Anom serians	ANSERIAN	Chrysophyta	Anomeoneis	serians				
Anom vitrea	ANVITREA	Chrysophyta	Anomeoneis	vitreia				
Aste formosa	ASFORMOS	Chrysophyta	Asterionella	formosa				
Aste ralfsii	ASRALFSI	Chrysophyta	Asterionella	ralfsii				
Aula sp. A Cascades	AU0Arscs	Chrysophyta	Aulacoseira	sp. PRS 0A				
Aula sp. B Cascades	AU0Brscs	Chrysophyta	Aulacoseira	sp. PRS 0B				
Aula sp. C Cascades	AU0Crscs	Chrysophyta	Aulacoseira	sp. PRS 0C				
Aula sp. D Cascades	AU0Drscs	Chrysophyta	Aulacoseira	sp. PRS 0D				
Aula ambigua	AUAMBIGU	Chrysophyta	Aulacoseira	ambigua				
Aulo distans humilis	AUDISNIV	Chrysophyta	Aulacoseira	distantis				
Aulo distans nivalis	AUDISNLO	Chrysophyta	Aulacoseira	distantis				
Aulo dist nivaloides	AUDISTAN	Chrysophyta	Aulacoseira	distantis				
Aulo distans tenella	AUDisten	Chrysophyta	Aulacoseira	distantis				
Aula granulata	AUGRANUL	Chrysophyta	Aulacoseira	granulata				
Aula italicica	AITALIC	Chrysophyta	Aulacoseira	italicica				
Aula italicica subarct	AUitasba	Chrysophyta	Aulacoseira	italicica				
Aulo italic tenu	AUITATEN	Chrysophyta	Aulacoseira	italicica				
Aulo lirata	AULIRATA	Chrysophyta	Aulacoseira	lirata				
Aula pergabra	AUPERGLA	Chrysophyta	Aulacoseira	perglabra				
Aula paffiana	AUPFAFFI	Chrysophyta	Aulacoseira	paffiana				

Diatom Counts

Long name	TAXANAME	Division	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta	Genus Achnanthes	Species sp. PRS 0A		
Achn sp. 1 ?	AC1?	Chrysophyta	Achnanthes	sp. 1 ?		
Achn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA		
Achn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA		
Aula species	AUSPP	Chrysophyta	Aulacoseira	spp.		
Aula validida	AUVALIDA	Chrysophyta	Aulacoslera	valida		
Calo bacillum	CABACILL	Chrysophyta	Caloneis	bacillum		
Calo hyalina	CAHYALIN	Chrysophyta	Caloneis	hyalina		
Calo silicula	CASILICU	Chrysophyta	Caloneis	silicula		
Calo tenuis	CATENUIS	Chrysophyta	Caloneis	tenuis		
Cocc placentula	CCPLACEN	Chrysophyta	Cocconeis	placentula		
Cocc placen euglypta	CCPLAEUG	Chrysophyta	Cocconeis	placentula		
Chae sp. 1?	CH1?	Chrysophyta	Chaetoceros	sp. 1?		
Cymb sp. A Cascades	CM0Arscs	Chrysophyta	Cymbella	sp. PRS 0A		
Cymb sp. B Cascades	CM0Brscs	Chrysophyta	Cymbella	sp. PRS 0B		
Cymb sp. 1 ?	CM1?	Chrysophyta	Cymbella	sp. 1 ?		
Cymb sp. 18	CM18PRL	Chrysophyta	Cymbella	sp. 18 PIRLA		
Cymb sp. 2 ?	CM2?	Chrysophyta	Cymbella	sp. 2 ?		
Cymb sp. 20	CM20PRL	Chrysophyta	Cymbella	sp. 20 PIRLA		
Cymb sp. 21	CM21PRL	Chrysophyta	Cymbella	sp. 21 PIRLA		
Cymb sp. 6	CM6PRL	Chrysophyta	Cymbella	sp. 6 PIRLA		
Cymb sp. 7	CM7PRL	Chrysophyta	Cymbella	sp. 7 PIRLA		
Cymb aequalis	CMAEQUAL	Chrysophyta	Cymbella	aequalis		
Cymb amphicephala	CMAMPHIC	Chrysophyta	Cymbella	amphicephala		
Cymb brehmii	CMBREHMI	Chrysophyta	Cymbella	brehmii		
Cymb caespitosa	CMCAESP1	Chrysophyta	Cymbella	caespitosa		
Cymb cesatii	CMCESAT1	Chrysophyta	Cymbella	cesatii		
Cymb cistula	CMcistul	Chrysophyta	Cymbella	cistula		
Cymb descripta	CMDESCRI	Chrysophyta	Cymbella	descripta		
Cymb elginensis	CMELGINE	Chrysophyta	Cymbella	elginensis		
Cymb falaisensis	CMFALAI5	Chrysophyta	Cymbella	falaensis		
Cymb gaeumannii	CMGAEUMA	Chrysophyta	Cymbella	gaeumannii		
Cymb hauckii	CMHAUCKI	Chrysophyta	Cymbella	hauckii		
Cymb hebridica	CMHEBRID	Chrysophyta	Cymbella	hebridica		
Cymb heterostrobra	CMhetsub	Chrysophyta	Cymbella	heteropleura		
Cymb inaequalis	CMInaequ	Chrysophyta	Cymbella	inaequalis		
Cymb lunata	CMLUNATA	Chrysophyta	Cymbella	lunata		
				0.00200		
				0.00600		
				0.00400		
				0.00200		
				0.01400		
				0.01200		

Diatom Counts

Long name	TAXANAME	Division	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta				
Achn sp. 1 ?	AC1?	Chrysophyta				
Achn sp. 11	AC1PRL	Chrysophyta				
Achn sp. 12	AC12PRL	Chrysophyta				
Cymb mesiana	CMMESIAN	Chrysophyta				
Cymb mexicana	CMMEXCAN	Chrysophyta				
Cymb microcephala	CMMICROC	Chrysophyta				
Cymb minut latens	CMinlat	Chrysophyta				
Cymb minut silesiaca	CMminsil	Chrysophyta				
Cymb minuta	CMMINUTA	Chrysophyta				
Cymb muelleri	CMMUELLE	Chrysophyta				
Cymb naviculiformis	CMnavicu	Chrysophyta				
Cymb perpusilla	CMSINUAT	Chrysophyta				
Cymb sinuata	CMSPP	Chrysophyta				
Cymb spp.	CTSOLEA	Chrysophyta				
Cyma solea						
Cycl sp. 1	CY1PRL	Chrysophyta				
Cycl bod lemanica	CYBODLEM	Chrysophyta				
Cycl kuetzin radiosa	CYKuetra	Chrysophyta				
Cycl ocellata	CYOCELLA	Chrysophyta				
Cycl pseudostelligera	Cypseste	Chrysophyta				
Cycl stelligera	CYSTELLI	Chrysophyta				
Diat anceps	DAANCEPS	Chrysophyta				
Diat hiemale	DAhiemal	Chrysophyta				
Diat mesodon	DAMESODO	Chrysophyta				
Diat tenuis	DATENUIS	Chrysophyta				
Dipl sp. 1 ?	DP1?	Chrysophyta				
Dipl elliptica	DPELLIPT	Chrysophyta				
Dipl finnica	DPFINNIC	Chrysophyta				
Dipl marginistriata	DPMARGIN	Chrysophyta				
Dipl modica	DPMODICA	Chrysophyta				
Epit adnata	EPADNATA	Chrysophyta				
Epit sorex	EPSOREX	Chrysophyta				
Epit turgida	EPTURGID	Chrysophyta				
Epit turgida gracil	EPTURGRA	Chrysophyta				
Euno sp. 1 ?	EU1?	Chrysophyta				
Euno sp. 42	EU42PRL	Chrysophyta				
			Species sp. PRS OA			
			sp. 1 ?			
			sp. 11 PIRLA			
			sp. 12 PIRLA			
			mesiana			
			mexicana			
			microcephala			
			minuta			
			minuta			
			minuta			
			muelleri			
			naviculiformis			
			perpusilla			
			sinuata			
			spp.			
			solea			
			sp. 1 PIRLA			
			bodanica			
			kuetzingiana			
			ocellata			
			pseudostelligera			
			stelligera			
			anceps			
			hiemale			
			mesodon			
			tenue			
			sp. 1 ?			
			elliptica			
			finnica			
			marginistriata			
			modica			
			adnata			
			sorex			
			turgida			
			turgida			
			sp. 1 ?			
			sp. 42 PIRLA			
			0.00083			
			0.000200			
			0.01400			
			0.000200			
			0.000200			

Diatom Counts

Long name	TAXANAME	Division	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta	Genus	Species		
Achn sp. 1 ?	AC1?	Chrysophyta	Achnanthes	sp. PRS 0A		
Achn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 1 ?		
Achn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA		
Euno sp. 44	EU44PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA		
Euno sp. 45	EU45PRL	Chrysophyta	Eunotia	sp. 44 PIRLA		
Euno sp. 46	EU46PRL	Chrysophyta	Eunotia	sp. 45 PIRLA		
Euno arcus	EUARCUS	Chrysophyta	Eunotia	sp. 46 PIRLA		
Euno bidentula var 1	EUbi1PRL	Chrysophyta	Eunotia	arcus		
Euno bidentula	EUBIDENT	Chrysophyta	Eunotia	bidentula		
Euno bilun mucophila	EUBLMUC	Chrysophyta	Eunotia	bilunaris		
Euno bilunaris	EUBLUNA	Chrysophyta	Eunotia	bilunaris		
Euno curvata	EUcurvat	Chrysophyta	Eunotia	curvata		
Euno denticulata	EUdentic	Chrysophyta	Eunotia	denticulata		
Euno exigua	EUEXIGUA	Chrysophyta	Eunotia	exigua		
Euno flexuosa	EUFLLEXUO	Chrysophyta	Eunotia	flexuosa		
Euno hemicyclus	EUHEMICY	Chrysophyta	Eunotia	hemicyclus		
Euno implicata	EUIMPLIC	Chrysophyta	Eunotia	implicata		
Euno incisa var 1	EUin1PRL	Chrysophyta	Eunotia	incisa		
Euno incisa var 2	EUin2PRL	Chrysophyta	Eunotia	incisa		
Euno incisa var 6	EUin6PRL	Chrysophyta	Eunotia	incisa		
Euno incisa	EUINCISA	Chrysophyta	Eunotia	intermedia		
Euno intermedia	EUInterm	Chrysophyta	Eunotia	meisteri		
Euno meisteri	EUMEISTE	Chrysophyta	Eunotia	microcephala		
Euno microcephala	EUMicroc	Chrysophyta	Eunotia	microcephala		
Euno micr tridentata	EUMictri	Chrysophyta	Eunotia	minor		
Euno minor	EUMINOR	Chrysophyta	Eunotia	monodon		
Euno monodon	EUMONODO	Chrysophyta	Eunotia	muscicola		
Euno musci trident	EUMUSTRI	Chrysophyta	Eunotia	naegelii		
Euno naegelii	EUNAEGL	Chrysophyta	Eunotia	paludosa		
Euno palud trinac	EUPALTRI	Chrysophyta	Eunotia	paludos		
Euno paludosa	EUPALUDO	Chrysophyta	Eunotia	pectinalis		
Euno pectinali minor	EUpecmin	Chrysophyta	Eunotia	pectinalis		
Euno pectinalis	EUPECTIN	Chrysophyta	Eunotia	praerupta		
Euno praerupta	EUPRAERU	Chrysophyta	Eunotia	quaternaria		
Euno quaternaria	EUquater	Chrysophyta	Eunotia	rhomboidea		
Euno rhomboidea	EURHOMBO	Chrysophyta	Eunotia			

Diatom Counts

Long name	TAXANAME	Division	Genus	Species	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta	Achnanthes	sp. PRS 0A				
Achn sp. 1 ?	AC1?	Chrysophyta	Achnanthes	sp. 1 ?				
Achn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA				
Achn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA				
Euno serra diadema	EUserdia	Chrysophyta	Eunotia	serra				
Euno serra	EUSERRA	Chrysophyta	Eunotia	serra				
Euno spp.	EUSPP	Chrysophyta	Eunotia	spp.	0.000400			
Euno subarcuatooides	EUSUBARC	Chrysophyta	Eunotia	subarcuatooides	0.00200			
Euno sudetica	EUSUDETTI	Chrysophyta	Eunotia	sudetica	0.00600			
Euno tenella	EUTENELL	Chrysophyta	Eunotia	tenella				
Euno trinacria	EURINAC	Chrysophyta	Eunotia	trinacia				
Euno vanheurckii	EUVANHEU	Chrysophyta	Eunotia	vanheurckii				
Euno sp. 74	EUVANINT	Chrysophyta	Eunotia	vanheurckii				
Frag sp. A Cascades	FR0Arsces	Chrysophyta	Fragilaria	sp. PRS 0A				
Frag sp. B Cascades	FR0Brscs	Chrysophyta	Fragilaria	sp. PRS 0B				
Frag sp. 11	FR11PRL	Chrysophyta	Fragilaria	sp. 11 PIRLA				
Frag sp. 15	FR15PRL	Chrysophyta	Fragilaria	sp. 15 PIRLA				
Frag sp. 17	FR17PRL	Chrysophyta	Fragilaria	sp. 17 PIRLA				
Frag sp. 9	FR9PRL	Chrysophyta	Fragilaria	sp. 9 PIRLA				
Frag arcus	FRARCUS	Chrysophyta	Fragilaria	arcus				
Frag bidens	FRBIDENS	Chrysophyta	Fragilaria	bidens				
Frag brevistriata	FRBREVIS	Chrysophyta	Fragilaria	brevistriata				
Frag capu rumpens	FRCAPRUM	Chrysophyta	Fragilaria	capucina				
Frag capicina	FRCAPUCI	Chrysophyta	Fragilaria	capucina				
Frag capu vaucheriae	FRCAPVAU	Chrysophyta	Fragilaria	capucina				
Frag constru binodis	FRCONBIN	Chrysophyta	Fragilaria	construens				
Frag constru pumila	FRCONPUM	Chrysophyta	Fragilaria	construens				
Frag construens	FRCONSTU	Chrysophyta	Fragilaria	construens				
Frag constru subsali	FRCONSUB	Chrysophyta	Fragilaria	construens				
Frag constru venter	FRCONVEN	Chrysophyta	Fragilaria	construens				
Frag crotensis	FRCROTON	Chrysophyta	Fragilaria	crotensis				
Frag constricta	FRctastr	Chrysophyta	Fragilaria	constricta				
Frag exigua	FREXIGUA	Chrysophyta	Fragilaria	exigua				
Frag fasciculata	FRFASCIC	Chrysophyta	Fragilaria	fasciculata				
Frag lapponica	FRLappon	Chrysophyta	Fragilaria	lapponica				
Frag leptostau dubia	FRLEPDUB	Chrysophyta	Fragilaria	leptostauron				
Frag oldenburgiana	FROLDENB	Chrysophyta	Fragilaria	oldenburgiana				

Diatom Counts

Long name	TAXANAME	Division	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta	Achnanthes	sp. PRS 0A		
Achn sp. 1 ?	AC1?	Chrysophyta	Achnanthes	sp. 1 ?		
Achn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA		
Achn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA		
Frag parasitica	FRPARASI	Chrysophyta	Fragilaria	parasitica		
Frag pinn lancettula	FRPINLAN	Chrysophyta	Fragilaria	pinnata	0.00200	
Frag pinnata	FRPINNAT	Chrysophyta	Fragilaria	pinnata	0.13400	0.43000
Frag pseudoconstruens	FRPSECON	Chrysophyta	Fragilaria	pseudoconstruens	0.12600	0.32500
Frag spp.	FRSPP	Chrysophyta	Fragilaria	spp.	0.01400	0.00200
Frag vaucheriae	FRvauche	Chrysophyta	Fragilaria	vaucheriae		
Frag virescens	FRVIRESC	Chrysophyta	Fragilaria	virescens		
Frag vires exi	FRvirexi	Chrysophyta	Fragilaria	virescens		
Frus rhom amphipleur	FSRHOAMP	Chrysophyta	Frustulia	rhomboides		
Frus rhomboides CAF	FSrhoCAF	Chrysophyta	Frustulia	rhomboides	0.02600	
Frus rhom crassiner	FSRHOCRA	Chrysophyta	Frustulia	rhomboides	0.07800	
Frus rhomboides	FSRHOMBO	Chrysophyta	Frustulia	rhomboides		
Frus rhombo saxonica	FSrhosax	Chrysophyta	Frustulia	rhomboides		
Gomp sp. A Cascades	GO0ARSCS	Chrysophyta	Gomphonema	sp. PRS 0A		
Gomp sp. 1 ?	GO1?	Chrysophyta	Gomphonema	sp. 1 ?		
Gomp sp. 11	GO11PRL	Chrysophyta	Gomphonema	sp. 11 PIRLA		
Gomp sp. 18	GO18PRL	Chrysophyta	Gomphonema	sp. 18 PIRLA		
Gomp sp. 1	GO1PRL	Chrysophyta	Gomphonema	sp. 1 PIRLA		
Gomp angustum	GOANGSTM	Chrysophyta	Gomphonema	angustum		
Gomp angustatum	GOANGUST	Chrysophyta	Gomphonema	angustatum		
Gomp consector	GOconsec	Chrysophyta	Gomphonema	consector		
Gomp gracile	GOGRACIL	Chrysophyta	Gomphonema	gracile		
Gomp grovei lingulat	GOGROLIN	Chrysophyta	Gomphonema	grovei		
Gomp olivcm mintsnum	GOOLIMIN	Chrysophyta	Gomphonema	olivaceum		
Gomp olivaceum	GOOLIVCM	Chrysophyta	Gomphonema	olivaceum		
Gomp parvulum	GOPARVUL	Chrysophyta	Gomphonema	parvulum		
Gomp puig aequatoria	GOpuiaeq	Chrysophyta	Gomphonema	puiggarianum		
Gomp spp.	GOSPP	Chrysophyta	Gomphonema	spp.		
Gomp subcl mexicanum	GOsubmex	Chrysophyta	Gomphonema	subclavatum		
Gomp tack brevistria	GOtacbre	Chrysophyta	Gomphonema	tackei		
Gomp turris	GOTURRIS	Chrysophyta	Gomphonema	turris		
Hann arcus	HNarcus	Chrysophyta	Hannaea	arcus		
Meri circulare	MDCIRCUL	Chrysophyta	Meridion	circulare		

Diatom Counts

Long name	TAXANAME	Division	Genus	Species	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta	Achnanthes	sp. PRS 0A	0.00400			
Achn sp. 1?	AC1?	Chrysophyta	Achnanthes	sp. 1?				
Actn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA				
Actn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA				
Melo sp. 1?	ME1?	Chrysophyta	Melosira	sp. 1?				
Melo sp. 13	ME13PRL	Chrysophyta	Melosira	sp. 13 PIRLA				
Melo sp. 4	ME4PRL	Chrysophyta	Melosira	sp. 4 PIRLA				
Melo sp. 8	ME8PRL	Chrysophyta	Melosira	sp. 8 PIRLA				
Navi sp. 1 Mt. Rain.	NA01mwm	Chrysophyta	Navicula	sp. 1 MW				
Navi sp. A Cascades	NA0Arscs	Chrysophyta	Navicula	sp. PRS 0A				
Navi sp. B Cascades	NA0Brscs	Chrysophyta	Navicula	sp. PRS 0B				
Navi sp. C Cascades	NA0Crscs	Chrysophyta	Navicula	sp. PRS 0C				
Navi sp. D Cascades	NA0Drscs	Chrysophyta	Navicula	sp. PRS 0D				
Navi sp. E Cascades	NA0Erscs	Chrysophyta	Navicula	sp. 0E				
Navi sp. 1?	NA1?	Chrysophyta	Navicula	sp. 1?				
Navi sp. 14	NA14PRL	Chrysophyta	Navicula	sp. 14 PIRLA				
Navi sp. 1	NA1PRL	Chrysophyta	Navicula	sp. 1 PIRLA				
Navi sp. 2?	NA2?	Chrysophyta	Navicula	sp. 2?				
Navi sp. 20	NA20PRL	Chrysophyta	Navicula	sp. 20 PIRLA				
Navi sp. 21	NA21PRL	Chrysophyta	Navicula	sp. 21 PIRLA				
Navi sp. 24	NA24PRL	Chrysophyta	Navicula	sp. 24 PIRLA				
Navi sp. 25	NA25PRL	Chrysophyta	Navicula	sp. 25 PIRLA				
Navi sp. 3?	NA3?	Chrysophyta	Navicula	sp. 3?				
Navi sp. 45	NA45PRL	Chrysophyta	Navicula	sp. 45 PIRLA				
Navi sp. 47	NA47PRL	Chrysophyta	Navicula	sp. 47 PIRLA				
Navi sp. 51	NA51PRL	Chrysophyta	Navicula	sp. 51 PIRLA				
Navi sp. 52	NA52PRL	Chrysophyta	Navicula	sp. 52 PIRLA				
Navi sp. 6	NA6PRL	Chrysophyta	Navicula	sp. 6 PIRLA				
Navi sp. 74	NA74PRL	Chrysophyta	Navicula	sp. 74 PIRLA				
Navi sp. 91	NA91PRL	Chrysophyta	Navicula	sp. 91 PIRLA				
Navi aboensis	NAABOENS	Chrysophyta	Navicula	aboenensis				
Nav absoluta	NAABSOLU	Chrysophyta	Navicula	absoluta				
Navi acceptata	NAACCEPT	Chrysophyta	Navicula	acceptata				
Navi accomoda	NAACCOMO	Chrysophyta	Navicula	accommoda				
Navi angusta	NAangust	Chrysophyta	Navicula	angusta				
Navi arvensis	NAARVEN	Chrysophyta	Navicula	arvensis				
Navi bacillum	NABACLUM	Chrysophyta	Navicula	bacillum				

Diatom Counts

Long name	TAXANAME	Division	Genus	Species	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta	Achnanthes	sp. PRS 0A	0.00400			
Achn sp. 1 ?	AC1?	Chrysophyta	Achnanthes	sp. 1 ?				
Achn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA				
Achn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA				
Navi bremensis	NABREMEN	Chrysophyta	Navicula	bremensis				
Navi bryophila	NABRYPHL	Chrysophyta	Navicula	bryophilila				
Navi cincta	NACINCTA	Chrysophyta	Navicula	cincta				
Navi cocconeiformis	NACOCCON	Chrysophyta	Navicula	cocconeiformis				
Navi cohni	NACOHNII	Chrysophyta	Navicula	cohni				
Navi concentrica	NACONCEN	Chrysophyta	Navicula	concentrica	0.00200			
Navi crucicula	NACRUCIC	Chrysophyta	Navicula	crucicula				
Navi cryptocephala	NACRYPTO	Chrysophyta	Navicula	cryptocephala	0.00400	0.00200		
Navi cryptotenella	NACRYTEN	Chrysophyta	Navicula	cryptotenella	0.00200	0.01400	0.000083	
Navi disjuncta	NADISJUN	Chrysophyta	Navicula	disjuncta				
Navi eligensis	NAELGINE	Chrysophyta	Navicula	eligensis				
Navi gallica	NAGALIC	Chrysophyta	Navicula	gallica				
Navi harderii	NAHARDER	Chrysophyta	Navicula	harderii				
Navi jaemelefeldti	NAJAERNE	Chrysophyta	Navicula	jaemelefeldti				
Navi laevissima	NALAEVIS	Chrysophyta	Navicula	laevissima				
Navi leptostriata	NALEPTOS	Chrysophyta	Navicula	leptostrata				
Navi libonensis	NALIBONE	Chrysophyta	Navicula	libonensis				
Navi medioconvexa	NAMEDCON	Chrysophyta	Navicula	medioconvexa				
Navi mediocris	NAMEDIOC	Chrysophyta	Navicula	mediocris				
Navi mediopunctata	NAMEDIOP	Chrysophyta	Navicula	mediopunctata				
Navi menisculus	NAMENSCL	Chrysophyta	Navicula	menisculus				
Navi minima	NAMINIMA	Chrysophyta	Navicula	minima				
Navi minuscula v. A.	Namins0A	Chrysophyta	Navicula	minuscula				
Navi modica	NAMODICA	Chrysophyta	Navicula	modica				
Navi mutica	NAPELLIC	Chrysophyta	Navicula	mutica				
Navi pelliculosa	NAPICNTA	Chrysophyta	Navicula	pelliculosa				
Navi placenta	NAPORIFFE	Chrysophyta	Navicula	placenta				
Nav porifera	NAPSEMUR	Chrysophyta	Navicula	porifera	0.00200			
Navi pseudomuralis	NAPSESCU	Chrysophyta	Navicula	pseudomuralis		0.01000		
Navi pseudoscutiform	NAPSEVEN	Chrysophyta	Navicula	pseudoscutiformis	0.00600	0.000083		
Navi pseudoventralis	NAPUPELL	Chrysophyta	Navicula	pseudoventralis				
Navi pup elliptica	NAPUPREC	Chrysophyta	Navicula	pupula	0.00200			
Navi pup rectangular				pupula				

Diatom Counts

Long name	TAXANAME	Division	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta	Achnanthes	sp. PRS 0A	0.00400	
Achn sp. 1 ?	AC1?	Chrysophyta	Achnanthes	sp. 1 ?		
Achn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA		
Achn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA		
Navi pupula	NAPUPULA	Chrysophyta	Navicula	pupula	0.00600	
Navi radiosua	NARADIOS	Chrysophyta	Navicula	radiosa	0.00400	0.00167
Navi radiosua parva	NARADPAR	Chrysophyta	Navicula	radiosa		
Navi rhynchocephala	NARHYNCH	Chrysophyta	Navicula	rhynchocephala	0.00400	0.00200
Navi seminuloides	NASEMDES	Chrysophyta	Navicula	seminuloides	0.00200	
Navi seminulum	NASEMLUM	Chrysophyta	Navicula	seminulum	0.01000	0.00400
Navi spp.	NASPP	Chrysophyta	Navicula	spp.	0.00400	
Navi cf subtil var 2	NASU2PRL	Chrysophyta	Navicula	subtilissima		
Navi cf subtil var 4	NASU4PRL	Chrysophyta	Navicula	subtilissima		
Navi cf subtil var 5	NASU5PRL	Chrysophyta	Navicula	subtilissima	0.01400	
Navi subatomoides	NASUBATO	Chrysophyta	Navicula	subatomoides	0.00200	
Navi subhyalina	NASUBHYA	Chrysophyta	Navicula	subhyalina		
Navi submuralis	NASUBMUR	Chrysophyta	Navicula	submuralis		
Navi subplacentula	NASUBPLA	Chrysophyta	Navicula	subplacentula		
Navi subrotundata	NASUBROT	Chrysophyta	Navicula	subrotundata		
Navi subtilissima	NASUBTIL	Chrysophyta	Navicula	subtilissima		
Navi tenuicephala	NATENUIC	Chrysophyta	Navicula	tenuicephala		
Navi tridentula	NATRIDEN	Chrysophyta	Navicula	tridentula		
Navi trivialis	NATRIVIA	Chrysophyta	Navicula	trivialis		
Nav veneta	NAVENETA	Chrysophyta	Navicula	veneta		
Navi viridula	NAVIRDLA	Chrysophyta	Navicula	viridula		
Navi vitabunda	NAVITABU	Chrysophyta	Navicula	vitabunda		
Neid sp. 13	NE13PRL	Chrysophyta	Neidium	sp. 13 PIRLA		
Neid sp. 4	NE4PRL	Chrysophyta	Neidium	sp. 4 PIRLA		
Neid affine	NEAFFINE	Chrysophyta	Neidium	affine		
Neid affin longiceps	NEAFFLON	Chrysophyta	Neidium	affine		
Neid alpinum	NEALPINU	Chrysophyta	Neidium	alpinum		
Neid alp quadripunct	NEalpqua	Chrysophyta	Neidium	alpinum		
Neid bisu baicalense	NEBISBAI	Chrysophyta	Neidium	bisulcatum		
Neid bisulcatum	NEBISULC	Chrysophyta	Neidium	bisulcatum		
Neid hertcyn f subros	NEHERSUB	Chrysophyta	Neidium	hercynicum		
Neid iridi ampliatum	NEIRIAMP	Chrysophyta	Neidium	iridis		
Neid ir amphigomphus	NEIRIAUS	Chrysophyta	Neidium	iridis		

Diatom Counts

Long name	TAXANAME	Division	Genus	Species	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta	Achnanthes	sp. FRS 0A	0.00400			
Achn sp. 1 ?	AC1?	Chrysophyta	Achnanthes	sp. 1 ?				
Achn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA				
Achn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA				
Neid iridis	NEIRIDIS	Chrysophyta	Neidium	iridis	0.00200			
Neid spp.	NESPP	Chrysophyta	Neidium	spp.	0.00400			
Nitz sp. A Cascades	NI0ARSCS	Chrysophyta	Nitzschia	sp. FRS 0A				
Nitz sp. 1 ?	NI1?	Chrysophyta	Nitzschia	sp. 1 ?				
Nitz sp. 12	NI12PRL	Chrysophyta	Nitzschia	sp. 12 PIRLA				
Nitz sp. 14	NI14PRL	Chrysophyta	Nitzschia	sp. 14 PIRLA				
Nitz sp. 15	NI15PRL	Chrysophyta	Nitzschia	sp. 15 PIRLA				
Nitz sp. 16	NI16PRL	Chrysophyta	Nitzschia	sp. 16 PIRLA				
Nitz sp. 17	NI17PRL	Chrysophyta	Nitzschia	sp. 17 PIRLA				
Nitz sp. 26	NI26PRL	Chrysophyta	Nitzschia	sp. 26 PIRLA				
Nitz sp. 34	NI34PRL	Chrysophyta	Nitzschia	sp. 34 PIRLA				
Nitz sp. 35	NI35PRL	Chrysophyta	Nitzschia	sp. 35 PIRLA				
Nitz sp. 36	NI36PRL	Chrysophyta	Nitzschia	sp. 36 PIRLA				
Nitz sp. 37	NI37PRL	Chrysophyta	Nitzschia	sp. 37 PIRLA				
Nitz sp. 38	NI38PRL	Chrysophyta	Nitzschia	sp. 38 PIRLA				
Nitz alpina	NIALPINA	Chrysophyta	Nitzschia	alpina	0.00600	0.00750		
Nitz amphibia	NIAMPHIB	Chrysophyta	Nitzschia	amphibia	0.00600	0.00600		
Nitz bacillariaeform	NIBACFOR	Chrysophyta	Nitzschia	bacillariaeformis				
Nitz brrophila	NIBRYOPH	Chrysophyta	Nitzschia	bryophila				
Nitz capitellata	NICAPITE	Chrysophyta	Nitzschia	capitellata				
Nitz dissipata	NIdissip	Chrysophyta	Nitzschia	dissipata				
Nitz dissip undulata	NIdisund	Chrysophyta	Nitzschia	dissipata				
Nitz fonticola	NIFONTIC	Chrysophyta	Nitzschia	fonticola				
Nitz fossilis	NIFOSSIL	Chrysophyta	Nitzschia	fossilis				
Nitz frustulum var 3	NIf3PRL	Chrysophyta	Nitzschia	frustulum				
Nitz frustulum var 8	NIf8PRL	Chrysophyta	Nitzschia	frustulum				
Nitz gracilis	NIGRACIL	Chrysophyta	Nitzschia	gracilis				
Nitz graciliformis	NIGRCLFM	Chrysophyta	Nitzschia	graciliformis	0.00400			
Nitz inconspicua	NIINCONS	Chrysophyta	Nitzschia	inconspicua				
Nitz lacuum	NILACUUM	Chrysophyta	Nitzschia	lacuum	0.00800			
Nitz leistikowii	NILEISTI	Chrysophyta	Nitzschia	leistikowii				
Nitz modesta	NIMODEST	Chrysophyta	Nitzschia	modesta	0.00250			
Nitz palea debilis	NIPALDEB	Chrysophyta	Nitzschia	palea				

Diatom Counts

Long name	TAXANAME	Division	Genus	Species	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	ACOARSSCS	Chrysophyta	Achnanthes	sp. PRS 0A	0.00400			
Achn sp. 1 ?	AC1?	Chrysophyta	Achnanthes	sp. 1 ?				
Achn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA				
Achn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA				
Nitz palea	NIPALEA	Chrysophyta	Nitzschia	palea				
Nitz paleaformis	NIPALEAF	Chrysophyta	Nitzschia	paleaeformis				
Nitz perminuta	NIPERMIN	Chrysophyta	Nitzschia	perminuta				
Nitz recta	NIRECTA	Chrysophyta	Nitzschia	recta				
Nitz spp.	NISPP	Chrysophyta	Nitzschia	spp.				
Nitz tropica	NITROPIC	Chrysophyta	Nitzschia	tropica				
Nitz vermicularis	NIVERMCL	Chrysophyta	Nitzschia	vermicularis				
Opep sp. 1	OP1PRL	Chrysophyta	Opephora	sp. 1 PIRLA				
Opep martyi	OPMARTYI	Chrysophyta	Opephora	martyi				
Opep Olsenii	OPOLSENI	Chrysophyta	Opephora	olsemini				
Pero fibula	PEFIBULA	Chrysophyta	Peronia	fibula				
Pinn sp. A Cascades	PI0Arscs	Chrysophyta	Pinnularia	sp. PRS 0A				
Pinn sp. B Cascades	PI0Brscs	Chrysophyta	Pinnularia	sp. PRS 0B				
Pinn sp. C Cascades	PI0Crscs	Chrysophyta	Pinnularia	sp. PRS 0C				
Pinn sp. D Cascades	PI0Drscs	Chrysophyta	Pinnularia	sp. PRS 0D				
Pinn sp. 24	PI24PRL	Chrysophyta	Pinnularia	sp. 24 PIRLA				
Pinn sp. 25	PI25PRL	Chrysophyta	Pinnularia	sp. 25 PIRLA				
Pinn sp. 28	PI28PRL	Chrysophyta	Pinnularia	sp. 28 PIRLA				
Pinn sp. 2 PIRLA	PI2PRL	Chrysophyta	Pinnularia	sp. 2 PIRLA				
Pinn sp. 30	PI30PRL	Chrysophyta	Pinnularia	sp. 30 PIRLA				
Pinn sp. 32	PI32PRL	Chrysophyta	Pinnularia	sp. 32 PIRLA				
Pinn sp. 36	PI36PRL	Chrysophyta	Pinnularia	sp. 36 PIRLA				
Pinn sp. 46	PI46PRL	Chrysophyta	Pinnularia	sp. 46 PIRLA				
Pinn abaujensis var 2	Plab2PRL	Chrysophyta	Pinnularia	abaujensis				
Pinn abaujensis	Plabaje	Chrysophyta	Pinnularia	sp. 32 PIRLA				
Pinn biceps	PIBICEPS	Chrysophyta	Pinnularia	biceps				
Pinn borealis rectan	PIBORREC	Chrysophyta	Pinnularia	borealis				
Pinn braunii	PIBRAUNI	Chrysophyta	Pinnularia	braunii				
Pinn brebissonii	Pibrebis	Chrysophyta	Pinnularia	brebissonii				
Pinn divergentissima	PIDIVTIS	Chrysophyta	Pinnularia	divergentissima				
Pinn gibba	PIGIBBA	Chrysophyta	Pinnularia	gibba				
Pinn gibba mesogong	PIGBMES	Chrysophyta	Pinnularia	gibba				
Pinn hemiptera	PIHEMIPT	Chrysophyta	Pinnularia	hemiptera				

Diatom Counts

Long name	TAXANAME	Division	Genus	Species	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta	Achnanthes	sp. PRS 0A	0.00400			
Achn sp. 1?	AC1?	Chrysophyta	Achnanthes	sp. 1?				
Achn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA				
Achn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA				
Pinn maior	PIMAIOR	Chrysophyta	Pinnularia	major				
Pinn microstauron	PIMICBRE	Chrysophyta	Pinnularia	microstauron				
Pinn nodosa	PINODOSA	Chrysophyta	Pinnularia	microstauron	0.06600			
Pinn obscura	PIOBSCUR	Chrysophyta	Pinnularia	nodosa				
Pinn rupestris	PISPP	Chrysophyta	Pinnularia	obscura				
Pinn spp.	PISUBCAP	Chrysophyta	Pinnularia	rupestris				
Pinn subcapitata	PISUBSTO	Chrysophyta	Pinnularia	spp.	0.01000			
Pinn substomatophora	PISUDETTI	Chrysophyta	Pinnularia	subcapitata				
Pinn sudetica	Pitermit	Chrysophyta	Pinnularia	substomatophora	0.00400			
Pinn temnitina	PIVIRIDI	Chrysophyta	Pinnularia	sudetica				
Pinn viridis	PN1UNK	Chrysophyta	Pinnularia	termitina				
Unkn penn 1	PN2UNK	Chrysophyta	Pinnularia	vindis				
Unkn penn 2	PN3UNK	Chrysophyta	Pinnularia	Unknown Pennate	0.00200			
Unkn penn 3	PN5UNK	Chrysophyta	Pinnularia	Unknown Pennate				
Unkn penn 5	PNSPP	Chrysophyta	Unknown Pennate	sp. 1				
Unkn pennate spp.	RHeriens	Chrysophyta	Unknown Pennate	sp. 2				
Rhiz eriensis	ROCURVAT	Chrysophyta	Unknown Pennate	sp. 3				
Rhoi curvata	RPGIBBER	Chrysophyta	Unknown Pennate	sp. 5				
Rhop gibberula	SNINTERM	Chrysophyta	Rhizosolenia	sp. 6	0.00600			
Sten intermedia	SSANCEPS	Chrysophyta	Rhoicosphenia	eriensis				
Stau anceps	SSANCGRA	Chrysophyta	Rhopalodia	curvata				
Stau anceps gracilis	SSKRIEGE	Chrysophyta	Stenopterobia	gibberula				
Stau kriegeri	SSliving	Chrysophyta	Stauroneis	intermedia	0.00800			
Stau livingstonii	SSPHOENI	Chrysophyta	Stauroneis	anceps				
Stau phoenicenteron	SSPHOGRA	Chrysophyta	Stauroneis	anceps				
Stau phoeni gracilis	SSSMINC	Chrysophyta	Stauroneis	kriegeri				
Stau smithii incisa	SSSPP	Chrysophyta	Stauroneis	livingstonii				
Stau spp.	SSTrunca	Chrysophyta	Stauroneis	phoenicenteron				
Stau truncata	STHANTZS	Chrysophyta	Stauroneis	smithii				
Step hantzschii	STMEDIUS	Chrysophyta	Stephanodiscus	spp.	0.00200			
Step medius	STMINUTU	Chrysophyta	Stephanodiscus	truncata				
Step minutus			Stephanodiscus	hantzschii				
			Stephanodiscus	medius				
			Stephanodiscus	minutus				

Diatom Counts

Long name	TAXANAME	Division	Genus	Species	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE
Achn sp. A Cascades	AC0ARSCS	Chrysophyta	Achnanthes	sp. PRS 0A	0.00400			
Achn sp. 1?	AC1?	Chrysophyta	Achnanthes	sp. 1?				
Achn sp. 11	AC11PRL	Chrysophyta	Achnanthes	sp. 11 PIRLA				
Achn sp. 12	AC12PRL	Chrysophyta	Achnanthes	sp. 12 PIRLA				
Step nigraeae	STNIAGAR	Chrysophyta	Stephanodiscus	niagarae				
Step parvus	STPARVUS	Chrysophyta	Stephanodiscus	parvus	0.06800	0.01833		
Suri sp. A Cascades	SU0Arscs	Chrysophyta	Suriella	sp. PRS 0A				
Suri sp. B Cascades	SU0Brscs	Chrysophyta	Suriella	sp. PRS 0A				
Suri sp. 1?	SU1?	Chrysophyta	Suriella	sp. 1?				
Suri sp. 3	SU3PRL	Chrysophyta	Suriella	sp. 3 PIRLA				
Suri sp. 6	SU6PRL	Chrysophyta	Suriella	sp. 6 PIRLA				
Suri angusta	SUANGUST	Chrysophyta	Suriella	angusta				
Suri delicatis var 1	SUde1PRL	Chrysophyta	Suriella	delicatissima				
Suri delicatissima	SUDELICA	Chrysophyta	Suriella	delicatissima				
Suri linear constrict	SULINCON	Chrysophyta	Suriella	linearis				
Suri linearis	SULINEAR	Chrysophyta	Suriella	linearis				
Suri splendida	SUSPLEND	Chrysophyta	Suriella	splendida				
Suri spp.	SUSPP	Chrysophyta	Suriella	spp.				
Syne sp. 1	SY1PRL	Chrysophyta	Synedra	sp. 1 PIRLA				
Syne sp. 8	SY8PRL	Chrysophyta	Synedra	sp. 8 PIRLA				
Syne cyclopum	SYCYCLOP	Chrysophyta	Synedra	cyclopum				
Syne delicatissima	SYDELICA	Chrysophyta	Synedra	delicatissima				
Syne filiform exilis	SYFILEXI	Chrysophyta	Synedra	filiformis				
Syne parasitica	SYparasi	Chrysophyta	Synedra	parasitica				
Syne ulna	SYULNA	Chrysophyta	Synedra	ulna				
Tabe binalis	TABINALI	Chrysophyta	Tabellaria	binalis				
Tabe fenestrata	TAFENEST	Chrysophyta	Tabellaria	fenestrata				
Tabe flocculosa	TAFLOCCU	Chrysophyta	Tabellaria	flocculosa				
Tabe flocculosa str4	TAflost4	Chrysophyta	Tabellaria	flocculosa				
	UN1?GOK	(Undetermined)	(Undetermined)	sp. 1? ANS GOK				
	UN2?GOK	(Undetermined)	(Undetermined)	sp. 2? ANS GOK				

Diatom Counts

Diatom Counts

	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE	SOUTH HEAVENLY Twin	LITTLE TWIN LAKE	HEAD LAKE	SUMMIT LAKE	CORA LAKE
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11							0.01800				
Achn sp. 12							0.00400				
Achn minutissima							0.01400				
Achn nodosa							0.00234				
Achn pusilla											
Achn spp.											
Achn subatomoides											
Achn suchlandii											
Amph sp. A Cascades											
Amph libyca											
Amph ovalis							0.00600				
Amph ovali pediculus											
Amph perpusilla							0.00429				
Anom brachysira							0.00350				
Anom serians											
Anom vitrea											
Aste formosa							0.00200				
Aste ralfsii											
Aula sp. A Cascades											
Aula sp. B Cascades											
Aula sp. C Cascades											
Aula sp. D Cascades											
Aula ambigua							0.01800				
Aulo distans humilis											
Aulo distans nivalis							0.00200				
Aulo dist nivaloides											
Aula distans							0.03600				
Aulo distans tenella											
Aula granulata											
Aula italicica							0.00467				
Aulo italicica subarct											
Aulo italic tenu											
Aulo litata											
Aula perglabra											
Aula paffiana											

Diatom Counts

	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE	SOUTH HEAVENLY Twin	LITTLE TWIN LAKE	HEAD LAKE	SUMMIT LAKE	CORA LAKE
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											
Achn sp. 12											
Aula species											
Aula validia											
Calo bacillum											
Calo hyalina											
Calo silicula											
Calo tenuis											
Cocc placentula											
Cocc placen euglypta											
Chae sp. 1?											
Cymb sp. A Cascades											
Cymb sp. B Cascades											
Cymb sp. 1 ?											
Cymb sp. 18											
Cymb sp. 2 ?											
Cymb sp. 20											
Cymb sp. 21											
Cymb sp. 6											
Cymb sp. 7											
Cymb aequalis											
Cymb amphicephala											
Cymb brehmii											
Cymb caespitosa											
Cymb cesatii											
Cymb cistula											
Cymb descripta											
Cymb elginensis											
Cymb falaisensis											
Cymb gaeumannii	0.02800										
Cymb hauckii											
Cymb hebridica											
Cymb heter subrostra											
Cymb imaequalis											
Cymb lunata											
	0.00600										
	0.01752										
	0.00200	0.01752									
	0.00467										
	0.01800	0.00467									
	0.00117										
	0.02000										
	0.00667										
	0.01000										
	0.02800										
	0.01000										
	0.00400										
	0.00400										

Diatom Counts

Long name	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE	SOUTH HEAVENLY Twin	LITTLE TWIN LAKE	HEAD LAKE	SUMMIT LAKE	CORA LAKE
Achn sp. A Cascades											
Achn sp. 1?											
Achn sp. 11											
Achn sp. 12											
Cymb mesiana											
Cymb mexicana											
Cymb microcephala											
Cymb minutula latens											
Cymb minut silesiaca											
Cymb minutula											
Cymb muelleri											
Cymb naviculiformis											
Cymb perpusilla											
Cymb sinuata											
Cymb spp.											
Cyma solea											
Cycl sp. 1											
Cycl bod lemanica											
Cycl kuetzin radiosa											
Cycl ocellata											
Cycl pseudostelliger											
Cycl stelligera											
Diat anceps											
Diat hemicale											
Diat mesodon											
Diat tenuis											
Dipl sp. 1?											
Dipl elliptica											
Dipl finnica											
Dipl marginestrata											
Dipl modica											
Epit adnata											
Epit sonex											
Epit turgida											
Epit turgida gracil											
Euno sp. 1?											
Euno sp. 42											

Diatom Counts

	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE	SOUTH HEAVENLY Twin	LITTLE TWIN LAKE	HEAD LAKE	HEAD LAKE	SUMMIT LAKE	CORA LAKE
Long name												
Achn sp. A Cascades												
Achn sp. 1 ?												
Achn sp. 11												
Achn sp. 12												
Euno sp. 44												
Euno sp. 45												
Euno sp. 46												
Euno arcus							0.00234					
Euno bidentula var 1												
Euno bidentula												
Euno bilun mucophila												
Euno bilunaris												
Euno curvata												
Euno denticulata												
Euno exigua												
Euno flexuosa												
Euno hemicyclus												
Euno implicata												
Euno incisa var 1												
Euno incisa var 2												
Euno incisa var 6												
Euno incisa												
Euno intermedia												
Euno meisteri												
Euno microcephala												
Euno micr tridentata												
Euno minor												
Euno monodon												
Euno musci trident												
Euno naegelii												
Euno palud trinac												
Euno paludososa												
Euno pectinali minor												
Euno pectinalis												
Euno praerupta												
Euno quaternaria												
Euno rhomboidea												

Diatom Counts

Long name	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE	SOUTH HEAVENLY Twin	LITTLE TWIN LAKE	HEAD LAKE	LAKE SUMMIT	CORA LAKE
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11							0.00117				
Achn sp. 12							0.00234				
Euno serra diadema											
Euno serra											
Euno spp.	0.00400										
Euno subarcuatooides											
Euno sudetica											
Euno tenella											
Euno trinacria											
Euno vanheurckii											
Euno sp. 74							0.00111				
Frag sp. A Cascades											
Frag sp. B Cascades											
Frag sp. 11											
Frag sp. 15											
Frag sp. 17											
Frag sp. 9											
Frag arcus											
Frag bidens	0.00222										
Frag brevistriata	0.21556										
Frag capu rumpens											
Frag capucina											
Frag capu vaucheriae	0.00286										
Frag constru binodis	0.00429										
Frag construe pumila											
Frag construens	0.00143										
Frag constru subsalii	0.00222										
Frag construe venter	0.02222										
Frag crotensis	0.11286										
Frag constricta											
Frag exigua							0.01000				
Frag fasciculata											
Frag lapponica								0.01000			
Frag leptostau dubia	0.00286										
Frag oldenburgiana											

Diatom Counts

	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE	SOUTH HEAVENLY Twin	LITTLE TWIN LAKE	HEAD LAKE	LAKE SUMMIT	CORA LAKE
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											0.00200
Achn sp. 12											0.04800
Frag parasitica											0.00400
Frag pinn lancettula											
Frag pinnata	0.04556	0.00857					0.02200				
Frag pseudoconstruen	0.01778	0.00429					0.16200				
Frag spp.											
Frag vaucheriae											
Frag virescens											
Frag vires exi											
Frus rhom amphipleur											
Frus rhomboides CAF											
Frus rhom crassiner	0.01600										
Frus rhomboides	0.00600										
Frus rhombo saxonica											
Gomp sp. A Cascades											
Gomp sp. 1 ?											
Gomp sp. 11											
Gomp sp. 18											
Gomp sp. 1											
Gomp angustum											
Gomp angustatum											
Gomp consector											
Gomp gracile											
Gomp grovei lingulat											
Gomp olivcm mintsnum											
Gomp parvulum											
Gomp puig aequatoria											
Gomp spp.											
Gomp subcl mexicanum											
Gomp tack brevistria											
Gomp turris											
Ham arcus											
Meri circulate											

Diatom Counts

Long name	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE	SOUTH HEAVENLY TWIN	LITTLE TWIN LAKE	HEAD LAKE	SUMMIT LAKE	CORA LAKE
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											
Achn sp. 12											
Melo sp. 1 ?											
Melo sp. 13											
Melo sp. 4											
Melo sp. 8											
Navi sp. 1 Mt. Rain.											
Navi sp. A Cascades											
Navi sp. B Cascades											
Navi sp. C Cascades											
Navi sp. D Cascades											
Navi sp. E Cascades											
Navi sp. 1 ?											
Navi sp. 14											
Navi sp. 1											
Navi sp. 2 ?											
Navi sp. 20											
Navi sp. 21											
Navi sp. 24											
Navi sp. 25											
Navi sp. 3 ?											
Navi sp. 45											
Navi sp. 47											
Navi sp. 51											
Navi sp. 52											
Navi sp. 6											
Navi sp. 74											
Navi sp. 91											
Navi aboensis											
Nav absoluta											
Navi acceptata											
Navi accommoda											
Navi angusta											
Navi arvensis											
Navi bacillum											

Diatom Counts

	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE	SOUTH HEAVENLY Twin	LITTLE TWIN LAKE	HEAD LAKE	LAKE SUMMIT	CORA LAKE
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											
Achn sp. 12											
Navi bremensis											
Navi bryophila											
Navi cincta											
Navi coccineiformis											
Navi cohnii											
Navi concentrica											
Navi crucicula											
Navi cryptocephala											
Navi cryptotenella											
Navi disjuncta											
Navi elginensis											
Navi gallica											
Navi harderii											
Navi jaernefeldii											
Navi laevissima											
Navi leptostriata											
Navi libonensis											
Navi medioconvexa											
Navi mediocris											
Navi mediopunctata											
Navi menisculus											
Navi minima											
Navi minuscula v. A											
Navi modica											
Navi mutica											
Navi pelliculosa											
Navi placenta											
Nav porifera											
Navi pseudomuralis											
Navi pseudoscutiform											
Navi pseudoventralis											
Navi pupul elliptica											
Navi pup rectangular											

Diatom Counts

Long name	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE	SOUTH HEAVENLY Twin	LITTLE TWIN LAKE	HEAD LAKE	SUMMIT LAKE	CORA LAKE
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											
Achn sp. 12											
Navi pupula											
Navi radiosa											
Navi radiosa parva											
Navi rhynchocephala											
Navi seminuloides											
Navi seminulum											
Navi spp.											
Navi cf subtil var 2											
Navi cf subtil var 4											
Navi cf subtil var 5											
Navi subatomoides											
Navi subhyalina											
Navi submuralis											
Navi subplacentula											
Navi subrotundata											
Navi subtilissima											
Navi tenuicephala											
Navi tridentula											
Navi trivialis											
Nav veneta											
Navi viridula											
Navi vitabunda											
Neid sp. 13											
Neid sp. 4											
Neid affine											
Neid affin longiceps											
Neid alpinum											
Neid alp quadripunct											
Neid bisulcata											
Neid hercyn f subros											
Neid iridi ampliatum											
Neid ir amphigomphus											

Diatom Counts

	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE	SOUTH HEAVENLY Twin	LITTLE TWIN LAKE	HEAD LAKE	SUMMIT LAKE	CORA LAKE
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											
Achn sp. 12											
Neid iridis											
Neid spp.											
Nitz sp. A Cascades											
Nitz sp. 1 ?											
Nitz sp. 12											
Nitz sp. 14											
Nitz sp. 15											
Nitz sp. 16											
Nitz sp. 17											
Nitz sp. 26											
Nitz sp. 34											
Nitz sp. 35											
Nitz sp. 36											
Nitz sp. 37											
Nitz sp. 38											
Nitz alpina											
Nitz amphibia											
Nitz bacillariaeform											
Nitz brizophila											
Nitz capitellata											
Nitz dissipata											
Nitz dissip undulata											
Nitz fonticola											
Nitz fossilis											
Nitz frustulum var 3											
Nitz frustulum var 8											
Nitz gracilis											
Nitz gracilliformis											
Nitz inconspicua											
Nitz lacuum											
Nitz leistikowii											
Nitz modesta											
Nitz palea debilis											

Diatom Counts

Diatom Counts

	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE	SOUTH HEAVENLY Twin	LITTLE TWIN LAKE	HEAD LAKE	LAKE SUMMIT	CORA LAKE
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											0.00200
Achn sp. 12											
Pinn major											
Pinn microstauron	0.01800				0.04400	0.01285		0.07600	0.10000	0.02400	0.00400
Pinn nodosa											0.00400
Pinn obscura											
Pinn rupestris											
Pinn spp.	0.00800						0.01400	0.01000		0.00400	0.00200
Pinn subcapitata	0.00400										
Pinn substromatophora							0.00600				
Pinn sudetica											
Pinn terminalia											
Pinn viridis											
Unkn penn 1											
Unkn penn 2											
Unkn penn 3											
Unkn penn 5											
Unkn pennate spp.											
Rhiz eriensis											
Rhoi curvata							0.00143				
Rhop gibberula											
Sten intermedia											
Stau anceps											
Stau anceps gracilis											
Stau kriegeri											
Stau livingstonii											
Stau phoenicenteron											
Stau phoeni gracilis											
Stau smithii incisa											
Stau spp.											
Stau truncata											
Step hantzschii											
Step medius											
Step minutus											

Diatom Counts

	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE	SOUTH HEAVENLY Twin	LITTLE TWIN LAKE	HEAD LAKE	LAKE SUMMIT	CORA LAKE
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											
Achn sp. 12											
Step niagarae											
Step parvus											
Suri sp. A Cascades											
Suri sp. B Cascades											
Suri sp. 1 ?											
Suri sp. 3											
Suri sp. 6											
Suri angusta											
Suri delicatis var 1											
Suri delicatissima											
Suri linear constrict											
Suri linearis											
Suri splendida											
Suri spp.											
Syne sp. 1											
Syne sp. 8											
Syne cyclopum											
Syne delicatissima											
Syne filiform exilis											
Syne parasitica											
Syne una											
Tabe binialis											
Tabe fenestrata											
Tabe flocculosa											
Tabe flocculosa str4											
	0.03333		0.01429								
		0.00400		0.00467							
			0.00111		0.00200		0.00200				
				0.00117		0.00200					
					0.00117						
						0.00200					
							0.00200				
								0.00200			
									0.00200		
										0.00200	
											0.00200

Diatom Counts

	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM LAKE	MARION LAKE
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Achn sp. 13										
Achn sp. 14										
Achn sp. 16										
Achn sp. 19										
Achn sp 2 Mt. Rain.										
Achn acares										
Achn altaica										
Achn austriaca										
Achn bioreti										
Achn calcar										
Achn chlidanos										
Achn clevei										
Achn curtissima										
Achn daui alaskaensi										
Achn detha										
Achn didyma										
Achn exigua										
Achn flexella										
Achn grana										
Achn grisichuna										
Achn helvetica										
Achn holstii										
Achn hungarica										
Achn kuelbsii										
Achn lanceolata										
Achn lanceolat dubia										
Achn lan ssp. frequ										
Achn laterostrata										
Achn levanderi										
Achn levan helvetica										
Achn linearis curta										
Achn linearis										
Achn marginulata										

Diatom Counts

	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM LAKE	MARION LAKE
Long name										0.01000
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Achn minutissima										
Achn nodosa										
Achn pusilla										
Achn spp.										
Achn subatomoides										
Achn suchlandii										
Amph sp. A Cascades										
Amph libyca										
Amph ovalis										
Amph ovali pediculus										
Amph perpusilla										
Anom brachysira										
Anom serians										
Anom vitrea										
Aste formosa										
Aste ralfsii										
Aula sp. A Cascades										
Aula sp. B Cascades										
Aula sp. C Cascades										
Aula sp. D Cascades										
Aula ambiguua										
Aulo distans humilis										
Aulo distans nivalis										
Aulo dist nivaloides										
Aula distans										
Aulo distans tenella										
Aula granulata										
Aula italicica										
Aulo italicica subarct										
Aulo italic tenu										
Aulo lirata										
Aula perglabra										
Aula paffiana										

Diatom Counts

	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM	MARION LAKE
Long name										
Achn sp. A Cascades				0.16071						
Achn sp. 1 ?										0.01000
Achn sp. 11										
Achn sp. 12										
Aula species										
Aula validula	0.02000									
Calo bacillum										0.00800
Calo hyalina										
Calo silicula										0.00400
Calo tenuis										0.00400
Cocc placenta										
Cocc placen euglypta										
Chae sp. 1?										0.00167
Cymb sp. A Cascades										
Cymb sp. B Cascades										
Cymb sp. 1 ?										
Cymb sp. 18										
Cymb sp. 2 ?										
Cymb sp. 20										
Cymb sp. 21										
Cymb sp. 6										
Cymb sp. 7										
Cymb aequalis										
Cymb amphicephala										
Cymb brehmii										
Cymb caespitosa										
Cymb cesatii										
Cymb cistula										
Cymb descripta										
Cymb elginensis										
Cymb falaisensis	0.00400									
Cymb elginiensis										
Cymb gaeumannii	0.01200									0.03200
Cymb hauckii										
Cymb hebridica	0.01200									0.00400
Cymb heter subrostra										
Cymb inaequalis										
Cymb lunata	0.01800	0.00600	0.00400	0.00992	0.01600	0.05000	0.00800	0.00400	0.00400	

Diatom Counts

	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM	MARION LAKE
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Cymb mesiana										
Cymb mexicana										
Cymb microcephala										
Cymb minuta latens										
Cymb minut sillesiaca										
Cymb minuta	0.01000	0.00800				0.00595				
Cymb muelleri										
Cymb naviculiformis										
Cymb perpusilla										
Cymb sinuata										
Cymb spp.	0.00600	0.00200				0.00200	0.00400	0.00400		
Cyma solea										
Cycl sp. 1										
Cycl bod lemanica										
Cycl kuetzin radiosa										
Cycl ocellata										
Cycl pseudostelliger										
Cycl stelligera										
Diat anceps										
Diat hiemale										
Diat mesodon										
Diat tenuis										
Dipl sp. 1 ?										
Dipl elliptica										
Dipl finnica										
Dipl marginestriata										
Dipl modica										
Epit adnata										
Epit sorex										
Epit turgida										
Epit turgida gracil										
Euno sp. 1 ?										
Euno sp. 42										

Diatom Counts

	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM LAKE	MARION LAKE
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Euno sp. 44										
Euno sp. 45										
Euno sp. 46										
Euno arcus										
Euno bidentula var 1										
Euno bidentula										
Euno bilun mucophila										
Euno bilunaris										
Euno curvata										
Euno denticulata										
Euno exigua										
Euno flexuosa										
Euno hemicyclus										
Euno implicata										
Euno incisa var 1										
Euno incisa var 2										
Euno incisa var 6										
Euno incisa										
Euno intermedia										
Euno meisteri										
Euno microcephala										
Euno micr tridentata										
Euno minor										
Euno monodon										
Euno musci trident										
Euno naegelii										
Euno palud trinac										
Euno paludososa										
Euno pectinali minor										
Euno pectinalis										
Euno praerupta										
Euno quaternaria										
Euno rhomboidea										

Diatom Counts

Diatom Counts

	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM	MARION LAKE
Long name										0.01000
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Frag parasitica										
Frag pinn lancettula	0.00600									
Frag pinnata	0.00400									
Frag pseudoconstruen	0.11000									
Frag spp.	0.06800									
Frag vaucheriae										
Frag virescens										
Frag vires exi										
Frus rhom amphipleur										
Frus rhombooides CAF	0.00200									
Frus rhom crassinerv	0.00200									
Frus rhombooides	0.00200									
Frus rhombo saxonica										
Gomp sp. A Cascades	0.00200									
Gomp sp. 1 ?										
Gomp sp. 11										
Gomp sp. 18										
Gomp sp. 1										
Gomp angustum										
Gomp angustatum										
Gomp consector										
Gomp gracile	0.00200									
Gomp grovel lingulat										
Gomp olivcm minstium										
Gomp olivaceum										
Gomp parvulum										
Gomp puig aequatoria										
Gomp spp.										
Gomp subcl mexicanum										
Gomp tack brevistria										
Gomp turris										
Hann arcus										
Meri circulare	0.03968									

Diatom Counts

	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM	MARION LAKE
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Melo sp. 1 ?										
Melo sp. 13										
Melo sp. 4										
Melo sp. 8										
Navi sp. 1 Mt. Rain.										
Navi sp. A Cascades										
Navi sp. B Cascades										
Navi sp. C Cascades										
Navi sp. D Cascades										
Navi sp. E Cascades										
Navi sp. 1 ?										
Navi sp. 14										
Navi sp. 1										
Navi sp. 2 ?										
Navi sp. 20										
Navi sp. 21										
Navi sp. 24										
Navi sp. 25										
Navi sp. 3 ?										
Navi sp. 45										
Navi sp. 47										
Navi sp. 51										
Navi sp. 52										
Navi sp. 6										
Navi sp. 74										
Navi sp. 91										
Navi aboensis										
Nav absoluta										
Navi acceptata										
Navi accomoda										
Navi angusta										
Navi arvensis										
Navi bacillum										
				0.16071	0.00400	0.000200	0.00200	0.000200	0.000200	0.01000

Diatom Counts

	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM LAKE	MARION LAKE
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Navi bremensis										0.00200
Navi bryophila										
Navi cincta										
Navi cocconeiformis										
Navi cohnii										
Navi concentrica										
Navi crucicula										
Navi cryptocephala										
Navi cryptotenia										
Navi disjuncta										
Navi elginensis										
Navi gallica										
Navi harderii										
Navi jaernefeldti										
Navi laevissima	0.00200									
Navi leptostriata	0.00400									
Navi libonensis										
Navi mediocconvexa										
Navi mediocris										
Navi mediopunctata										
Navi menisculus										
Navi minima										
Navi minuscula v. A										
Navi modica										
Navi mutica										
Navi pelliculosa										
Navi placenta										
Nav porifera										
Navi pseudomurialis										
Navi pseudoscutiform										
Navi pseudoventralis										
Navi pup elliptica										
Navi pup rectangular										
										0.00200

Diatom Counts

	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM	MARION LAKE
Long name										
Achn sp. A Cascades				0.16071						
Achn sp. 1?										0.01000
Achn sp. 11										
Achn sp. 12										
Navi pupula	0.00200	0.01400								
Navi radiosa	0.00200	0.00200								
Navi radiosa parva	0.00200	0.00200								
Navi rhynchocephala					0.00400					0.00167
Navi seminuloides					0.05400					
Navi seminulum	0.01600	0.02600			0.00600					
Navi spp.					0.00400					
Navi cf subtil var 2										0.00500
Navi cf subtil var 4										
Navi cf subtil var 5	0.02200	0.01400			0.00198					
Navi subatomoides						0.00400				
Navi subhyalina						0.00600				
Navi submuralis	0.02400	0.00800			0.00800					
Navi subplacentula						0.00200				
Navi subrotundata						0.00200				
Navi subtilissima						0.05600				
Navi tenuephala						0.00992				
Navi tridentula	0.00400	0.04000			0.04800					
Navi trivialis										
Nav veneta										
Navi viridula										
Navi vitabunda	0.01000									
Neid sp. 13										
Neid sp. 4										
Neid affine										
Neid affin longiceps										
Neid alpinum										
Neid ap quadripunct										
Neid bisulcata										
Neid bisulcatum										
Neid hecycn f subros										
Neid iridi ampliatum	0.00200									
Neid ir amphigomphus										

Diatom Counts

Diatom Counts

	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM LAKE	MARION LAKE
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										0.00333
Achn sp. 12										0.00167
Nitz palea					0.00600	0.00200	0.00200	0.00600		
Nitz paleaeformis					0.00600	0.00200				
Nitz perminuta					0.00600	0.00600				
Nitz recta					0.00200					
Nitz spp.					0.00400					
Nitz tropica										
Nitz vermicularis					0.00800					
Opep sp. 1										0.00800
Opep martyi										
Opep olsenii					0.01200					
Pero fibula										0.00198
Pinn sp. A Cascades										
Pinn sp. B Cascades										
Pinn sp. C Cascades										
Pinn sp. D Cascades										
Pinn sp. 24										
Pinn sp. 25										
Pinn sp. 28										
Pinn sp. 2 PIRLA					0.02600					
Pinn sp. 30										
Pinn sp. 32										
Pinn sp. 36										
Pinn sp. 46										
Pinn abaljensis var 2										
Pinn abaljensis										
Pinn biceps										
Pinn borealis rectan										
Pinn braunii										
Pinn brebissonii										
Pinn divergentissima										
Pinn gibba										
Pinn mesogong										
Pinn hemiptera										

Diatom Counts

	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM LAKE	MARION LAKE
Long name										0.01000
Achn sp. A Cascades										
Achn sp. 1 ?				0.16071						
Achn sp. 11										
Achn sp. 12										
Pinn major				0.00200	0.00198					
Pinn microbrisss										
Pinn microstauron										
Pinn nodosa										
Pinn obscura										
Pinn rupestris										
Pinn spp.				0.00400						
Pinn subcapitata										
Pinn substiomatophora										
Pinn sudetica										
Pinn terminalia										
Pinn viridis										
Unkn penn 1										
Unkn penn 2										
Unkn penn 3										
Unkn penn 5										
Unkn pennate spp.										
Rhiz eriensis										
Rhoi curvata										
Rhop gibberula										
Sten intermedia	0.00400				0.00198					
Stau anceps	0.00200	0.00200				0.00200	0.00200	0.00600		0.00600
Stau anceps gracilis	0.01000	0.00400				0.00800	0.03400			0.01400
Stau kriegeri										
Stau livingstonii										
Stau phoenicapteron										
Stau phoeni gracilis										
Stau smithii incisa										
Stau spp.										
Stau truncata										
Step hantzschii										0.01167
Step mediis										0.18667
Step minutus										0.02500

Diatom Counts

Diatom Counts

	SUTTLE LAKE	Upper Palisades Lake	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9
Long name										
Achn sp. A Cascades										
Achn sp. 1?										
Achn sp. 11										
Achn sp. 12										
Achn sp. 13										
Achn sp. 14										
Achn sp. 16										
Achn sp. 19										
Achn sp 2 Mt. Rain.										
Achn acares										
Achn altaica										
Achn austriaca										
Achn bioreti										
Achn calcar										
Achn chlidanos										
Achn clevei										
Achn curtissima										
Achn daui alaskaensi										
Achn detha										
Achn didyma										
Achn exigua										
Achn flexella										
Achn grana										
Achn grischuna										
Achn helvetica										
Achn holstii										
Achn hungarica										
Achn kuelbsii										
Achn lanceolata										
Achn lanceolat dubia										
Achn lan ssp. frequ										
Achn laterostrata										
Achn levanderi										
Achn levan helvetica										
Achn linearis curta										
Achn linearis										
Achn marginulata										
	0.09449	0.09195	0.19737	0.08119	0.07059	0.05167	0.03901	0.01822	0.06958	0.01992

Diatom Counts

Diatom Counts

	SUTTLE LAKE	Upper Palisades Lake	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Aula species										
Aula valida										
Calo bacillum										
Calo hyalina										
Calo silicula										
Calo tenuis										
Cocc placenta										
Cocc placen euglypta										
Chae sp. 1?										
Cymb sp. A Cascades										
Cymb sp. B Cascades										
Cymb sp. 1 ?										
Cymb sp. 18										
Cymb sp. 2 ?										
Cymb sp. 20										
Cymb sp. 21										
Cymb sp. 6										
Cymb sp. 7										
Cymb aequalis	0.00984	0.01149	0.01504	0.06139	0.05490	0.00667	0.06738	0.05688	0.03976	0.01594
Cymb amphicephala										
Cymb brehmii										
Cymb caespitosa										
Cymb cesatii										
Cymb cistula										
Cymb descripta										
Cymb elginensis										
Cymb falaiensis	0.05512	0.07663	0.01316	0.01782	0.02157	0.06000	0.01950	0.00810	0.01193	0.01793
Cymb gaeumannii										
Cymb hauckii										
Cymb hebridica	0.00197									
Cymb heter subrostra										
Cymb inaequalis										
Cymb lunata										
										0.02191

Diatom Counts

	SUTTLE LAKE	Upper Palisades Lake	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9
Long name										
Achn sp. A Cascades										
Achn sp. 1?										
Achn sp. 11										
Achn sp. 12										
Cymb mesiana										
Cymb mexicana										
Cymb microcephala										
Cymb minuta latens										
Cymb minut silesiaca										
Cymb minuta										
Cymb muelleri										
Cymb naviculiformis										
Cymb perpusilla										
Cymb sinuata										
Cymb spp.										
Cyma solea										
Cycl sp. 1										
Cycl bid lemancia										
Cycl kuetzin radiosa										
Cycl ocellata										
Cycl pseudostelliger										
Cycl stelligera										
Diat anceps										
Diat hemale										
Diat mesodon										
Diat tenuis										
Dipl sp. 1?										
Dipl elliptica										
Dipl finnica										
Dipl marginestriata										
Dipl modica										
Epit adnata										
Epit sorex										
Epit turgida										
Euno sp. 1?										
Euno sp. 42										

Diatom Counts

Diatom Counts

	SUTTLE LAKE	Upper Palisades Lake	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Euno serra diadema										
Euno serra										
Euno spp.										
Euno subarcuatooides										
Euno sudetica										
Euno tenella										
Euno trinacia										
Euno vanheurckii										
Euno sp. 74										
Frag sp. A Cascades										
Frag sp. B Cascades										
Frag sp. 11										
Frag sp. 15										
Frag sp. 17										
Frag sp. 9										
Frag arcus										
Frag bidens										
Frag brevistriata										
Frag capu rumpens										
Frag capucina										
Frag capu vaucheriae										
Frag constru binodis										
Frag construe pumila										
Frag construens										
Frag constru subsali										
Frag construe venter										
Frag crotonensis										
Frag constricta										
Frag exigua										
Frag fasciculata										
Frag lapponica										
Frag leptostau dubia										
Frag oldenburgiana										

Diatom Counts

	Long name	SUTTLE LAKE	Upper Pallisades Lake	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9
Achn sp. A Cascades												
Achn sp. 1 ?												
Achn sp. 11												
Achn sp. 12												
Frag parasitica												
Frag pinn lanceolata												
Frag pinnata	0.15200											
Frag pseudoconstruens												
Frag spp.												
Frag vaucheriae												
Frag virescens												
Frag vires exilis												
Frus rhomb amphipleur												
Frus rhomboides CAF												
Frus rhom crassinerv	0.01181				0.00383							
Frus rhomboides	0.00197					0.01128						
Frus rhombo saxonica												
Gomp sp. A Cascades												
Gomp sp. 1 ?												
Gomp sp. 11												
Gomp sp. 18												
Gomp sp. 1												
Gomp angustum												
Gomp angustatum												
Gomp consector												
Gomp gracile												
Gomp grovei lingulata												
Gomp olivaceum minstimum												
Gomp olivaceum												
Gomp parvulum												
Gomp puig aequatoria												
Gomp spp.												
Gomp subcl mexicanum												
Gomp tach brevistria												
Hann arcus												
Meri circulare												

Diatom Counts

	SUTTLE LAKE	Upper Palisades Lake	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Melo sp. 1 ?										
Melo sp. 13										
Melo sp. 4										
Melo sp. 8										
Navi sp. 1 Mt. Rain.										
Navi sp. A Cascades										
Navi sp. B Cascades										
Navi sp. C Cascades										
Navi sp. D Cascades										
Navi sp. E Cascades										
Navi sp. 1 ?										
Navi sp. 14										
Navi sp. 1										
Navi sp. 2 ?										
Navi sp. 20										
Navi sp. 21										
Navi sp. 24	0.05315	0.01724	0.02632	0.00792	0.00196	0.01500	0.11170	0.00607	0.01193	
Navi sp. 25										
Navi sp. 3 ?										
Navi sp. 45										
Navi sp. 47										
Navi sp. 51										
Navi sp. 52										
Navi sp. 6										
Navi sp. 74										
Navi sp. 91										
Navi aboensis										
Nav absoluta										
Navi acceptata										
Navi accomoda										
Navi angusta										
Navi arvensis										
Navi bacillum										

Diatom Counts

	SUTTLE LAKE	Upper Palisades Lake	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Navi bremensis										
Navi bryophila										
Navi cincta										
Navi coccineiformis										
Navi cohni										
Navi concentrica										
Navi crucicula										
Navi cryptocephala										
Navi cryptotenella										
Navi disjuncta										
Navi elginensis										
Navi gallica										
Navi harderii										
Navi jaernefeldti										
Navi laevissima										
Navi leptostriata										
Navi ibonensis										
Navi medioconvexa										
Navi mediopunctata										
Navi menisculus										
Navi minima										
Navi minuscula v. A										
Navi modica										
Navi mutica										
Navi pelliculosa										
Navi placenta										
Nav porifera										
Navi pseudomurialis										
Navi pseudoscutiform										
Navi pseudoventralis										
Navi pup elliptica										
Navi pup rectangular										

Diatom Counts

	SUTTLE LAKE	Upper Palisades Lake	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											
Achn sp. 12											
Navi pupula	0.00200										
Navi radiosa											
Navi radiosa parva											
Navi rhynchocephala											
Navi seminuloides	0.01200										
Navi seminulum											
Navi spp.		0.00196									
Navi cf subtil var 2											
Navi cf subtil var 4											
Navi cf subtil var 5											
Navi subatomoides											
Navi subhyalina											
Navi submuralis	0.00800										
Navi subplacentula											
Navi subrotundata											
Navi subtilissima											
Navi tenuicephala											
Navi tridentula											
Navi trivialis											
Nav veneta			0.00400								
Navi viridula											
Navi vitabunda											
Neid sp. 13											
Neid sp. 4											
Neid affine											
Neid affin longiceps											
Neid alpinum											
Neid alp quadripunct											
Neid bisulcata											
Neid hercyn f subros											
Neid iridi ampliatum											
Neid ir amphigomphus											
	0.00394	0.00383	0.00376	0.00594	0.00167	0.01241	0.00202	0.00598	0.00795		

Diatom Counts

	Upper Palisades Lake	SUTTLE LAKE	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Neid iridis										
Neid spp.										
Nitz sp. A Cascades										
Nitz sp. 1 ?										
Nitz sp. 12										
Nitz sp. 14										
Nitz sp. 15										
Nitz sp. 16										
Nitz sp. 17										
Nitz sp. 26										
Nitz sp. 34										
Nitz sp. 35										
Nitz sp. 36										
Nitz sp. 37										
Nitz sp. 38										
Nitz alpina										
Nitz amphibia										
Nitz bacillariaeform										
Nitz bruophila										
Nitz capitellata										
Nitz dissipata										
Nitz dissip undulata										
Nitz fonticola										
Nitz fossilis										
Nitz frustulum var 3										
Nitz frustulum var 8										
Nitz gracilis										
Nitz graciliformis										
Nitz inconspicua										
Nitz lacuum										
Nitz leistikowii										
Nitz modesta										
Nitz palea debilis										
										0.00398
										0.00028
										0.00199
										0.00996

Diatom Counts

	Upper Palisades Lake	SUTTLE LAKE	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Nitz palea										
Nitz paleaformis	0.00200									
Nitz perminta										
Nitz recta										
Nitz spp.										
Nitz tropica										
Nitz vermicularis										
Opep sp. 1										
Opep martyi										
Opep olsenii										
Pero fibula										
Pinn sp. A Cascades										
Pinn sp. B Cascades										
Pinn sp. C Cascades										
Pinn sp. D Cascades										
Pinn sp. 24										
Pinn sp. 25										
Pinn sp. 28										
Pinn sp. 2 PIRLA										
Pinn sp. 30										
Pinn sp. 32										
Pinn sp. 36										
Pinn sp. 46										
Pinn abaujensi var 2										
Pinn abaujensis										
Pinn biceps										
Pinn borealis rectan										
Pinn braunii										
Pinn brebissonii										
Pinn divergentissima										
Pinn gibba										
Pinn gibba mesogong										
Pinn hemiptera										
0.00197	0.07283	0.001316	0.02970	0.00980	0.02167	0.13830	0.02632	0.03777	0.02590	0.00405
0.00591		0.00192	0.00376							
0.00199										
0.00199										

Diatom Counts

	Upper Palisades Lake	SUTTLE LAKE	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											
Achn sp. 12											
Pinn maior											
Pinn micros brevis											
Pinn microstauron	0.00400	0.00984		0.00192	0.02820	0.00198	0.01176	0.00167	0.01950	0.02024	0.00795
Pinn nodosa											0.00398
Pinn obscura											
Pinn rupestris											
Pinn spp.											
Pinn subcapitata											
Pinn substomatophora											
Pinn sudetica											
Pinn terminalia											
Pinn viridis											
Unkn penn 1											
Unkn penn 2											
Unkn penn 3											
Unkn penn 5											
Unkn pennate spp.											
Rhiz eriensis											
Rhoi curvata											
Rhop gibberula											
Sten intermedia											
Stau anceps											
Stau anceps gracilis											
Stau kriegeri											
Stau livingstonii											
Stau phoenicenteron											
Stau phoeni gracilis											
Stau smithii incisa											
Stau spp.											
Stau truncata											
Step hantzschii	0.00400										
Step medius	0.02200										
Step minutus	0.02200										

Diatom Counts

	Upper Palisades Lake	SUTTLE LAKE	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9
Long name										
Achn sp. A Cascades										
Achn sp. 1 ?										
Achn sp. 11										
Achn sp. 12										
Step niagareae										
Step parvus										
Suri sp. A Cascades										
Suri sp. B Cascades										
Suri sp. 1 ?										
Suri sp. 3										
Suri sp. 6										
Suri angusta										
Suri delicatis var 1										
Suri delicatissima										
Suri linear constrict										
Suri linearis										
Suri splendida										
Suri spp.										
Syne sp. 1										
Syne sp. 8										
Syne cyclopum										
Syne delicatissima										
Syne filiform exilis										
Syne parasitica										
Syne ulna										
Tabe binialis										
Tabe fenestrata										
Tabe flocculosa										
Tabe flocculosa str4										

Diatom Counts

	Unnamed Lake10	Unnamed Lake6	Green Lake	Mowich Lake	Lake Notasha	Reflection Lake B	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11					0.00155	0.02727	0.01127	0.00930	0.00733	0.16934	
Achn sp. 12					0.10248						
Achn sp. 13					0.00932						
Achn sp. 14											
Achn sp. 16											
Achn sp. 19											
Achn sp 2 Mt. Rain.											
Achn acares											
Achn altaica											
Achn austriaca											
Achn bioreti											
Achn calcar											
Achn chlidanos											
Achn clevei											
Achn curtissima											
Achn daui alaskaensi											
Achn detha											
Achn didyma											
Achn exigua											
Achn flexella											
Achn grana											
Achn grischuna											
Achn helvetica											
Achn holstii											
Achn hungarica											
Achn kuelbsii											
Achn lanceolata											
Achn lanceolat dubia											
Achn lan ssp. frequ											
Achn laterostrata											
Achn levanderi											
Achn levan helvetica											
Achn linearis curta											
Achn linearis											
Achn marginulata	0.00980	0.05545	0.00151	0.00932	0.02351	0.05000	0.16264	0.01240	0.08791	0.05255	0.28155

Diatom Counts

Diatom Counts

	Unnamed Lake 10	Unnamed Lake 6	Green Lake	Mowich Lake	Lake Notasha	Reflection Lake B	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											
Achn sp. 12											
Aula species											
Aula validia											
Calo bacillum											
Calo hyalina											
Calo silicula											
Calo tenuis											
Cocc placentula											
Cocc placen euglypta											
Chae sp. 1?											
Cymb sp. A Cascades	0.00784	0.05743									
Cymb sp. B Cascades											
Cymb sp. 1 ?											
Cymb sp. 18											
Cymb sp. 2 ?											
Cymb sp. 20											
Cymb sp. 21											
Cymb sp. 6											
Cymb sp. 7											
Cymb aequalis	0.01373										
Cymb amphicephala	0.00196										
Cymb brehmii											
Cymb caespitosa											
Cymb cesatii											
Cymb cistula											
Cymb descripta											
Cymb elginensis											
Cymb fataisensis	0.00980	0.02178									
Cymb gaeumannii											
Cymb hauckii	0.00392	0.00990									
Cymb hebridica											
Cymb heter subrostra											
Cymb inaequalis	0.00392										
Cymb lunata											

Diatom Counts

	Long name	Unnamed Lake 10	Unnamed Lake 6	Green Lake	Mowich Lake Notasha	Lake	Reflection Lake B	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
Achn sp. A Cascades						0.00155						
Achn sp. 1 ?						0.10248						
Achn sp. 11						0.00932						
Achn sp. 12												
Cymb mesiana												
Cymb mexicana												
Cymb microcephala												
Cymb minut latens												
Cymb minut silesiaca												
Cymb minuta												
Cymb muelleri												
Cymb naviculiformis												
Cymb perpusilla						0.00196						
Cymb sinuata												
Cymb spp.												
Cyma solea												
Cycl sp. 1												
Cycl bod lemanica												
Cycl kuetzin radiosa												
Cycl ocellata												
Cycl pseudostelliger												
Cycl stelligera												
Diat anceps												
Diat hemale												
Diat mesodon												
Diat tenuis												
Dipl sp. 1 ?												
Dipl elliptica												
Dipl finnica												
Dipl marginestriata												
Dipl modica												
Epit adnata												
Epit sorex												
Epit turgida												
Epit turgida gracil												
Euno sp. 1 ?												
Euno sp. 42												

Diatom Counts

Long name	Unnamed Lake10	Unnamed Lake6	Green Lake	Mowich Lake	Lake Notasha	Reflection Lake B	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											
Achn sp. 12											
Euno sp. 44											
Euno sp. 45											
Euno sp. 46											
Euno arcus											
Euno bidentula var 1											
Euno bidentula											
Euno bilun mucophila											
Euno bilunaris	0.00196										
Euno curvata											
Euno denticulata											
Euno exigua	0.00392	0.00396									
Euno flexuosa											
Euno hemicyclus											
Euno implicata	0.00392										
Euno incisa var 1											
Euno incisa var 2											
Euno incisa var 6											
Euno incisa											
Euno intermedia											
Euno meisteri											
Euno microcephala											
Euno micr tridentata											
Euno minor											
Euno monodon											
Euno musci trident											
Euno naegelii											
Euno palud trinac	0.00784	0.00594									
Euno paludosa											
Euno pectinali minor											
Euno pectinalis											
Euno praerupta											
Euno quaternaria											
Euno rhomboidea											

Diatom Counts

Long name	Unnamed Lake10	Unnamed Lake6	Green Lake	Mowich Lake Notasha	Lake	Reflection Lake B	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
Achn sp. A Cascades					0.00155						
Achn sp. 1 ?					0.10248						
Achn sp. 11					0.00932						
Achn sp. 12					0.00151						
Euno serra diadema											
Euno serra											
Euno spp.											
Euno subarctooides					0.00198						
Euno sudetica											
Euno tenella					0.00151	0.00311					
Euno trinacria											
Euno vanheurckii											
Euno sp. 74											
Frag sp. A Cascades											
Frag sp. B Cascades											
Frag sp. 11											
Frag sp. 15											
Frag sp. 17											
Frag sp. 9											
Frag arcus											
Frag bidens											
Frag brevistriata											
Frag capu rumpens											
Frag capucina											
Frag capu vaucheriae											
Frag constru binodis											
Frag construe pumila											
Frag construens											
Frag constru subalii											
Frag construe venter											
Frag crotensis											
Frag constricta											
Frag exigua											
Frag fasciculata											
Frag laponica											
Frag leptostau dubia											
Frag oldenburgiana											

Diatom Counts

	Unnamed Lake10	Unnamed Lake6	Green Lake	Mowich Lake	Lake Notasha	Reflection Lake B	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
Long name											
Achn sp. A Cascades					0.00155						
Achn sp. 1 ?					0.10248						
Achn sp. 11					0.00932						
Achn sp. 12											
Frag parasitica											
Frag pinn lancettula											
Frag pinnata											
Frag pseudoconstruen											
Frag spp.											
Frag vaucheriae											
Frag virescens											
Frag vires exi											
Frus rhom amphileur											
Frus rhomboides CAF											
Frus rhom crassimerv											
Frus rhomboides											
Frus rhombo saxonica											
Gomp sp. A Cascades											
Gomp sp. 1 ?											
Gomp sp. 11											
Gomp sp. 18											
Gomp sp. 1											
Gomp angustum											
Gomp angustatum											
Gomp consector											
Gomp gracile											
Gomp grovei lingulat											
Gomp olivcm mintnum											
Gomp olivaceum											
Gomp parvulum											
Gomp puig aequatoria											
Gomp spp.											
Gomp subcl mexicanum											
Gomp tack brevistria											
Hann arcus											
Meri circulare											

Diatom Counts

	Unnamed Lake10	Unnamed Lake6	Green Lake	Mowich Lake	Lake Notasha	Reflection Lake B	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11	0.00155										
Achn sp. 12	0.10248										
Melo sp. 1 ?	0.00932										
Melo sp. 13											
Melo sp. 4	0.00303										
Melo sp. 8	0.00455										
Navi sp. 1 Mt. Rain.	0.08939										
Navi sp. A Cascades	0.06924										
Navi sp. B Cascades	0.04848										
Navi sp. C Cascades	0.22383										
Navi sp. D Cascades											
Navi sp. E Cascades	0.00155										
Navi sp. 1 ?	0.00155										
Navi sp. 14	0.00155										
Navi sp. 1											
Navi sp. 2 ?	0.00152										
Navi sp. 20	0.00152										
Navi sp. 21	0.00152										
Navi sp. 24	0.00396										
Navi sp. 25	0.00151										
Navi sp. 3 ?	0.00621										
Navi sp. 45	0.00155										
Navi sp. 47											
Navi sp. 51	0.00303										
Navi sp. 52	0.00303										
Navi sp. 6											
Navi sp. 74											
Navi sp. 91											
Navi abeoensis											
Nav absoluta											
Navi acceptata											
Navi accomoda											
Navi angusta											
Navi arvensis											
Navi bacillum											
	0.00152										
	0.01127										
	0.00549										
	0.00583										

Diatom Counts

	Unnamed Lake 10	Unnamed Lake 6	Green Lake	Mowich Lake	Lake Notasha	Reflection Lake B	Shaddow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											
Achn sp. 12											
Navi bremensis											
Navi bryophila											
Navi cincta											
Navi coccineiformis											
Navi cohnii											
Navi concentrica											
Navi crucicula											
Navi cryptocephala											
Navi cryptotenella											
Navi disjuncta											
Navi elginiensis											
Navi gallica											
Navi harderii											
Navi jaemefeldti											
Navi laevissima											
Navi leptostriata											
Navi libonensis											
Navi medioconvexa											
Navi mediocris											
Navi mediopunctata											
Navi menisculus											
Navi minima											
Navi minuscula v. A											
Navi modica											
Navi mutica											
Navi pelliculosa											
Navi placenta											
Nav porifera											
Navi pseudomuralis											
Navi pseudoscutiform											
Navi pseudoventralis											
Navi pup elliptica											
Navi pup rectangular											

Diatom Counts

Diatom Counts

	Long name	Unnamed Lake 10	Unnamed Lake 6	Green Lake	Mowich Lake Notasha	Lake	Reflection Lake B	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
Achn sp. A Cascades						0.00155						
Achn sp. 1 ?						0.10248						
Achn sp. 11						0.00932						
Achn sp. 12							0.00362					
Neid iridis		0.00196										
Neid spp.												
Nitz sp. A Cascades												
Nitz sp. 1 ?												
Nitz sp. 12						0.00776						
Nitz sp. 14						0.00155						
Nitz sp. 15						0.00311						
Nitz sp. 16						0.00621						
Nitz sp. 17							0.00151					
Nitz sp. 26							0.00466					
Nitz sp. 34								0.01395				
Nitz sp. 35								0.00155				
Nitz sp. 36								0.14729				
Nitz sp. 37								0.00310				
Nitz sp. 38									0.00152			
Nitz alpina										0.00146		
Nitz amphibia											0.00438	
Nitz bacillariaeform												
Nitz bruophila												
Nitz capitellata												
Nitz dissipata						0.01553						
Nitz dissip undulata						0.00311						
Nitz fonticola												
Nitz fossilis												
Nitz frustulum var 3												
Nitz frustulum var 8												
Nitz gracilis												
Nitz graciliformis												
Nitz inconspicua												
Nitz lacuum												
Nitz leistikowii												
Nitz modesta												
Nitz palea debilis												

Diatom Counts

	Unnamed Lake 10	Unnamed Lake 6	Green Lake	Mowich Lake	Lake Notasha	Reflection Lake B	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
Long name											
Achn sp. A Cascades											
Achn sp. 1 ?											
Achn sp. 11											
Achn sp. 12											
Nitz palea											
Nitz paleaeformis											
Nitz perminuta											
Nitz recta											
Nitz spp.											
Nitz tropica											
Nitz vermicularis											
Opep sp. 1											
Opep martyi											
Opep Olsenii											
Pero fibula											
Pinn sp. A Cascades											
Pinn sp. B Cascades											
Pinn sp. C Cascades											
Pinn sp. D Cascades											
Pinn sp. 24											
Pinn sp. 25											
Pinn sp. 28											
Pinn sp. 2 PIRLA											
Pinn sp. 30											
Pinn sp. 32											
Pinn sp. 36											
Pinn sp. 46											
Pinn abaujensi var 2											
Pinn abaujensis											
Pinn biceps											
Pinn borealis rectan											
Pinn braunii											
Pinn brebissonii											
Pinn divergentissima											
Pinn gibba											
Pinn gibba mesogong											
Pinn hemiptera											

Diatom Counts

	Long name	Unnamed Lake10	Unnamed Lake6	Green Lake	Mowich Lake	Lake Notasha	Reflection Lake B	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
Achn sp. A Cascades												
Achn sp. 1 ?												
Achn sp. 11												
Achn sp. 12												
Pinn major												
Pinn micros brevis												
Pinn microstauron												
Pinn nodosa												
Pinn obscura												
Pinn rupestris												
Pinn spp.												
Pinn subcapitata												
Pinn substomatophora												
Pinn sudetica												
Pinn terminalia												
Pinn viridis												
Unkn penn 1												
Unkn penn 2												
Unkn penn 3												
Unkn penn 5												
Unkn pennate spp.												
Rhiz eriensis												
Rhoi curvata												
Rhop gibberula												
Sten intermedia												
Stau anceps												
Stau anceps gracilis												
Stau kriegeri												
Stau livingstonii												
Stau phoenicenteron												
Stau phoeni gracilis												
Stau smithii incisa												
Stau spp.												
Stau truncata												
Step hantzschii												
Step medius												
Step minutus												

Diatom Counts

APPENDIX 2

Compressed Diatom Taxa List Used in Ordination and Modeling for the Cascade Diatom Calibration Set

TAXANAME	Long name	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY
CMBREHMI	<i>Cymb bremii</i>										
CMFALALIS	<i>Cymb falaisensis</i>										
CMGAEUMA	<i>Cymb gaeumannii</i>										
CMHEBRID	<i>Cymb hebridica</i>										
CMLUNATA	<i>Cymb lunata</i>										
CMMINUTA	<i>Cymb minuta</i>										
CMPPERPUS	<i>Cymb perpusilla</i>										
CYOCELLA	<i>Cycl ocellata</i>										
CYPSESTE	<i>Cycl pseudostelliger</i>										
CYSTELLI	<i>Cycl stelligera</i>	0.11200									
EUBILMUC	<i>Euno bilunaria</i>										
EUBLILUNA	<i>Euno exigua</i>										
EUEXIGUA	<i>Euno incisa vars</i>										
EUINCIVS	<i>Euno minor</i>										
EUMINOR	<i>Euno palud trinac</i>										
EUPALTRI	<i>Euno rhomboidea</i>										
EURHOMBO	<i>Euno subarcuatooides</i>										
EUSUBARC	<i>Euno vanheurck vars</i>										
EUVANHVs	<i>Frag sp. A Cascades</i>										
FR0AstsCS	<i>Frag sp. 1'</i>										
FR11PRL	<i>Frag sp. 15</i>										
FR15PRL	<i>Frag sp. 9</i>										
FR9PRL	<i>Frag brevistriata</i>	0.18000	0.07200	0.01000							
FRBREVIS	<i>Frag capucina vars</i>										
FRCAPUvs	<i>Frag crotoneensis</i>										
FRCRDTON	<i>Frag construens vars</i>	0.27800	0.17000	0.09500							
FRCSRNvs	<i>Frag exigua & viresc</i>										
FREXI&VR	<i>Frag pinn lancettula</i>	0.13400	0.13600	0.43000	0.00200						
FRPINLAN	<i>Frag pinnata</i>	0.12600	0.18000	0.32500							
FRPINNAT	<i>Frag pseudoconstruens</i>	0.01400									
FRPSEC0N	<i>Frus rhom crassinerv</i>										
FSRHOCRA	<i>Frus rhomboides</i>										
FSRHOMBO											

TAXA NAME	Long name	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE
GOangust	Gomp angustatum										0.00400
GOGRACIL	Gomp gracie										0.00701
ME13PRL	Melo sp. 13										
ME4PRL	Melo sp. 4										
ME8PRL	Melo sp. 8										
NA0ErsCS	Navi sp. E Cascades										
NA20PRL	Navi sp. 20										
NA24PRL	Navi sp. 24										
NA25PRL	Navi sp. 25										
NA45PRL	Navi sp. 45										
NACRYPT0	Navi cryptocephalia	0.00400	0.00200	0.01400	0.00083	0.00100	0.00222				
NACRYTEN	Navi cryptotenella	0.00200	0.01000	0.01000	0.00167	0.00167					
NALAEVIS	Navi laevissima										0.00200
NALEPTOS	Navi leptostriata										0.00200
NAMEDIOC	Navi mediocris										
NAPUPUvs	Navi pupula vars										
NARADIVs	Navi radiosa vars										
NASEMDES	Navi seminuloides	0.00600	0.00200	0.00400	0.00167	0.00167	0.00143				
NASEMLUM	Navi seminulum	0.00200	0.01000	0.00400	0.00200	0.00200	0.00111	0.00143			
NASUBATO	Navi subatomoides	0.00200	0.02000	0.01000	0.00200	0.00111	0.00111	0.00200			0.00600
NASUBMUR	Navi submuralis										
NASUBTvs	Navi subtil vars										
NATENUJC	Navi tenuicephala	0.01000									
NEAFFVs	Neid affine vars										
NEAL&BIV	Neid alp & basic vars										
NEIRIDVs	Neid iridis vars										
NI36PRL	Nitz sp. 36										
NIFONTIC	Nitz fonticola										
NIff3PRL	Nitz frustulum var 3	0.02200	0.00167								
NIGRACIL	Nitz gracilis										
NIPALEVs	Nitz palea vars	0.00400									
NIPERMIN	Nitz permunita										
NITROPIC	Nitz tropica	0.00417									

TAXANAME	Long name	BURNT LAKE	DIAMOND LAKE	LAVA LAKE	MINK LAKE	BIG LAKE	PAULINA	EAST LAKE	LUCKY LAKE	CHARLTON LAKE	AGENCY LAKE
P10Cscs	Pinn sp. C Cascades										
PI28PRL	Pinn sp. 28										
PIBICEPS	Pinn biceps										
PIBRAUNI	Pinn braunii										
Pibrebis	Pinn brebissonii										
PIGIBVs	Pinn gibba vars										
PIMICRvs	Pinn micros vars										
PISUBCAP	Pinn subcapitata										
SNINTERM	Sten intermedia										
SSANCEvs	Stau anceps vars										
STHANTZS	Step hantzschii										
STMEDIUS	Step medius	0.00400									
STMINUTU	Step minutus	0.06200	0.02667								
STNIAGAR	Step niagare	0.06800	0.01833								
STPARVUS	Step parvus	0.02583									
SUde1PRL	Suri delicatis var 1										
SUDELICA	Suri delicatissima										
TAspp	Tabe spp.										

	SOUTH	HEAVENLY	LITTLE	SUMMIT	CORA LAKE	GERTRUDE	CASKEY	MIDDLE	PYRAMID	HEATHER
	Twin	TWIN LAKE	HEAD LAKE	LAKE	CORA LAKE	LAKE	NO NAME	THORNTON	LAKE	LAKE
Long name										
Achn sp. A Cascades										0.00400
Achn sp. 11										
Achn bioreti										0.00200
Achn curtissima										
Achn detha										
Achn lanceolata vars										
Achn linearis vars										
Achn margin & levand	0.05200	0.00778	0.09400	0.02000	0.01000	0.02600	0.01600	0.00198	0.03800	0.04000
Achn minutissima										
Achn pusilla										
Achn subatomoides										
Achn suchlandii										
Amph ovalis										
Amph perpusilla										
Anom brachysira	0.01600	0.00222	0.00200		0.00200	0.00800	0.00200	0.00600	0.0198	0.01200
Anom vitrea										
Aste formosa										
Aula sp. A Cascades										
Aula sp. B Cascades	0.00600									
Aula sp. C Cascades										
Aula sp. D Cascades										
Aula ambigua										
Aula distans vars	0.02000	0.03444	0.15600	0.00800	0.00600	0.03000	0.09000	0.03800	0.01000	0.01400
Aula granulata										
Aula italica vars										
Aula paffiana										
Cocc placent vars										
Cymb sp. A Cascades	0.04000									
Cymb sp. 18										
Cymb sp. 20										
Cymb sp. 6	0.13000									
Cymb aequalis	0.01600	0.05889	0.02200							
Cymb amphicephala										

	SOUTH Twin	HEAVENLY TWIN LAKE	LITTLE TWIN LAKE	HEAD LAKE	SUMMIT LAKE	CORA LAKE	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE
Long name												0.00800
<i>Cymb brehmii</i>												
<i>Cymb falaisensis</i>												
<i>Cymb gaeumannii</i>												0.00400
<i>Cymb hebridica</i>	0.02000	0.00667	0.01000	0.02800	0.01000	0.00400	0.00400	0.00400	0.01200	0.01200	0.01200	
<i>Cymb lunata</i>												0.01600
<i>Cymb minuta</i>												0.01000
<i>Cymb perpusilla</i>												0.00595
<i>Cycl ocellata</i>												0.06400
<i>Cycl pseudostelliger</i>												
<i>Cycl stelligera</i>												
<i>Euno bilun mucophila</i>	0.00444	0.00600	0.01200	0.09600	0.02800	0.02800	0.02800	0.02800	0.14600	0.38800	0.00200	
<i>Euno bilunaris</i>												0.01200
<i>Euno exigua</i>												0.00400
<i>Euno incisa vars</i>												0.00200
<i>Euno minor</i>												0.00800
<i>Euno palud trinac</i>												
<i>Euno rhomboidea</i>												
<i>Euno subarcuatooides</i>	0.00333	0.00200	0.01200	0.00400	0.00200	0.00200	0.00200	0.00200	0.03571	0.03571	0.00200	
<i>Euno vanheurck vars</i>												
<i>Frag sp. A Cascades</i>												
<i>Frag sp. 11</i>												
<i>Frag sp. 15</i>												
<i>Frag sp. 9</i>												
<i>Frag brevistriata</i>												
<i>Frag capucina vars</i>												
<i>Frag crotonensis</i>												
<i>Frag construens vars</i>												
<i>Frag exigua & viresc</i>												
<i>Frag pinn lanceifolia</i>												
<i>Frag pinnata</i>												
<i>Frag pseudoconstruens</i>												
<i>Frus rhom crassinerv</i>	0.02000	0.00667	0.00400	0.04800	0.00400	0.00200	0.00200	0.00200	0.04960	0.04960	0.00800	
<i>Frus rhomboides</i>												0.00600

	SOUTH Twin	HEAVENLY TWIN LAKE	LITTLE HEAD LAKE	SUMMIT LAKE	CORA LAKE LAKE	GERTRUDE LAKE	CASKEY LAKE	NO NAME	MIDDLE THORNTON LAKE	PYRAMID LAKE	HEATHER LAKE
Long name											
Gomp angustatum	0.00200	0.00222			0.00400	0.00200	0.00200		0.00200	0.00400	
Gomp gracile	0.00200	0.00222			0.00400	0.00200	0.00200		0.00200	0.00400	
Melo sp. 13											
Melo sp. 4											
Melo sp. 8											
Navi sp. E Cascades					0.03800	0.00800	0.00800		0.00600	0.00400	
Navi sp. 20					0.01800	0.00200	0.00200		0.00200	0.00200	
Navi sp. 24					0.01800	0.00200	0.00200		0.00200	0.00200	
Navi sp. 25					0.01800	0.00200	0.00200		0.00200	0.00200	
Navi sp. 45					0.00400	0.00400	0.00400		0.00400	0.00400	
Navi cryptocephala					0.02400	0.00200	0.00200		0.00200	0.00200	
Navi cryptotenia					0.01600	0.05400	0.00400	0.01800	0.02600	0.01400	
Navi laevissima					0.00800	0.01600	0.00400	0.01600	0.02200	0.00794	0.00400
Navi leptostriata					0.00200	0.00200	0.01800	0.01400	0.02000	0.01400	0.00200
Navi mediociris											
Navi pupula vars											
Navi radiosa vars											
Navi seminuloides											
Navi seminulum											
Navi subatomoides											
Navi submuralis											
Navi subtil vars	0.06400	0.01333	0.00600	0.04600	0.00600	0.01200	0.04000	0.00800	0.00200	0.00198	0.00400
Navi tenuiceps	0.39400	0.00333	0.35000	0.19800	0.00600	0.01200	0.04800	0.00800	0.00200	0.00600	0.00600
Neid affine vars											
Neid alp & bslc vars											
Neid iridis vars	0.00800	0.02667	0.01000	0.02000	0.00200	0.00600	0.01000	0.00200	0.00794	0.00794	0.00400
Nitz sp. 36											
Nitz fonticola											
Nitz frustulum var 3											
Nitz gracilis	0.01200				0.00200	0.01000	0.00400	0.01000	0.00600	0.00397	0.02400
Nitz palea vars											
Nitz permunita											
Nitz tropica											

	SOUTH	HEAVENLY	LITTLE	TWIN LAKE	HEAD LAKE	LAKE	SUMMIT	CORA LAKE	GERTRUDE	CASKEY	LAKE	NO NAME	MIDDLE	THORNTON	PYRAMID	LAKE	HEATHER
Long name																	
Pinn sp. C Cascades																	
Pinn sp. 28																	
Pinn biceps	0.08800						0.00600	0.10200	0.01200	0.01000	0.02000	0.02000	0.08929				
Pinn braunii																	
Pinn brebissonii																	
Pinn gibba vars																	
Pinn micros vars																	
Pinn subcapitata																	
Sten intermedia																	
Stau anceps vars																	
Step hantzschii																	
Step medius																	
Step minutus																	
Step niagraeae																	
Step panvus																	
Suri delicatis var 1	0.00400						0.00400	0.03000					0.00400	0.00198			0.00400
Suri delicatissima	0.00400						0.02222	0.02400	0.00400				0.01000	0.00198			0.00200
Tabe spp.								0.00200		0.00600			0.00200				0.00400

Long name	DUMBBELL LAKE	SHELLROCK LAKE	K LAKE	SPHAGNUM NORDRUM LAKE	MARION LAKE	SUTTLE LAKE	Upper Palisades Lake	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake
Achn sp. A Cascades					0.01000						
Achn sp. 11											
Achn bioretii											
Achn curtissima	0.00400										
Achn dethia											
Achn lanceolata vars											
Achn linearis vars											
Achn margin & levand	0.05400	0.03800	0.00250	0.17200	0.00500	0.00167	0.05364	0.00376	0.00396	0.00392	0.00392
Achn minutissima											
Achn pusilla	0.00400										
Achn subatomoides											
Achn suchlandii											
Amph ovalis											
Amph perpusilla											
Anom brachysira	0.01600	0.02000	0.01375	0.04200	0.00333	0.00600	0.00333	0.00600	0.00198	0.00198	0.00198
Anom vitrea											
Aste formosa											
Aula sp. A Cascades	0.04600	0.03800	0.13500	0.04000	0.35500	0.09800	0.00197	0.00394	0.00940	0.00396	0.01176
Aula sp. B Cascades	0.04800	0.01600									
Aula sp. C Cascades											
Aula sp. D Cascades											
Aula ambigua											
Aula cistans vars	0.11600	0.17400	0.50000	0.05400	0.48228	0.26437	0.32519	0.22376	0.43529	0.02107	
Aula granulata	0.00200	0.00600		0.02800	0.05333	0.01600					
Aula italicica vars											
Aula paffiana	0.04600	0.11400	0.20000	0.03800	0.00167	0.00600					
Cocc placent vars											
Cymb sp. A Cascades	0.03400	0.06600	0.01625	0.00400	0.00984	0.01149	0.01504	0.006139	0.05490	0.05490	
Cymb sp. 18											
Cymb sp. 20											
Cymb sp. 6	0.01600	0.01400	0.02000	0.00600	0.00600	0.00400	0.01181	0.00376	0.00376	0.00376	
Cymb aequalis	0.03000	0.03000	0.00500								
Cymb amphicephala											

Long name	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM LAKE	MARION LAKE	SUTTLE LAKE	Upper Palisades Lake	Unnamed Lake						
Cymb brehmii	0.01000	0.00200	0.00250	0.00400									
Cymb falaensis					0.05512	0.07663	0.01316	0.00564	0.01386	0.01782	0.02157	0.00392	
Cymb gaeumannii	0.06600	0.03800	0.00500	0.03200									
Cymb hebridica	0.00400	0.03200	0.01375	0.00400								0.00594	
Cymb lunata	0.05000	0.00800		0.00400								0.01386	
Cymb minuta					0.00333								
Cymb perpusilla													
Cycl ocellata													
Cycl pseudostelliger													
Cycl stelligera					0.04000	0.00667	0.00400						0.00198
Euno bilun mucophila					0.03200								
Euno bilunaris					0.01800								0.00396
Euno exigua					0.01000	0.00250	0.01000						0.00198
Euno incisa vars					0.00125								0.00594
Euno minor													
Euno palud trinac													
Euno rhomboidea													
Euno subarciatoides													
Euno vanheurck vars													
Frag sp. A Cascades													
Frag sp. 11													
Frag sp. 15													
Frag sp. 9													0.00188
Frag brevistriata													
Frag capucina vars													
Frag crotonensis													
Frag construens vars													
Frag exigua & viresc													
Frag pinn lancettula													
Frag pinnata													
Frag pseudoconstruens													
Frus rhom crassirerv	0.00800	0.01600	0.00125	0.03000								0.00196	
Frus rhomboides	0.01600	0.06600	0.00750	0.02000								0.00196	

	DUMBBELL LAKE	SHELLROC K LAKE	SPHAGNUM NORDRUM LAKE	MARION LAKE	SUTTLE LAKE	Upper Paisades Lake	Unnamed Lake	Unnamed Lake3(Lost Lake)	Unnamed Lake4	Unnamed Lake
Long name										
Gomp angustatum										
Gomp gracile										
Melo sp. 13										
Melo sp. 4										
Melo sp. 8										
Navi sp. E Cascades										
Navi sp. 20										
Navi sp. 24	0.02200	0.00200								
Navi sp. 25										
Navi sp. 45										
Navi cryptocephala										
Navi cryptotenella	0.01000	0.00800								
Navi laevissima										
Navi leptostriata	0.00800	0.03000	0.01625	0.00600						
Navi mediocris	0.00200	0.02200	0.00500	0.02400						
Navi pupula vars										
Navi radiosa vars	0.01600			0.00400						
Navi seminuloides	0.00800	0.00800		0.00400						
Navi seminulum										
Navi subatomoides										
Navi submuralis	0.00400				0.00800					
Navi subtil vars	0.00400	0.00500	0.01800	0.00167						
Navi tenuicephala	0.01800	0.04600	0.00375	0.03800						
Neid affine vars	0.00400	0.01200	0.00250	0.01200						
Neid alp & bsic vars	0.01600	0.00400		0.00800						
Neid iris vars	0.00200		0.00125	0.00400						
Nitz sp. 36	0.01000	0.00400	0.00250		0.01500	0.00400				
Nitz fonticola										
Nitz frustulum var.3										
Nitz gracilis	0.01400	0.01400								
Nitz palea vars	0.02400	0.00200								
Nitz perminuta										
Nitz tropica										

Long name	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9	Unnamed Lake10	Unnamed Lake6	Green Lake	Mowich Lake	Lake Notasha	Reflection Lake B
Achn sp. A Cascades											
Achn sp. 11	0.01500										0.02727
Achn biorefi											
Achn curtissima											
Achn detha											
Achn lanceolata vars											
Achn linearis vars											
Achn margin & levand	0.05167	0.03901	0.01822	0.06958	0.01992	0.00980	0.04147	0.00151	0.05124	0.02351	0.00909
Achn minutissima							0.06452	0.00151	0.03177	0.01242	0.00606
Achn pusilla							0.003307	0.01059	0.08385	0.00152	0.00152
Achn subatomoides											
Achn suchlandti											
Amph ovalis											
Amph perpusilla											
Anom brachysira											
Anom vitrea											
Aste formosa											
Aula sp. A Cascades											
Aula sp. B Cascades	0.07833	0.00532	0.01822		0.11753	0.37647					
Aula sp. C Cascades					0.00398	0.00784					
Aula sp. D Cascades	0.01667		0.01215	0.04175	0.05976	0.07255					
Aula ambigua											
Aula distans vars	0.57667	0.12943	0.31579	0.48509	0.33466	0.19020	0.04916		0.02174	0.00362	0.15758
Aula granulata											
Aula italica vars											
Aula paffiana	0.01500				0.00994	0.06574	0.07255				
Cocc placent vars											
Cymb sp. A Cascades	0.00667	0.06738	0.05668		0.03976	0.01594	0.00784				
Cymb sp. 18											
Cymb sp. 20											
Cymb sp. 6											
Cymb aequalis											
Cymb amphicephala											

	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9	Unnamed Lake10	Unnamed Lake6	Green Lake	Mowich Lake	Lake Notasha	Reflection Lake B
Long name											
Gomp angustatum											0.00455
Gomp gracile											0.00303
Melo sp. 13											0.00455
Melo sp. 4											0.08939
Melo sp. 8											0.04848
Navi sp. E Cascades											
Navi sp. 20	0.01500	0.11170	0.00607	0.01193							
Navi sp. 24											0.00152
Navi sp. 25											0.00606
Navi sp. 45											0.00303
Navi cryptocephala											0.00303
Navi cryptotenella											0.01061
Navi taevissima											
Navi leptostriata	0.00833	0.00177	0.00202	0.00199	0.01392	0.02590	0.00588				
Navi medioris	0.00333										
Navi pupula vars											
Navi radiosa vars											
Navi seminuloides											
Navi seminulum											
Navi subatomoides											
Navi submuralis											
Navi subtil vars	0.00500	0.01418	0.01417	0.01193	0.00398						0.00909
Navi tenuicephala	0.07667	0.32447	0.43725	0.11730	0.06972	0.10000					0.36528
Neid affine vars											0.00606
Neid alp & bsic vars	0.01000				0.00795		0.00196				0.00303
Neid iridis vars	0.00167	0.01241	0.00202	0.00398	0.00598	0.00784					0.01667
Nitz sp. 36											
Nitz fonticola											
Nitz frustulum var 3											0.02727
Nitz gracilis											
Nitz palea vars											
Nitz permunita											
Nitz tropica											

Long name	Unnamed Lake5	Unnamed Lake7	Unnamed Lake8	Reflection Lake A	Unnamed Lake9	Unnamed Lake10	Unnamed Lake6	Green Lake	Mowich Lake Natasha	Lake	Reflection Lake B
Pinn sp. C Cascades											
Pinn sp. 28	0.02167	0.13830	0.02632	0.03777	0.02590	0.01373	0.02151		0.00155	0.01627	0.00606
Pinn biceps				0.00405							
Pinn braunii										0.05063	
Pinn brebissonii										0.06691	
Pinn gibba vars	0.00177	0.01950	0.02024	0.01193	0.00398	0.00588				0.00904	0.03333
Pinn micros vars											
Pinn subcapitata											
Sten intermedia											
Stau anceps vars	0.00532										
Step hantzschii											
Step medius											
Step minutus											
Step niagarae											
Step parvus											
Suri delicatis var 1	0.00810								0.00181		
Suri delicatissima	0.00887		0.02982	0.01992	0.00980	0.00154			0.00904	0.00152	0.00152
Tabe spp.											

	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	LAKE	SUMMIT
Long name						
Achn sp. A Cascades	0.01127	0.00930	0.00733	0.16934		
Achn sp. 11	0.00805			0.00438		
Achn bioreti						
Achn curtissima						
Achn detha		0.02171		0.04818		
Achn lanceolata vars						
Achn linearis vars						
Achn margin & levand						
Achn minutissima	0.16908	0.02016	0.08791	0.05255	0.28155	
Achn pusilla		0.01860		0.01314	0.01359	
Achn subatomoides		0.00620		0.00146		
Achn suchlandti						
Amph ovalis						
Amph perpusilla						
Anom brachysira	0.01449	0.00155	0.02198	0.00146	0.00388	
Anom vitrea			0.00366			
Aste formosa						
Aula sp. A Cascades						
Aula sp. B Cascades						
Aula sp. C Cascades						
Aula sp. D Cascades						
Aula ambigua						
Aula distans vars	0.07246	0.00155	0.05861	0.02482	0.20777	
Aula granulata						
Aula italicica vars	0.03221					
Aula paffiana						
Cocc placent vars						
Cymb sp. A Cascades						
Cymb sp. 18	0.01932		0.03480	0.00292		
Cymb sp. 20						
Cymb sp. 6						
Cymb aequalis	0.00155				0.08932	
Cymb amphicephala						

Long name	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
<i>Cymb brehmii</i>					
<i>Cymb falaisensis</i>					
<i>Cymb gaeumannii</i>	0.01288	0.00620	0.08608	0.00146	
<i>Cymb hebridica</i>					
<i>Cymb lunata</i>	0.00483	0.01550	0.05861		0.00194
<i>Cymb minuta</i>	0.01449	0.13023	0.01099	0.02190	
<i>Cymb perpusilla</i>					
<i>Cycl ocellata</i>					
<i>Cycl pseudostelliger</i>					
<i>Cycl stelligera</i>	0.00310				
<i>Euno bilun mucophila</i>					
<i>Euno bilunaris</i>		0.00183	0.00292		0.00194
<i>Euno exigua</i>					0.00583
<i>Euno incisa vars</i>			0.00292		0.04466
<i>Euno minor</i>	0.00161				
<i>Euno palud trinac</i>					0.04272
<i>Euno rhomboidea</i>				0.00146	
<i>Euno subarcuoides</i>					
<i>Euno vanheurck vars</i>					
<i>Frag sp. A Cascades</i>					
<i>Frag sp. 11</i>	0.02171			0.00146	
<i>Frag sp. 15</i>		0.01550		0.00146	
<i>Frag sp. 9</i>		0.13023			
<i>Frag brevistriata</i>					
<i>Frag capucina vars</i>					
<i>Frag crotoneensis</i>					
<i>Frag construens vars</i>					
<i>Frag exigua & viresc</i>					
<i>Frag pinn lancettula</i>					
<i>Frag pinnata</i>					
<i>Frag pseudoconstruens</i>					
<i>Frus rhom crassirerv</i>	0.00483	0.00155	0.01832		0.00388
<i>Frus rhomboides</i>		0.00465	0.01282	0.00146	

		Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	LAKE	SUMMIT
Long name		0.01449	0.01085	0.00366	0.00292		
Gomp angustatum							
Gomp gracile							
Melo sp. 13		0.01288				0.04380	
Melo sp. 4		0.06924	0.00155	0.05128	0.12117		
Melo sp. 8		0.22383	0.00155	0.09341	0.22482		
Navi sp. E Cascades							
Navi sp. 20				0.00366	0.01606		
Navi sp. 24							
Navi sp. 25		0.00322		0.00183	0.00876		
Navi sp. 45		0.02415		0.00366	0.00584		
Navi cryptocephala							
Navi cryptotenella		0.00966	0.01085	0.02381			
Navi laevissima				0.00549			
Navi leptostriata							
Navi medioris				0.01465			
Navi pupula vars		0.00161	0.00155		0.00438		
Navi radiosa vars				0.03101	0.00549		
Navi seminuloides							
Navi seminulum							
Navi subatomoides							
Navi submuralis							
Navi subtil vars		0.00161				0.14175	
Navi tenuicephala		0.00322		0.03114	0.00292		0.00194
Neid affine vars				0.03480			
Neid alp & bslc vars		0.00483				0.00146	
Neid iridis vars					0.00183		
Nitz sp. 36			0.14729				
Nitz fonticola							
Nitz frustulum var 3		0.00322	0.10388	0.02747			
Nitz gracilis							
Nitz palea vars							
Nitz permunita							
Nitz tropica							

Long name	Shadow Lake	Tipsoo Lake	Sunrise Lake	Snow Lake	SUMMIT LAKE
Pinn sp. C Cascades	0.14976		0.06410	0.02920	
Pinn sp. 28					0.00388
Pinn biceps					
Pinn braunii			0.01099		
Pinn brebissonii			0.00549	0.00292	
Pinn gibba vars					0.05243
Pinn micros vars	0.00322				
Pinn subcapitata					
Sten intermedia	0.00644	0.00155	0.00916		
Stau anceps vars	0.03060		0.00549	0.00584	
Step hantzschii					
Step medius					
Step minutus					
Step niagarae					
Step parvus					
Suri delicatis var 1					0.00194
Suri delicatissima	0.00161		0.00916		
Tabe spp.					